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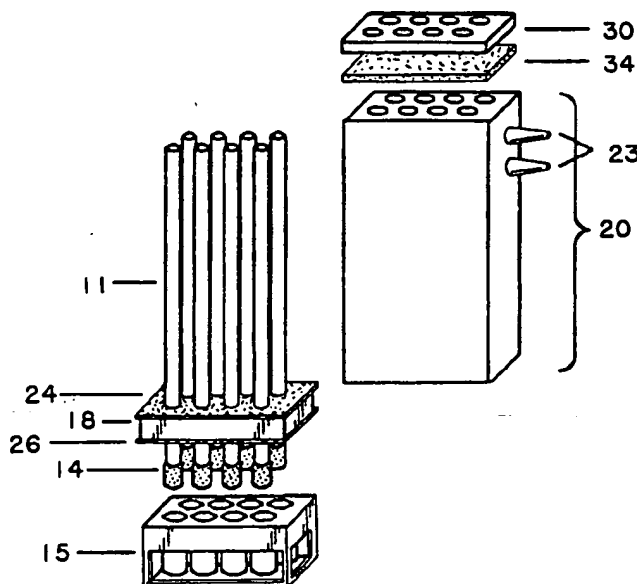


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(54) Title: APPARATUS AND METHOD FOR MULTIPLE SIMULTANEOUS SYNTHESIS**(57) Abstract**

An apparatus and method which provides a suitable location for multiple, simultaneous synthesis of compounds. The apparatus consists of: a reservoir block (15) having a plurality of wells (16); a plurality of reaction tubes (11), usually gas dispersion tubes, having filters (14) on their lower ends; a holder block (18), having a plurality of apertures (19); and a manifold (20), which may have ports (23) to allow introduction/maintenance of a controlled environment. The manifold top wall (22) has apertures (29) and a detachable plate (30) with identical apertures (31). The apparatus is constructed from materials which will accommodate heating, cooling, agitation, or corrosive reagents. Gaskets (24, 26, 34) are placed between the components. Rods (32) or clamps (35, 36, 37) are provided for fastening the components together. Apparatus operation involves placing the filters (14) on the lower ends of the reaction tubes (11) in the reservoir block wells (16), and the upper ends (12), passing through the holder block apertures (19) and into the manifold (20). The apparatus provides in excess of 1 mg of each product with structural knowledge and control over each compound. The apparatus can be adapted to manual, semiautomatic or fully automatic performance. Using the apparatus a series of building blocks are covalently attached to a solid support. These building blocks are then modified by covalently adding additional different building blocks or chemically modifying some existing functionality until the penultimate structure is achieved. This is then cleaved from the solid support by another chemical reaction into the solution within the well yielding an array of newly synthesized individual compounds, which after postreaction modification, if necessary, are suitable for testing for activity.



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APPARATUS AND METHOD FOR MULTIPLE
SIMULTANEOUS SYNTHESIS

5 BACKGROUND OF THE INVENTION

Field of the Invention

10 The invention relates to an apparatus and method which are used for the multiple, simultaneous synthesis of compounds, including but not limited to organic compounds.

Description of Related Art

15 It is well known in the art that peptides or oligonucleotides may be multiply and simultaneously synthesized. In a basic, single synthesis of a peptide, amino acids are simultaneously coupled to a functionalized solid support. Several methods have been developed by which peptides or oligonucleotides
20 may be multiply synthesized. One such methodology for peptide synthesis was disclosed in Geysen, et al, International Publication Number WO 90/09395. Geysen's method involves functionalizing the termini of polymeric rods and sequentially immersing the termini
25 in solutions of individual amino acids. Geysen's approach has proven to be impractical for commercial production of peptides since only very minute quantities of polypeptides may be generated. In addition, this method is extremely labor intensive. A
30 second method of peptide or oligonucleotide synthesis was developed by Affymax Technologies N.V. and disclosed in United States Patent Number 5,143,854. The Affymax method involves sequentially using light for illuminating a plurality of polymer sequences on a
35 substrate and delivering reaction fluids to said substrate. This method of synthesis has numerous

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drawbacks, including the fact that the products are noncleavable and that the process produces large numbers, but only minute quantities of products. A further method and device for producing peptides or oligonucleotides is disclosed in Houghton, European Patent Number 196174. Houghton's apparatus is a polypropylene mesh container, similar to a tea-bag, which encloses reactive particles. The containers, however, are not amenable to general organic synthesis techniques. Further apparatus are disclosed in German Published Patent Application Number DE 4005518 and European Patent Number 0355582, issued to Boehringer Ingelheim KG. Like the earlier devices, these apparatus are not suitable for the synthesis of general organic compounds and are directed toward peptide or oligonucleotide synthesis.

The synthesis of general organic compounds, poses many difficulties which are absent in the synthesis of peptides or oligonucleotides. An approach describing the synthesis of unnatural, oligomeric peptides is reported by Simon, et al, in Proceedings of the National Academy of Sciences USA 1992;89:9367. Accordingly, none of the disclosed devices or methods for the multiple, simultaneous synthesis of peptides or oligonucleotides are useful for the synthesis of general organic compounds. Among the many special problems found in the synthesis of general organic compounds, as opposed to peptide or oligonucleotide synthesis, is the problem of providing a device which will accommodate the wide range of synthetic manipulations required for organic synthesis. The synthesis of general organic compounds often requires such varied conditions as an inert atmosphere, heating, cooling, agitation, and an environment to facilitate reflux. Additionally, such synthesis requires chemical compatibility between the materials used in the

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apparatus for multiple synthesis and the reactants and solvents. Consequently, the apparatus must be constructed of materials which are resistant to organic synthesis conditions and techniques. Organic synthesis also often requires agitation. Such agitation may be accomplished by magnetic stirring, sonicating, or rotational shaking. None of the prior art devices are suitable for use under these special conditions required for general organic synthesis.

While undeniably useful, peptides or oligonucleotides have significant limitations in their application to pharmaceutical discovery programs. The chemical leads discovered from these collections of compounds require extensive modification due to the general unsuitability of peptides or nucleotides as stable, orally active drugs. The building blocks utilized are, in general, limited even allowing for the use of unnatural enantiomers or artificial amino acids and modified nucleotides. The peptides or oligonucleotides generated possess a repetitive linkage, amide or phosphate moiety, which limits their structural diversity.

The principal object of the present invention, therefore, is to overcome the limitations of the previous apparatus and methods which are limited to peptide or oligonucleotide synthesis and to provide an apparatus and method which will accommodate multiple, simultaneous synthesis of general organic compounds including, but not limited to, nonpeptide or nonnucleotide compounds.

SUMMARY OF THE INVENTION

The present invention is directed to an apparatus and method which will provide a suitable environment

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for the synthesis of organic compounds. Additionally, the synthesis of inorganic compounds, organometallic compounds, and the like is entirely within the scope of the invention.

5 Central to the demonstration of this concept is the need to devise a "general" method for multiple, simultaneous synthesis of organic compounds. The synthesis method developed must satisfy the following criteria. The compounds should be simultaneously
10 synthesized in an array format, which is compatible with the standard techniques of organic synthesis. The final compounds should be produced individually (not as mixtures) in soluble form. The quantity generated should be greater than 1 mg and in sufficiently pure
15 form to allow direct biological testing. Additionally, to the extent possible, sample handling should be carried out using automated systems for speed, accuracy, and precision. A final requirement is that the growing compounds must be readily separable from
20 by-products and reagents. Solid phase synthesis techniques commonly used in peptide or oligonucleotide synthesis enable achievement of this criteria. Typical solid supports (resins) include cross-linked divinylbenzene-styrene (polystyrene), controlled pore
25 glass (CPG), polyacrylamides, poly(ethylene glycol)-monomethyl ether and poly(ethylene glycol) (PEG), silica gel, cellulose, acrylic acid grafted polypropylene, and the like. Additionally, the solid support contains a reactive moiety. Thus, a
30 functionalized solid support is an insoluble material containing an accessible reactive moiety such as, for example, a carboxylic acid, alcohol, amine, halomethyl and the like which is used to covalently attach an incoming building block. A further objective of the
35 present invention is to synthesize products with

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molecular weights of less than, but not limited to, 1500 g/mol.

5 The approach described herein greatly increases the flexibility and diversity of structures that can be produced by a parallel, solid phase synthesis technology. Since neither the building blocks nor the methods for connecting them are in any way limited, the chemistries compatible with this apparatus and method are very broad, encompassing nearly all organic reactions. The key feature which allows for the success of this method is containment of a solid support within a gas dispersion tube. This feature provides a unique means to segregate and manipulate the growing compounds on a solid support. Equipment
10 designed to simultaneously manipulate the plurality of gas dispersion tubes and provide an environment to perform standard organic synthesis techniques enables the multiple, simultaneous synthesis of, for example, 8, 40, 100, or more reactions at one time.

20 The steps necessary to perform a synthesis are 1) development of a synthetic route that will be feasible on a solid support, 2) verification of the resin-based synthesis using several representative examples, and 3) execution of multiple, simultaneous synthesis within an array format to generate, for
25 example, 8, 20, 40 unit arrays and the like.

The method involves the sequential coupling of building blocks to form resin-bound intermediates until the final or penultimate compound at each location in
30 the array is constructed, but still resin-bound. In addition to coupling the building blocks directly, one may add, if required, a coupling agent or reagent which is intended to chemically participate in forming the covalent bond between the solid support and the
35 building block or between building blocks. Coupling reagents include catalysts, chemical reagents, and the

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like. The sequential coupling reactions can be performed as illustrated in the following procedures:

- I. a functionalized solid support, a building block, a coupling reagent and solvent are reacted together; or
- 5 II. a functionalized solid support, a reactive building block and solvent are reacted together; or
- 10 III. a solid support with attached building block, a second building block, a coupling reagent and a solvent are reacted together; or
- IV. a solid support with attached building block, a second reactive building block, and a solvent are reacted together.

15 Preferably, the sequential coupling reactions can be performed as illustrated in the following procedures:

- I. (a) charging the apparatus with a solid support with attached building block wherein the building block has a reactive moiety protected by a protecting group and a solvent;
- 20 (b) removing the protecting group from the reactive moiety with a deprotection reagent;
- (c) removing the deprotection reagent;
- 25 (d) sequentially adding additional reactive building blocks in solvents to synthesize the compounds; and
- (e) cleaving the compounds from the solid support within the apparatus to afford the desired compounds; or
- 30 II. (a) charging the apparatus with a solid support with attached building block wherein the building block has a reactive moiety protected by a protecting group and a solvent;
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- (b) removing the protecting group from the reactive moiety with a deprotection reagent;
- (c) removing the deprotection reagent;
- (d) adding a coupling reagent in a solvent;
- 5 (e) sequentially adding additional reactive building blocks and optionally coupling reagents in solvents to synthesize the compounds; and
- (f) cleaving the compounds from the solid support within the apparatus to afford the desired compounds; or
- 10
- III. (a) charging the apparatus with a solid support with attached building block and a solvent;
- (b) adding a reagent for changing the oxidation state of the reactive moiety;
- 15 (c) sequentially adding additional reactive building blocks in solvents to synthesize the compounds; and
- (d) cleaving the compounds from the solid support within the apparatus to afford the desired compounds; or
- 20
- IV. (a) charging the apparatus with a solid support with attached building block and a solvent;
- (b) adding a reagent for changing the oxidation state of the reactive moiety;
- 25 (c) adding a coupling reagent in a solvent;
- (d) sequentially adding additional reactive building blocks and optionally coupling reagents in solvents to synthesize the compounds; and
- 30 (e) cleaving the compounds from the solid support within the apparatus to afford the desired compounds.

Other strategies for constructing the growing compounds on the solid support are possible and are encompassed within the scope of the present invention.

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Cleavage of the final compound from the resin yields a product which can be readily separated from the spent resin. Several options are available for achieving this cleavage and these are illustrated in Scheme 1. A single, invariant cleavage reagent can be employed to attack the resin-bound product linkage to yield a final compound containing an invariant functionality. Cleavage can be affected utilizing a variety of incoming building blocks to attack the resin linkage and give a product with variations in structure at the site of detachment. An alternative strategy constructs a precursor compound (resin-bound) possessing a distal functionality which, when activated or unmasked, will attack the resin-linking bond resulting in ejection of the cyclized final compound into solution. Since "unreacted" compounds remain attached to the resin, the latter option provides a means to produce cleaner final products.

The use of a solid support to multiply and simultaneously synthesize a subset of related, individual compounds requires a means of preparing the compounds in an array format. The method for constructing a compound array is illustrated with the following two examples: In one variant, the final compound, prior to detachment from the solid support, can be constructed from two building blocks/portions/halves utilizing a single coupling reaction to join the two smaller parts. One starts by selecting the congeners of building block #1 (for example, 3; A, B, and C) to be directly attached to the solid support and the number of congeners of the second half (building block #2) of the final compound that will be attached to the first building block (for example, 3; X, Y, and Z). The number of congeners of building block #1 multiplied by the number of congeners of building block #2 gives the number of locations contained in the

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array, in this example $3 \times 3 = 9$. Each of the congeners of #1 is covalently attached to the solid support a number of times equal to the number of congeners of #2, herein each of the first building blocks (A, B, and C) is coupled to the support in three locations each (9 couplings total). The covalent joining of the second building block to the first building block is now carried out with each of the congeners of building block #2 (i.e., A, B, and C are each coupled once with Z giving AZ, BZ, and CZ). Completion of the progression of couplings yields all nine expected permutations. This is illustrated in Scheme A.

SCHEME A. Construction of an Array Involving One Coupling Reaction

Building Block #2	Building Block #1		
	A	B	C
X	AX	BX	CX
Y	AY	BY	CY
Z	AZ	BZ	Cz

To achieve additional structural variation, one can utilize the addition of a third set of building blocks (for example, 3; 1, 2, and 3) and thus a second coupling reaction to provide 27 elements from an array of $3 \times 3 \times 3 = 27$. To accomplish this in the array synthesis will require that the first array (building block #1 coupled to building block #2 which is nine elements) be replicated three times so that when the third building block is added, the final elements are produced separately. This would be carried out as is illustrated in Scheme B. The expansion of the number of congeners within each building block set and the

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expansion of the number of discrete building blocks can be carried to whatever level desired to prepare arrays of any desired size or structural variability.

5 An alternative array construction can be carried out using a large number of congeners of several building blocks but choosing not to prepare every permutation possible (for example, two building blocks each with 30 congeners leads to 900 possible compounds). In this instance quantitative structure
10 activity relationship techniques and statistical methods can be used to select the most desired subsets of congeners to employ in preparing a smaller array.

15 SCHEME B. Construction of an Array Involving Two Coupling Reactions

20	Building Block #2	Building Block #1 (A, B, C)								
		A	B	C	A	B	C	A	B	C
	X	AX1	BX1	CX1	AX2	BX2	CX2	AX3	BX3	CX3
	Y	AY1	BY1	CY1	AY2	BY2	CY2	AY3	BY3	CY3
	Z	AZ1	BZ1	CZ1	AZ2	BZ2	CZ2	AZ3	BZ3	CZ3
25		1			2			3		
		Building Block #3 (1, 2, 3)								

20 To achieve the foregoing objects and in accordance with the purpose of the invention, as embodied and
30 broadly described herein, the invention provides an apparatus for the multiple, simultaneous synthesis of compounds comprising: (a) a plurality of reaction tubes, with each reaction tube having a filter device
35 on its lower end; (b) a reservoir block which has a means for containing a plurality of reaction wells and receiving the filter devices on the lower ends of the plurality of reaction tubes; (c) a holder block with a plurality of apertures which correspond to the location
40 of the plurality of reaction wells in the reservoir

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block and the plurality of reaction tubes; (d) a manifold located adjacent to the holder block, with the lower end of the manifold open such that the manifold may be placed on the holder block and surround the upper ends of the reaction tubes which are protruding upward through the apertures in the holder block; (e) a means for providing a sealed connection which is impermeable to gases and liquids between the holder block and the manifold and the holder block and the reservoir block; and (f) a means for fastening together the components of the apparatus. Figures 1-10 illustrate the components and embodiments of the apparatus.

In a second embodiment, the upper end of the manifold has a plurality of apertures which correspond in location to the apertures in the holder block. In this second embodiment, a plate which has a plurality of apertures which correspond in location to the apertures in the upper end of the manifold, and a means for providing a seal between the plate and the upper end of the manifold is provided. Figures 1 and 6 illustrate the components of this particular embodiment.

A further advantage of the present invention is that the apparatus provides the ability to monitor the reaction process by removal of a filtrate aliquot from the reaction well and analyze the solution by common chromatographic methods, such as Gas Chromatography/Internal Standard (GC/ISTD); High Pressure Liquid Chromatography/Internal Standard (HPLC/ISTD) or Thin Layer Chromatography (TLC), titration, colorimetry, spectroscopic methods, and the like. Additionally, by providing a separate reaction vessel for each reaction, the apparatus allows for the integrity of the filtrates, intermediates, and compounds which are generated.

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The apparatus of the present invention has the additional advantages of being constructed of materials which are chemically compatible with organic reagents, such as corrosive acids and organic solvents, required for organic reactions. The present invention has the further advantage of having the ability to provide a suitable means for the manipulations, such as agitation, heating, cooling, refluxing, and an inert atmosphere, common to organic synthesis.

Additional objectives and advantages of the invention will be set forth in part in the description that follows, and in part will be obvious from this description, or may be learned by practice of the invention. The objects and advantages of the invention may be realized and attained by means of the instrumentalities and combinations particularly pointed out in the appended claims.

BRIEF DESCRIPTION OF THE DRAWINGS

While this specification concludes with claims particularly pointing out and distinctly claiming that which is regarded as the present invention, the objects and advantages of this invention may be more readily ascertained from the following description of a preferred embodiment when read in conjunction with the accompanying drawings.

Figure 1 is an illustration of the components of the preferred embodiment of the apparatus of the invention.

Figure 2 is a perspective view of the components of the preferred embodiment of the apparatus of the invention.

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Figure 3 is an exploded perspective view showing each of the components of the preferred embodiment of the apparatus of the invention.

Figure 4 is a cross-sectional view Figure 3.

5 Figure 5 is an enlarged partial cross-section of Figure 4.

Figure 6 is an exploded perspective view of the preferred embodiment of the invention.

10 Figure 7 is an alternative embodiment for pressure equalization by a jacketed gas dispersion tube.

Figure 8 is an alternative embodiment for pressure equalization by a capillary tube.

Figure 9 is an alternative embodiment of the components for providing a gas-tight seal.

15 Figure 10 is an illustration of the solid-phase extraction equipment for postcleavage manipulations.

DESCRIPTION OF THE PREFERRED EMBODIMENTS

20

Reference will now be made in detail to the presently preferred embodiments of the invention.

25 The apparatus of the present invention comprises a plurality of reaction vessels. For the sake of illustration only, the accompanying drawings and description describe a device containing eight such reaction vessels, unless otherwise described. A device having a greater or lesser number of reaction vessels is entirely within the scope of the invention.

30 Further, the apparatus is described in the accompanying drawings as having a horizontal cross-section which is rectangular in shape. An apparatus having a square or circular horizontal cross-section is also entirely within the scope of the present invention.

35 Additionally, the apparatus may be adapted to be fully

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automated or semiautomated, and such adaptations are also within the scope of invention.

An initial element of the apparatus (10) as shown in Figures 1-6 is reaction vessels which can be any
5 vessels capable of containing resin and organic reactants. In the preferred embodiment of the invention, the reaction vessels are a combination of reaction tubes and reaction wells. The reaction tubes are most preferably gas dispersion tubes (11). The
10 lower ends of the reaction tubes have filters (14) and one or more pressure equalization holes (40) pierced in the reaction tubes (11) above the filters (14). The total length of each reaction tube can be from 50 to 300 mm with a preferred length of 250 mm. The upper
15 ends of the reaction tubes may have an outside diameter of from 5 to 25 mm, with a preferred outside diameter of 8 mm. The inside diameter of the upper ends of the reaction tubes can be from 1 to 24 mm, with a preferred inside diameter of 5 mm. To allow for the materials in
20 the reaction tube to mix with reactants, a filter (14) should be located in the lower end of the reaction tube (11). The length of the filters (14) on the lower ends of the reaction tubes can be from 1 to 300 mm, with a preferred length of 25 mm. The filters (14) on
25 the lower ends of the reaction tubes may have an outside diameter of from 5 to 30 mm, with a preferred outside diameter of 12 mm. The filters (14) on the lower ends of the reaction tubes may have an inside diameter of from 1 to 24 mm, with a preferred inside
30 diameter of 5 mm. Preferably, in order to allow the maximum reaction between material placed in the reaction tube and surrounding reactant, this filter (14) is preferably constructed of fritted glass. The porosity of the frit may be modified or selected to
35 accommodate various mesh sizes of solid supports and to affect more efficient reaction mixing. The pore

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diameter may be from 10 to 400 μm , with a preferred pore diameter of 145 to 175 μm for 200 to 400 mesh resin. The reaction tubes, including the filters, must be constructed of a material which will not be harmed by the presence of the volatile and/or corrosive reactants which are often necessary to carry out organic chemical reactions. Such materials include glass, polytetrafluoroethylene (PTFE), stainless steel, ceramics, or alternate glasses, metals, or plastics, preferably glass. One or more (preferably one) pressure equalization holes (40) located on the reaction tube at a position above the filter and below the bottom face of the holder block provides a means to equilibrate any pressure differentials within the sealed apparatus. The position of the pressure equalization holes above the filter may be from 1 to 250 mm, with a preferred height of 20 mm above the top of the filter (14). The diameter of the pressure equalization holes must be small enough to prevent loss of resin through them or cause significant loss of air pressure during sparging. The pressure equalization holes may have a diameter from 100 to 2000 μm , with a preferred diameter of 1000 μm .

A second component of the apparatus of the present invention as shown in Figures 1-6 is a reservoir block (15). The reservoir block (15) contains a plurality of reaction wells (16). In a preferred embodiment, the reservoir block consists of a reservoir rack (41) which is adapted to hold at least as many removable reaction wells (16) as the number of reaction tubes (11). The reservoir rack is constructed of a material such as aluminum, stainless steel, PTFE, ceramics, or various types of plastics, glasses, or metals. The reaction wells (16) are constructed of a solid, nonporous material which is capable of withstanding the reactive components, manipulations,

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and pressures required for organic synthesis. Such materials include glass, stainless steel, PTFE, ceramics, and various types of plastics, metals, or glasses, preferably glass. The reaction wells (16) in the reservoir block (15) should be sufficient in number, depth and diameter to accommodate the filters (14) on the lower ends of the reaction tubes (11) with each individual reaction well (16) suitable for receiving the lower end of one reaction tube, and to simultaneously hold a quantity of reactant necessary to perform the required reactions. The reaction wells (16) should be located throughout the reservoir block (15), such that each reaction well (16) is not in direct contact with any neighboring reaction wells. The depth of the reaction well may be from 1 to 300 mm, with a preferred depth of 50 mm. The inner diameter of the reaction well may be from 7 to 100 mm, with a preferred inner diameter of 18 mm. While not necessary to fully utilize the apparatus of the present invention, in a preferred embodiment the reaction wells are constructed of a transparent material, such as glass, plexiglass, or alternate glasses or plastics, so that the reactions may be visually monitored.

A further component of the invention as shown in Figures 1-6 is a holder block (18) located adjacent to the reservoir block (15). A plurality of apertures (19) are located throughout the holder block (18). These apertures (19) correspond in location, and preferably in number, to the reaction wells (16) located in the reservoir block (15), such that when the holder block (18) is placed on top of the reservoir block as in Figures 1-6, the apertures (19) in the holder block (18) are located directly above the reaction wells (16) in the reservoir block (15). The apertures (19) in the holder block (18) must be of sufficient diameter so that the upper ends (12) of the

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reaction tubes (11) will pass through the apertures. The diameter of the apertures (19) must not be so large, however, that they do not closely surround and support the upper ends (12) of the reaction tubes (11).

5 Such support allows the holder block to secure the reaction tubes so as to prevent them from torquing, wobbling, or moving vertically. The holder block also provides a means for physically handling and manipulating the plurality of reaction tubes (11) as a
10 single unit. The holder block (18) is preferably constructed from a heat-resistant material, such as PTFE, stainless steel, ceramics, glass, or certain types of metals, glasses, or plastics, so that it will be able to withstand organic reaction conditions.

15 The invention further includes a manifold (20) which, as shown in Figures 1-6, is located adjacent to the holder block (18). The manifold (20) is in the form of a hollow chamber. Preferably, the manifold consists of four side walls (21) and a top wall (22),
20 and has an opening on the lower end. In the preferred embodiment of the invention, the upper ends (12) of the reaction tubes, (11) after passing upward through the apertures (19) in the holder block, (18) enter and are enclosed within the manifold (20). When located
25 adjacent to the holder block, the manifold provides an air-tight chamber which facilitates manipulations common to organic synthesis. In a preferred embodiment, the manifold (20) will have a plurality of ports (23), preferably two, located on at least one of
30 the side walls (21) of the manifold (20). These ports may be aligned in any configuration and can be adapted to perform as inlet and outlet ports so that gaseous and/or liquid materials may be introduced into and recycled out of the manifold. The ports (23) allow
35 control over the atmosphere within the manifold (20) and provide a means to sparge the reaction tubes (11).

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While not necessary to fully utilize the apparatus of the present invention, in a preferred embodiment the manifold is constructed of a transparent material, such as glass, plexiglass, or alternate glasses or plastics, so that the reactions may be visually monitored.

Figure 3 illustrates the interface of the manifold (20) and the holder block (18), which in the preferred embodiment of the invention contains a first gasket or plurality of gaskets (24) placed between the holder block and the manifold. Such placement of a first gasket(s) creates a sealing effect between the manifold and the holder block in order to allow manipulations such as pressurization, inert atmosphere, and chilled gas circulation to take place. The first gasket(s) (24) surrounds the reaction tubes (11). The first gasket(s) may be made from PTFE and/or various rubbers and/or various plastics, with preferred types of rubber being neoprene, silicone, and Viton® (copolymer of hexafluoropropylene and 1,1-difluoro-ethylene). In a further embodiment, a second gasket or plurality of gaskets (26) may be located at the interface of the reservoir block (15) and the holder block (18). The second gasket(s) may be made from PTFE and/or various rubbers and/or various plastics, with preferred types of rubber being neoprene, silicone, and Viton®.

Figure 6 illustrates one further preferred embodiment of the invention. In this embodiment, the top wall of the manifold (28) contains apertures (29) which, when the manifold (20) is located on the holder block (18), correspond in number and spatial location to the apertures (19) in the holder block (18). In conjunction with this embodiment, a plate (30) with apertures (31) corresponding in number and spatial location to the apertures (29) in the top wall (28) of the manifold (20) is placed on the top wall of the

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manifold. A third gasket or plurality of third gaskets (34) placed between the top wall of the manifold and the plate allows the manifold (20) to remain sealed despite the apertures (29) in the top wall (28) of the manifold. In this embodiment, the third gasket(s) (34) may be made from PTFE and/or various rubbers and/or various plastics, with preferred types of rubber being neoprene, silicone, and Viton® so that it is puncturable with a needle-like object and reseals following the puncture.

A further component of the invention is a means for fastening all of the various elements together. Any means may be used so long as the elements are securely fastened together. In one embodiment, the fastening means consist of spring clips affixed to the sides of the components of the apparatus so that the elements may be releasably connected to each other. As shown in Figures 2-6, one or more spring clips (35) may be used for releasably connecting the manifold (20) and the holder block (18). When spring clips (35) are fastened, the manifold and the holder block may be fastened together and moved in tandem, and separated from the reservoir block (15). Such an embodiment is especially useful for removing the reaction tubes, in combination with the holder block and the manifold, from the reservoir block and draining off any excess reactant. As shown in Figure 2, spring clips (36) are provided at the interface of the holder block (18) and the reservoir block (15) and, when fastened, allow the holder block (18) and the reservoir block (15) to be fastened together independently of the manifold. (20) Spring clips (35) and spring clips (36) may all be fastened simultaneously, at which point the manifold (20), the holder block (18), and the reservoir block (15) are all tightly fastened as a single unit.

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In a further preferred embodiment, illustrated in Figure 6, the fastening means consist of a plurality of rods (32) located within the apparatus and which extend for the entire length of the apparatus, from the bottom wall of the reservoir block through the top wall of the manifold, or in a second embodiment, the plate located adjacent the top wall of the manifold. The rods (32) are preferably grooved at each end so that tighteners (33), such as wing-nuts, may be tightly screwed onto the rods so as to fasten the elements together. In a preferred embodiment, such rods are located at each of the four corners of the apparatus and additional intermediate locations dependent on the size of the apparatus.

In the embodiment illustrated in Figure 6, a groove (38) is provided along the edge of the plate (30). Two or more spring clips (37) are attached to the side wall of the manifold (20) and snap into the grooves (38) on the upper side of the plate (30). When spring clips (37) are fastened, plate (30) and gasket(s) (34) are held tightly against the top wall (28) of the manifold (20), and allow the manifold to maintain a seal despite the presence of apertures (29). This particular embodiment is most useful when sensitive chemicals are being used in the reaction. The use of spring clips as a fastening means may be in addition to or instead of the use of the plurality of rods as earlier set forth.

Figures 7 and 8 illustrate alternative embodiments for equalization of pressure between the reaction tube (11) and the reaction well (16). Figure 7 illustrates a jacketed reaction tube (11). A hollow tube (45), constructed of a material such as glass, stainless steel, or PTFE, is placed as a jacket around the upper end of the reaction tube (11). The outside diameter of the jacket (45) may be from 8 to 36 mm,

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with a preferred outside diameter of 15 mm. The inside diameter of the jacket (45) may be from 6 to 33 mm, with a preferred inside diameter of 12 mm. The length of the jacket (45) can be from 1 to 300 mm, with a preferred length of 200 mm. The jacket may extend below the lower face of the holder block (18), into the reaction well (16). The length of the jacket (45) extends below the lower face of the holder block (18) by 0 to 300 mm and the reaction tube (11) is held in place by one or more protrusions (46) on the inside face of the jacket at two or more positions along the length of the jacket (45). These protrusions (46) are constructed of one or more of the following materials: glass, PTFE, or alternate plastics or glasses. Accordingly, the apertures (19) in the holder block (18) must be of sufficient diameter so that the jacket (45) will pass through the apertures.

Figure 8 illustrates a capillary pressure release tube (47). A hollow tube, constructed of a material such as glass, stainless steel, PTFE, or alternate plastics or glasses, is placed in an aperture (48) in the holder block (18) directly adjacent to and parallel to the reaction tube (11) and within the inner diameter of the reaction well (16). The apertures (19) in the holder block (18) must be of sufficient diameter so that the capillary tubes will pass through the apertures. The diameter of the apertures (19) must not be so large, however, that they do not closely surround and support the capillary. Such support allows the holder block to secure the capillary tubes so as to prevent them from torquing, wobbling, or moving vertically. The outside diameter of the capillary tube (47) may be from 2 to 13 mm, preferably 2 mm. The inside diameter of the capillary tube (47) may be from 1 to 10 mm, preferably 1 mm. The length of the capillary tube (47) may be from 1 to 300 mm, with a

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preferred length of 200 mm. The lower end of the capillary tube (47) may extend below the lower face of the holder block (18), into the reaction well (16). Preferably, the lower end of the capillary tube (47) extends below the lower face of the holder block (18) by 0 to 300 mm. Accordingly, in this embodiment a second set of apertures (48) in the holder block (18) and sealing gaskets (24 and 26) must be of sufficient diameter to accommodate the capillary tube (47). The upper end of the capillary tube extends into the manifold (20).

Figure 9 illustrates an alternative embodiment for the sealing and containment of the reaction utilizing an o-shaped ring (42) seal between the reaction tube (11), the holder block (18), and the reaction well (16) to provide a greaseless, vacuum tight connection. The first spherical joint (43), opening downward, tooled with a groove to accommodate the o-shaped ring (42) is fused to the outer wall of the reaction tube (11). A second spherical joint (44) having the same diameter and groove size of the first spherical joint and opening upward to accommodate the same o-shaped ring (42) is constructed at the top of the corresponding reaction well (16). The outside diameter of the first and second spherical joints (43 and 44) may be from 5 to 50 mm. The position of the first spherical joint (43) fused to the outer wall of the reaction tube (11) is above both the filter (14) and pressure equalization hole (40) and at a height necessary to adequately accommodate the glass frit within the sealed reaction well (16) when the seal is achieved. The position of the first spherical joint (43) may be from 1 to 299 mm above the top of the filter (14), with a preferred height of 25 mm above the filter, corresponding to the preferred height of the filter (14) and preferred depth of the reaction

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well (16) described above. The position of the corresponding second spherical joint (44) fused to the top of the reaction well (16) is also at a height necessary to adequately accommodate the filter (14) within the sealed reaction well (16) when the seal is achieved. The position of the second spherical joint (44) may be from 1 to 299 mm above the bottom of the reaction well (16), with a preferred height of 50 mm above the bottom of the reaction well (16). In this embodiment, the faces of the apertures on the lower side of the holder block (18) should be chamfered to accommodate the first spherical joint (43) attached to the outer wall of the reaction tube (11). Furthermore, the faces of the apertures on the upper side of the reservoir rack (41) should be chamfered to accommodate the second spherical joint (44) attached to the reaction well (16). The use of an o-shaped ring seal obviates the need for the second gasket (26) between the holder block (18) and the reservoir block (15).

Figure 10 illustrates the solid phase extraction (SPE) system used for post-cleavage manipulations of the final reaction solutions. The SPE system consists of a first rack (50) having a frame which supports one or more test tubes (51), preferably the same number of test tubes as compounds synthesized in the array set forth above, for example, 8, 40, or 100 compounds. The first rack can be the same as, or compatible with, the reservoir block (15), reservoir rack (41), and reaction wells (16) described above and illustrated in Figures 1-6. The test tubes (51) are constructed of a solid, nonporous material such as glass, stainless steel, PTFE, ceramics, or various types of plastic. Preferably, the test tubes are constructed of glass. During operation of the SPE system, the first rack, in combination with the test tubes, is placed in a vacuum

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box (52). One or more valves (53) control the pressure within the vacuum box. A second rack (54) which supports one or more removable cartridges (55) fits on top of the vacuum box. The first and second

5 racks (50 and 54) are constructed of a material such as aluminum, stainless steel, PTFE, ceramics, or various types of plastics, glasses, or metals. One or more gaskets (55) are adhered to the lower side of the base of the second rack (56) to provide a means for

10 achieving a seal between the vacuum box and the second rack. One or more spring clips (57) are attached to the side walls of the vacuum box (52), preferably two clips are located on opposing sides, and snap into the grooves (58) in the upper side of the base of the

15 second rack (54). When the spring clip (57) is fastened, the second rack (54) and gasket (55) are held tightly against the top wall (59) of the vacuum box (52) to maintain a seal. One or more needle-like objects (60), corresponding in number and location to

20 the test tubes (51), are mounted in the base of the second rack (54), protrude through the gasket (55), and open into the test tubes (51). Removable cartridges (61) are used in combination with the needle-like objects (60). The lower end of each

25 cartridge opens into the upper end of the corresponding needle-like object. The cartridges may be filled with solids or solid supports such as silica gel, drying agents, cellulose, and the like, and then eluted with solvents or reagent solutions which may be eluted

30 through the cartridges, through the needles, and into the corresponding test tubes.

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GENERAL DESCRIPTION OF EXPERIMENTAL PROCEDURE

A number of reaction tubes equal to the total number of compounds to be synthesized by the array method are loaded with 1 to 1000 mg, preferably 100 mg, of the appropriate functionalized solid support, preferably 1 to 3% cross-linked polystyrene. The individual reaction tubes are inserted into the holder block. The reaction tubes, in combination with the holder block and manifold, are inserted into the reservoir block so that each reaction tube is submerged in a volume, preferably 3 to 5 mL, of a solvent capable of swelling the polystyrene resin (such as, but not limited to, dichloromethane, chloroform, dimethylformamide (DMF), dioxane, toluene, tetrahydrofuran (THF), ethanol and the like). The polystyrene resin within the reaction tubes is preferably agitated for 15 to 30 minutes to affect swelling. Swelling times of 5 to 600 minutes, with or without agitation are also within the scope of the invention. The reaction tubes, in combination with the holder block and manifold, are removed from the reservoir block and the excess solvent within the reaction tubes is allowed to drain, preferably by gravity, although gas pressure applied to the manifold inlet (while closing the outlet) can be used to expel the solvent, if desired.

The reaction wells are emptied of solvent and the proper reactant solutions are dispensed into the new or clean reaction wells at appropriate locations in the reservoir block. If the reactant is actually one of the building blocks that is to become covalently attached to the growing compound on the solid support, the quantity of reactant is usually 1 to 100 equivalents based on the milliequivalents per gram (meq/g) loading of functionalized solid support (typically 0.1 to 1.0 meq/g for polystyrene resins).

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originally weighed into the reaction tube. Additional equivalents of reactants can be used if required to drive the reaction to completion in a reasonable time. The reaction tubes, in combination with the holder
5 block and manifold, are reinserted into the reservoir block and the apparatus is fastened together. Gas flow through the manifold is initiated to provide a controlled environment, for example, nitrogen, argon, air, and the like. The gas flow may also be heated or
10 chilled prior to flow through the manifold. Heating or cooling of the reaction wells is achieved by immersing the reservoir block in water baths, oil baths, isopropanol/dry ice baths, sand baths, and the like to perform synthetic reactions. Agitation is achieved by
15 shaking, sonication (preferred), or magnetic stirring (within the reaction well or within the reaction tube). Reflux is achieved by circulating chilled gas through the manifold while heating the reaction wells in the reservoir block. Reactants may be injected directly
20 into the reaction tubes through an injectable gasket on the top of the manifold. The reaction is allowed to proceed for an amount of time deemed necessary from the preliminary validation experiments or monitored by removal and quantitative analysis of filtrate aliquots
25 from selected wells by methods such as GC/ISTD or HPLC/ISTD. If necessary, the complete assembly is allowed to return to ambient temperature, then the holder block and manifold, in combination with the reaction tubes, are detached from and raised above the
30 reservoir block and the excess reagent solution is drained by gravity followed by gas pressure applied to the manifold inlet (while closing the outlet) to expel the excess reagents, solvents, and by-products. The resin-bound intermediate within each reaction tube is
35 washed clean of excess retained reagents, solvents, and by-products by repetitive exposure to clean solvent(s)

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by one of two methods: 1) the reaction wells are filled with solvent (preferably 1-5 mL), the reaction tubes, in combination with the holder block and manifold, are immersed and agitated for 5 to 300 minutes, preferably 15 minutes, and drained by gravity followed by gas pressure applied through the manifold inlet (while closing the outlet) to expel the solvent; 2) the manifold is removed from the holder block, aliquots of solvent (preferably 5 mL) are dispensed through the top of the reaction tubes and drained by gravity through the filter into a receiving vessel such as a test tube or vial. Both of the above washing procedures are repeated 1 to 50 times (preferably 10 times), monitoring the efficiency of reagent, solvent, and byproduct removal by methods such as TLC, GC, or visualization of the wash filtrates.

The above described procedure of reacting the resin-bound compound with reagents within the reaction wells followed by removal of excess reagents, byproducts, and solvents is repeated with each successive transformation until the final or penultimate resin-bound compound is prepared.

Detachment of the final product from the solid support is achieved by immersion of the reaction tubes, in combination with the holder block and manifold, in reaction wells containing a solution of the cleavage reagent (preferably 3-5 mL). Gas flow, temperature control, agitation, and reaction monitoring are implemented as above and as desired to affect the detachment reaction. The reaction tubes, in combination with the holder block and manifold, are disassembled from the reservoir block and raised above the solution level but below the upper lip of the reaction wells and gas pressure is applied through the manifold inlet (while closing the outlet) to efficiently expel the final product solution into the

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reservoir wells. The spent resin in the reaction tubes is then washed 2 to 5 times as above with 3 to 5 mL of an appropriate solvent to extract (wash out) as much of the detached product as possible. The final product solutions are combined, taking care to avoid cross-mixing. The individual solutions/extracts are then manipulated as needed to isolate the final compounds. Typical manipulations include, but are not limited to, evaporation, concentration, liquid/liquid extraction, acidification, basification, neutralization or additional reactions in solution.

The final compounds are individually tested for biological activity once they are isolated. For example, the method of Sweetnam, et al, Molecular Pharmacology 1986;29:299 employing bovine cortical membranes as a receptor source and a radiolabeled benzodiazepine, such as Flunitrazepam, to determine competitive ligand binding to the benzodiazepine receptor (central [brain] localized) and some quantitative measure of relative binding potency such as K_D or IC_{50} values. Performing this type of assay serves two purposes: 1) to discover new compounds with biological activity in a given biological screening assay and 2) the development of a relationship between the structural variations contained within the series and biological potency. This second utility is known as development of a structure activity relationship (SAR). This type of assay can be done using the compounds isolated from the array synthesis for any receptor binding screen or assay (screening for receptor agonists or antagonists), enzyme functional assays (measuring competitive or noncompetitive inhibition of the catalyzed reaction), and the like.

This strategy may be used to screen for pharmaceutical agents, veterinary agents, agricultural agents, diagnostic reagents, and the like.

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Typical compounds and pharmaceutical applications include: (1) nitrogen containing heterocyclic compounds; imidazopyridines having antiulcer or anxiolytic activities, dihydropyridines having calcium antagonist activity, nucleoside and nucleoside analogs having antiviral activity, indazoles having 5HT₂ antagonist activity, piperidines having antidopamine, antiserotonin, antidepressant, or antihistamine activities, benzazepines having antiparkinsonism and antidopamine activities, indoles and condensed indoles with 5HT antagonist activities, quinolines and isoquinolines having anti-infective and antiulcer activities, pyrrolidines having anti-infective and antihypertensive activities, aminopyrimidines having antihypertensive activities, pyrrolizidines having antiarrhythmic activities, guanidines having anticancer activities; tetrazoles having antiallergenic; (2) oxygen containing heterocycles; benzopyrans having potassium agonist and antagonist activities, coumarins having antiplatelet aggregating and antithrombotic activities, prostaglandins and prostacyclins having antiplatelet, antiulcer, labor inducing activities, psoralens having antipsoriasis activities, tetrahydrofurans and pyrans having antidiabetic activity; (3) nitrogen and sulfur containing compounds; beta-lactams and cephalosporins having anti-infective activities; (4) carbocyclic compounds; tocopherol analogs having antipsoriasis activities, vitamin D analogs having antipsoriasis activities, steroids having anti-inflammatory, bronchiodilating, antihyperplasia and antifertility activities, naphthalenes having antifungal activities, anthracene analogs having anticancer activities; (5) alicyclic compounds; polyunsaturated alkenes having antithrombotic activities, hydroxypropanolamines having adrenergic blocking activities, benzofused bicyclic

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amines having analgesic activities, aryl amides having
anesthetic, gastroprokinetic, antidepressant, and anti-
inflammatory activities; and (6) cyclic peptides and
cyclic nucleotide having anti-infective and
5 antiautoimmune activities; and the like.

In alternative uses, the apparatus and methods may
be implemented without using a solid support, for
example, standard solution techniques. An additional
alternative use of the apparatus is for the
10 optimization of chemical reactions, for example,
reaction yields or reaction times, on solid supports or
in solution. This use is accomplished by performing
the same reaction in all reaction well locations but
systematically determining and varying dependent
15 reaction variables across the array. For example, the
reagent concentration and/or reagent equivalency (mole
percentage) can be varied from near zero to the maximum
achievable with an undiluted reagent. Typically, one
varies this variable from 0.001 to 25 mol. Reaction
20 times can be varied within the array by withdrawal of
the contents of the reaction well, followed by for
example workup, and/or isolation, and/or purification,
and/or quantitation of the final solution. Statistical
experimental design strategies or quantitative
25 structure activity relationship (QSAR) strategies may
also be implemented to select a subset of locations
within the original array which will provide the
necessary information for final analysis and
conclusions, therefore reducing the number of reactions
30 necessary in the final array.

Synthesis of Dipeptides

In the operation of the present invention, the
synthesis of dipeptides is achieved using the apparatus
35 and applying 9-fluorenylmethyloxycarbonyl (Fmoc)
strategy, as taught in Meienhofer, et al, International

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Journal Peptide Protein Research 1979;13:35 and Atherton, et al, Bioorganic Chemistry, 1979;8:351 on a variety of commercially available polystyrene resins (see Table 1). For peptide acids the p-benzyloxy benzyl alcohol cross-linked divinylbenzene-styrene (WANG) resin, as taught in Wang, Journal American Chemistry Society 1973;95:1328, is utilized. Eight Fmoc amino acid resins including phenylalanine, glycine, alanine, isoleucine, leucine, proline, valine, and tryptophan are deprotected, reacted with either Fmoc alanine or Fmoc isoleucine, deprotected again, then cleaved from the resin to generate 16 discrete dipeptides. A total of 3 to 20 mg (28-85%) of each crude dipeptide is isolated as the trifluoroacetic acid (TFA) salt and analyzed by HPLC, mass spectroscopy (MS), and proton nuclear magnetic resonance spectroscopy (^1H NMR) (see Table 1).

Synthesis of Hydantoins

The synthesis of an array of 40 hydantoins, including phenytoin, is achieved using the apparatus described herein. Five samples of eight Fmoc or Boc protected amino acid resins (phenylalanine, glycine, isoleucine, leucine, alanine, valine, tryptophan, and 2,2-diphenylglycine) are deprotected then separately reacted with five isocyanates (trimethylsilyl, n-butyl, allyl, 2-trifluorotolyl, or 4-methoxyphenyl isocyanate) followed by treatment with aqueous 6N hydrochloric acid (HCl) to generate 40 discrete hydantoins (see Scheme 2 and Table 2).

The synthesis of 40 hydantoins in an array illustrates a standard protocol for a typical parallel array synthesis. In this example, a robotic sample processor (Tecan® 5032) is used for every liquid sample handling step. Some of the key features which demonstrate the strengths, flexibility and scope of the

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method and apparatus are highlighted below. Forty BOC or FMOC protected resin-bound amino acids are weighed into the reaction tubes, assembled in the apparatus, and simultaneously deprotected employing GC/ISTD calibration methods to determine the completion of the FMOC deprotection reactions. The ability to simultaneously perform different reactions with vastly different reagents (for example, piperidine, DMF and TFA) further demonstrates the flexibility of the apparatus. Residual solvents, reactants, or byproducts are removed by wash cycles which include submersion and sonication of the reaction tubes in a series of solvents, followed by robotic spotting of the filtrates on a TLC plate and observation of the results under ultraviolet (UV) light to insure the thoroughness of the wash cycles. Following the wash cycles, the resin-bound amines are reacted with the desired isocyanates in DMF. Again, GC/ISTD calibration methods are used to quantitatively monitor the uptake of the isocyanates. After washing, the resin-bound ureas are cyclized by heating in aqueous 6N HCl to produce the desired hydantoins. Following the HCl treatment, the spent resins in the reaction tubes are washed with methanol to completely extract the hydantoins from the resins. All filtrates are combined, concentrated on a commercial centrifugal vacuum system (SpeedVac[®]), weighed, and analyzed to yield the expected hydantoins, in all but one example (see Table 2). A total of 0 to 11.5 mg corresponding to 0 to 81% yield of each crude hydantoin is isolated and analyzed by TLC, MS, and ¹H NMR.

Synthesis of Benzodiazepines and Biological Testing

The investigation of an array synthesis of benzodiazepines as potential targets was prompted by a brief communication by Camps, et al, Anales De Quimica

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1974;70:848 who reported a one-step synthesis of several benzodiazepines starting with a resin-bound amino acid. A two-step route is outlined in Scheme 3. The first step in the sequence is the formation of an imine between a resin-bound amino acid and a 2-amino-benzophenone. Initially, a number of condensation methods were explored, but all proved unsatisfactory. Replacement of the imine condensation with a trans-
5 imination reaction, as taught by O'Donnell and Polt, Journal Organic Chemistry 1982;47:2663, to form a mixture of E and Z imine isomers proved satisfactory. The resin-bound amino acid imines are then converted to the corresponding benzodiazepines by heating in TFA.
10

The synthesis of 40 benzodiazepines in the array is similar to that outlined for the dipeptides and
15 hydantoins with several modifications, which demonstrate the flexibility of both the method and the apparatus (see Scheme 3 and Table 3). Five amino acid Merrifield resins (alanine, glycine, 2-bromobenzyloxy-carbonyl-tyrosine, tryptophan, and valine) as their TFA
20 salts are separately reacted with eight 2-aminobenzo-phenone imines. The reactions are not monitored, but are reacted long enough to accommodate the slowest example (valine resin with N-isopropyl 2-amino-
25 4-methylbenzophenone imine) based on previous validation studies. Following wash cycles to extract the unreacted imine, the resin bound imines are heated in TFA for a time sufficient to ensure cyclization of the slowest examples (imines of N-methyl 2-amino-
30 5-nitrobenzophenone), while minimizing decomposition of the tryptophan derived benzodiazepines. When the reaction is complete, the spent resins in the reaction tubes are washed and the combined filtrates concentrated to dryness. In this case, an aqueous
35 bicarbonate work-up is implemented (utilizing the Tecan 5032 robotic sample processor) to remove

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residual TFA. For each reaction tube the corresponding organic extracts are combined, dried, and concentrated to yield the expected benzodiazepines in all but one example (see Table 3). The 40 (39 desired) products are characterized by TLC, ¹H-NMR, and MS. The crude yields range from 7 to >100% and the estimated purities from NMR and TLC are greater than 60% in most cases.

To verify that the compounds produced could be used directly in a biological assay, the crude benzodiazepines are tested for activity in a benzodiazepine receptor binding assay without further purification. (The assay was performed using the commercially available NovaScreen® assay system contracted by Scios-Nova Pharmaceutical Corp., Baltimore, MD, whereby bovine cortical membrane preparations were used as the source of receptor and the radioligand employed was [³H]Flunitrazepam while the positive control was Clonazepam.) The calculated IC₅₀'s from three concentrations (average of two determinations) for the individual compounds are shown in Table 3.

The following nonlimiting examples are offered by way of illustration and are not intended to limit the invention in any manner.

25

EXAMPLE 1

Synthesis of Dipeptides

The synthesis of 16 dipeptides is summarized in Table 1.

101 to 153 mg each of eight Fmoc protected amino acids (phenylalanine, glycine, alanine, isoleucine, leucine, proline, valine, and tryptophan) on a commercially available WANG resin (loading = 0.37-0.60 meq/g, 200-400 mesh) was measured into each of 16 gas dispersion tubes (reaction tubes). The

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reaction tubes were fitted into the holder block with two gaskets, one above and one below the holder block. The array of reaction tubes was fitted into a matching array of reaction wells in an appropriate reservoir block, in this case a test tube rack, and 3 mL of DMF was dispensed through the aperture at the top of the reaction tubes to wash any residual resin to the bottom of the glass frit. The manifold was fitted over the holder block and a nitrogen atmosphere was initiated through the ports on the manifold. The apparatus was then agitated in a sonic bath for 15 minutes to swell the resin support in preparation for the first reaction. Following swelling of the resin, the holder block and the manifold, in combination with the reaction tubes, were raised above the reservoir block and the reaction tubes were allowed to drain by gravity.

To deprotect the FMOC amino acids, the reaction tubes were submerged in reaction wells containing 2 mL of a solution of 22% piperidine in DMF (v/v) with an internal standard (e.g., anthracene at 1.97 mg/mL). The apparatus was fastened together with clamps and agitated in a sonic bath for 2 hours, while maintaining a positive nitrogen flow through the manifold. The reaction progress was monitored by removing a sample of the filtrate (10-100 μ l) and analyzing for the FMOC-piperidine adduct and dibenzofulvene by GC/ISTD calibration methods. At the end of the reaction, the holder block and the manifold, in combination with the reaction tubes, were raised above the reservoir block and the reaction tubes were allowed to drain by gravity, then sparged with nitrogen. The reaction tubes were sparged by introducing nitrogen into one port of the manifold and closing off the second port.

The reaction tubes were subjected to a series of wash cycles to remove residual solvents, reagents, and

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byproducts. A standard protocol included sequentially submerging the reaction tubes in 2 to 5 mL each of 2 x DMF, 2 x methanol, 2 x water, 2 x water:dioxane (1:1), 2 x dioxane, and 2 x DMF, followed by agitation
5 by sonication for 10 to 15 minutes, and finally draining by gravity and nitrogen sparge. The efficiency of the wash cycles was monitored by TLC or GC/ISTD of the wash filtrates.

In order to couple the Fmoc amino acids, the
10 following DMF solutions were sequentially admixed in two 8-unit reservoir blocks, to generate the activated esters: 1.0 mL of 0.46 M Fmoc alanine or Fmoc isoleucine, 0.5 mL of 1.02 M Benzotriazol-1-yloxy-tris-(dimethylamino)-phosphonium hexafluorophosphate (BOP),
15 1.0 mL of 1.02 M N-Hydroxybenzotriazole (HOBT), and 0.5 mL of 1.84 M N-methylmorpholine. The apparatus was assembled as above and agitated on a rotational platform shaker for 18 hours, while maintaining a positive nitrogen flow through the manifold. The
20 reaction progress was monitored by removing a sample of the resin following the wash cycles and colorometrically analyzing for primary amines (Bromophenol blue or Kaiser/Ninhydrin). At the end of the reaction, the holder block and the manifold, in
25 combination with the reaction tubes, were raised above the reservoir block and the reaction tubes were allowed to drain by gravity then sparged with nitrogen.

The reaction tubes were subjected to the same series of wash cycles as previously set forth.

30 The deprotection of the above formed Fmoc protected dipeptides was repeated following the same procedure as previously set forth.

The reaction tubes were again subjected to the same series of wash cycles as previously set forth.

35 In order to cleave the product from the solid support, the reaction tubes were submerged in reaction

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wells containing 3 mL of a solution of anisole:TFA (5:95). The apparatus was sealed with clamps and agitated in a sonic bath, while maintaining a positive nitrogen flow through the manifold. After 3 hours, the holder block and the manifold, in combination with the reaction tubes, were raised above the reservoir block and the reaction tubes were allowed to drain by gravity then sparged with nitrogen.

To isolate and purify the products, a holder block was fitted with disposable glass pipets which were submerged in the TFA filtrates. The filtrates were concentrated in a well-ventilated hood or glove box fitted with a scrubber by initiating and maintaining a positive, subsurface nitrogen flow through the manifold. The TFA salt was dissolved in 2 mL of water and extracted with 2 mL of ethyl acetate. The reaction reservoirs were vortexed to affect efficient extraction and the upper organic layer was withdrawn via syringe. The extraction cycle was repeated until the organic layer contained no residues as determined by TLC or GC/ISTD (3 times). The aqueous layers were concentrated on a Speed-Vac® in tared vials. The dipeptides generated in the apparatus including, alanylphenylalanine; alanylglycine; alanylisoleucine; alanylleucine; alanylalanine; alanylproline; alanylvaline; alanyltryptophan; isoleucylphenylalanine; isoleucylglycine; isoleucylisoleucine; isoleucylleucine; isoleucylalanine; isoleucylproline; isoleucylvaline; and isoleucyltryptophan, are set forth in Table 1.

EXAMPLE 2

Synthesis of Hydantoins

The synthesis of 40 hydantoins is summarized in Scheme 2 and Table 2.

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95 to 105 mg each of seven FMOC protected amino acids (phenylalanine, glycine, isoleucine, leucine, alanine, valine, and tryptophan) on a commercially available WANG resin (loading = 0.37 to 0.60 meq/g, 200 to 400 mesh) and 95 to 105 mg of BOC protected diphenylglycine loaded on a commercially available cross-linked hydroxymethyl divinylbenzene-styrene resin (loading = 1.04 meq/g, 200-400 mesh) was measured into 40 reaction tubes. The apparatus was assembled, swelled with 4 mL of DMF, and drained as set forth in Example 1.

To deprotect the FMOC amino acids, the appropriate reaction tubes were submerged in reaction wells containing 3 mL of a solution of 25% piperidine in DMF (v/v) with an internal standard (anthracene at 1.74 mg/mL). To deprotect the BOC amino acids the appropriate reaction tubes are submerged in wells containing 3 mL of 50% TFA/DMF. The apparatus was assembled as set forth in Example 1 and agitated in a sonic bath while maintaining a positive nitrogen flow through the manifold. The reaction progress was monitored by removing a sample of the filtrate (e.g. 10-100 μ l) and analyzing for the FMOC-piperidine adduct and dibenzofulvene by GC/ISTD calibration methods. The reaction was complete after 6 hours in a sonic bath. At the end of the reaction, the reaction tube array was drained and sparged as set forth in Example 1.

The reaction tubes were subjected to the standard wash cycle set forth in Example 1.

To facilitate urea formation, the appropriate reaction tubes were submerged in reaction wells containing 3 mL of a solution of 0.19 to 0.23 M isocyanate (trimethylsilyl isocyanate, butyl isocyanate, allyl isocyanate, trifluoro-o-tolyl isocyanate, and 4-methoxyphenyl isocyanate) in DMF, containing an internal standard (e.g., anthracene at

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1.89-2.00 mg/mL). The apparatus was sealed as set forth in Example 1 and agitated in a sonic bath for 6 hours, while maintaining a positive nitrogen flow through the manifold. The reaction progress was
5 monitored by removing a sample of the filtrate (10-100 μ l), derivatization with an appropriate amine or alcohol, and analysis by GC/ISTD calibration methods. At the end of the reaction, the reaction tube array was drained and sparged as set forth in
10 Example 1.

The reaction tubes were subjected to the standard wash cycle set forth in Example 1.

To facilitate cleavage of the final product from the solid support, the reaction tubes, in combination with the holder block and manifold, were submerged in
15 reaction wells containing 3 mL each of 6N HCl. The apparatus was sealed with clamps and submerged in an oil bath (at 105°C) and heated at 85 to 100°C while maintaining a positive chilled nitrogen flow through
20 the manifold. (The chilled nitrogen flow through the manifold was affected by submersion of the nitrogen inlet tubing in an isopropanol/dry ice bath.) After 2 hours, the reaction tube array was cooled, drained and sparged as set forth in Example 1.

25 To isolate and purify the products, the reaction tubes, in combination with the holder block and manifold, were submerged in reaction wells containing 3 mL each of methanol. The apparatus was agitated in a sonic bath for 10 to 15 minutes to extract the
30 hydantoins from the resins then drained by gravity and nitrogen sparge. The methanol extraction protocol was repeated until the filtrates were free of any organic components as determined by TLC (4 times). The HCl and methanol filtrates were concentrated on a Speed-Vac in
35 tared vials to afford 39 of the 40 desired hydantoins. The hydantoins generated in the apparatus, including:

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- 5-methyl-2,4-imidazolidinedione; 5-(phenylmethyl)-
 2,4-imidazolidinedione; 2,4-imidazolidinedione;
 5-(1-methylpropyl)-2,4-imidazolidinedione;
 5-(2-methylpropyl)-2,4-imidazolidinedione;
 5 5-(1-methylethyl)-2,4-imidazolidinedione; 5-(1H-indol-
 2-ylmethyl)-2,4-imidazolidinedione; 5,5-diphenyl-
 2,4-imidazolidinedione; 3-butyl-5-methyl-
 2,4-imidazolidinedione; 3-butyl-5-(phenylmethyl)-
 2,4-imidazolidinedione; 3-butyl-2,4-imidazolidinedione,
 10 3-butyl-5-(1-methylpropyl)-2,4-imidazolidinedione;
 3-butyl-5-(2-methylpropyl)-2,4-imidazolidinedione;
 3-butyl-5-(1-methylethyl)-2,4-imidazolidinedione;
 3-butyl-5-(1H-indol-2-ylmethyl)-2,4-imidazolidinedione;
 3-butyl-5,5-diphenyl-2,4-imidazolidinedione; 5-methyl-
 15 3-(2-propenyl)-2,4-imidazolidinedione; 5-(phenyl-
 methyl)-3-(2-propenyl)-2,4-imidazolidinedione;
 3-(2-propenyl)-2,4-imidazolidinedione;
 5-(1-methylpropyl)-3-(2-propenyl)-2,4-imidazolidine-
 dione; 5-(2-methylpropyl)-3-(2-propenyl)-
 20 2,4-imidazolidinedione; 5-(1-methylethyl)-
 3-(2-propenyl)-2,4-imidazolidinedione; 5-(1H-indol-
 2-ylmethyl)-3-(2-propenyl)-2,4-imidazolidinedione;
 5,5-diphenyl-3-(2-propenyl)-2,4-imidazolidinedione;
 5-methyl-3-[2-(trifluoromethyl)phenyl]-
 25 2,4-imidazolidinedione; 5-(phenylmethyl)-
 3-[2-(trifluoromethyl)phenyl]-2,4-imidazolidinedione;
 3-[2-(trifluoromethyl)phenyl]-2,4-imidazolidinedione;
 5-(1-methylpropyl)-3-[2-(trifluoromethyl)phenyl]-
 2,4-imidazolidinedione; 5-(2-methylpropyl)-
 30 3-[2-(trifluoromethyl)phenyl]-2,4-imidazolidinedione;
 5-(1-methylethyl)-3-[2-(trifluoromethyl)phenyl]-
 2,4-imidazolidinedione; 5-(1H-indol-2-ylmethyl)-
 3-[2-(trifluoromethyl)phenyl]-2,4-imidazolidinedione;
 5,5-diphenyl-3-[2-(trifluoromethyl)phenyl]-
 35 2,4-imidazolidinedione; 3-(4-methoxyphenyl)-5-methyl-
 2,4-imidazolidinedione; 3-(4-methoxyphenyl)-

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5- (phenylmethyl) -2,4-imidazolidinedione;
3- (4-methoxyphenyl) -2,4-imidazolidinedione;
3- (4-methoxyphenyl) -5- (1-methylpropyl) -2,4-imidazol-
idinedione; 3- (4-methoxyphenyl) -5- (2-methylpropyl) -
5 2,4-imidazolidinedione; 3- (4-methoxyphenyl) -
5- (1-methylethyl) -2,4-imidazolidinedione; 5- (1H-indol-
2-ylmethyl) -3- (4-methoxyphenyl) -2,4-imidazolidinedione;
3- (4-methoxyphenyl) -5,5-diphenyl-
2,4-imidazolidinedione; are set forth in Table 2.

10

EXAMPLE 3

Synthesis of Benzodiazepines.

The synthesis of 40 benzodiazepines is summarized
in Scheme 3 and Table 3.

15 Five commercially available BOC amino acid
Merrifield resins (alanine, glycine, 2-bromobenzyloxy
carbonyl-tyrosine, tryptophan, and valine;
0.57-0.89 meq/g, 200-400 mesh) were deprotected in bulk
(1-5 g) instead of within the apparatus, using
20 TFA:CH₂Cl₂ (1:1) at room temperature overnight. After
washing with dioxane and CH₂Cl₂, the resins were dried
under vacuum and used directly.

99 to 107 mg of each amino acid resin prepared
above, as its TFA salt, was loaded into 40 reaction
25 tubes. The apparatus was assembled, swelled with 3 mL
of CH₂Cl₂, and drained as set forth in Example 1. The
appropriate reaction tubes are submerged in reaction
wells containing a solution of the appropriate
2-aminobenzophenoneimines (3-6 eq) in 3 mL of
30 dichloroethane (see Scheme 2). The apparatus was
sealed as set forth in Example 1 and heated at 60°C
(oil bath temp.), while maintaining a positive chilled
nitrogen flow through the manifold. In this case the
reactions were not monitored, but were reacted for
35 sufficient time (24 hours) to drive the slowest
reaction (valine resin with N-isopropyl 2-amino-

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4-methyl benzophenone imine) to completion based on earlier validation studies. The reaction tube array was allowed to cool and then drained and sparged as set forth in Example 1.

5 In this example, the reaction tubes were washed by adding 4 mL portions of CH_2Cl_2 through the top of each reaction tube repeatedly (12 times) until the washes were no longer colored (indicating the absence of
10 2-amino benzophenone imine or its corresponding ketone from hydrolysis).

 The reaction tube array was then sparged as set forth in Example 1 and the reaction tubes are submerged in reaction wells containing 3 mL of 100% TFA. The apparatus is sealed as set forth in Example 1 and
15 heated at 60°C (oil bath temp.) for 20 hours, while maintaining a positive chilled nitrogen flow through the manifold as set forth in Example 2. Again, the reactions were not monitored, but were allowed to react for a time sufficient to ensure conversion of the
20 slowest reactions (N-methyl 2-amino-5-nitro benzophenone imine), while minimizing decomposition of the tryptophan derived benzodiazepines. The reaction tube array was allowed to cool and then drained as set forth in Example 1.

25 The reaction tubes are then washed as above with 3 x 2 mL portions of CH_2Cl_2 .

 The appropriate washes are then combined and evaporated under a stream of nitrogen as set forth in Example 1, except in this case the pipet tips were
30 maintained above the surface of the liquid. A simple extraction procedure was implemented using the Tecan® robot. The residues from evaporation were dissolved in 3 mL CH_2Cl_2 and mixed with 3 mL saturated sodium bicarbonate. The organic phase was withdrawn and the
35 aqueous layer was extracted twice more with 1.5 mL of CH_2Cl_2 . The combined organic extracts were dried

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with MgSO_4 , filtered, and concentrated as before to yield the expected benzodiazepines in all but one example (see Table 3). The 40 (39 desired) products were characterized by TLC, $^1\text{H-NMR}$, and MS. The crude yields range from 4 to >100% and the estimated purities from NMR and TLC are greater than 60% in most cases. The desired products including: 1,3-dihydro-3-methyl-5-phenyl-2H-1,4-benzodiazepin-2-one; 7-chloro-1,3-dihydro-3-methyl-5-phenyl-2H-1,4-benzodiazepin-2-one; 1,3-dihydro-5-(4-methoxyphenyl)-3-methyl-2H-1,4-benzodiazepin-2-one; 1,3-dihydro-3-methyl-7-nitro-5-phenyl-2H-1,4-benzodiazepin-2-one; 1,3-dihydro-1-isopropyl-3,6-dimethyl-2H-1,4-benzodiazepin-2-one; 7-nitro-1,3-dihydro-1,3-dimethyl-5-phenyl-2H-1,4-benzodiazepin-2-one; 5-cyclohexyl-1,3-dihydro-3-methyl-2H-1,4-benzodiazepin-2-one; 1,3-dihydro-3-methyl-5-(2-thienyl)-2H-1,4-benzodiazepin-2-one; 1,3-dihydro-5-phenyl-2H-1,4-benzodiazepin-2-one; 7-chloro-1,3-dihydro-5-phenyl-2H-1,4-benzodiazepin-2-one; 1,3-dihydro-5-(4-methoxyphenyl)-2H-1,4-benzodiazepin-2-one; 1,3-dihydro-7-nitro-5-phenyl-2H-1,4-benzodiazepin-2-one; 1,3-dihydro-1-isopropyl-6-methyl-2H-1,4-benzodiazepin-2-one; 7-nitro-1,3-dihydro-1-methyl-5-phenyl-2H-1,4-benzodiazepin-2-one; 5-cyclohexyl-1,3-dihydro-2H-1,4-benzodiazepin-2-one; 1,3-dihydro-5-(2-thienyl)-2H-1,4-benzodiazepin-2-one; 1,3-dihydro-3-(4-hydroxyphenyl)-5-phenyl-2H-1,4-benzodiazepin-2-one; 7-chloro-1,3-dihydro-3-(4-hydroxyphenyl)-5-phenyl-2H-1,4-benzodiazepin-2-one; 1,3-dihydro-5-(4-methoxyphenyl)-3-(4-hydroxyphenyl)-2H-1,4-benzodiazepin-2-one; 1,3-dihydro-7-nitro-3-(4-hydroxyphenyl)-5-phenyl-2H-1,4-benzodiazepin-2-one; 1,3-dihydro-1-isopropyl-6-methyl-3-(4-hydroxyphenyl)-2H-1,4-benzodiazepin-2-one; 7-nitro-1,3-dihydro-1-methyl-3-(4-hydroxyphenyl)-5-phenyl-2H-1,4-benzodiazepin-2-one;

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5-cyclohexyl-1,3-dihydro-3-(4-hydroxyphenyl)-2H-
 1,4-benzodiazepin-2-one; 1,3-dihydro-
 3-(4-hydroxyphenyl)-5-(2-thienyl)-2H-1,4-benzodiazepin-
 2-one; 1,3-dihydro-3-(1H-indol-2-ylmethyl)-5-phenyl-2H-
 1,4-benzodiazepin-2-one; 7-chloro-1,3-dihydro-3-(1H-
 indol-2-ylmethyl)-5-phenyl-2H-1,4-benzodiazepin-2-one;
 1,3-dihydro-3-(1H-indol-2-ylmethyl)-
 5-(4-methoxyphenyl)-2H-1,4-benzodiazepin-2-one;
 1,3-dihydro-3-(1H-indol-2-ylmethyl)-7-nitro-5-phenyl-
 2H-1,4-benzodiazepin-2-one; 1,3-dihydro-3-(1H-indol-
 2-ylmethyl)-1-isopropyl-6-methyl-2H-1,4-benzodiazepin-
 2-one; 7-nitro-1,3-dihydro-3-(1H-indol-2-ylmethyl)-
 1-methyl-5-phenyl-2H-1,4-benzodiazepin-2-one;
 5-cyclohexyl-1,3-dihydro-3-(1H-indol-2-ylmethyl)-2H-
 1,4-benzodiazepin-2-one; 1,3-dihydro-3-(1H-indol-
 2-ylmethyl)-5-(2-thienyl)-2H-1,4-benzodiazepin-2-one;
 1,3-dihydro-3-(1-methylethyl)-5-phenyl-2H-
 1,4-benzodiazepin-2-one; 7-chloro-1,3-dihydro-
 3-(1-methylethyl)-5-phenyl-2H-1,4-benzodiazepin-2-one;
 1,3-dihydro-5-(4-methoxyphenyl)-3-(1-methylethyl)-2H-
 1,4-benzodiazepin-2-one; 1,3-dihydro-3-(1-methylethyl)-
 7-nitro-5-phenyl-2H-1,4-benzodiazepin-2-one;
 1,3-dihydro-1-isopropyl-6-methyl-3-(1-methylethyl)-2H-
 1,4-benzodiazepin-2-one; 7-nitro-1,3-dihydro-1-methyl-
 3-(1-methylethyl)-5-phenyl-2H-1,4-benzodiazepin-2-one;
 5-cyclohexyl-1,3-dihydro-3-(1-methylethyl)-2H-
 1,4-benzodiazepin-2-one; 1,3-dihydro-3-(1-methylethyl)-
 5-(2-thienyl)-2H-1,4-benzodiazepin-2-one; are set forth
 in Table 3.

30

EXAMPLE 4

Synthesis of Quinolones

The synthesis of 36 quinolones is summarized in
 Scheme 4 and Table 4.

35

Thirty-six gas dispersion tubes are loaded each
 with 100 ± 5 mg of hydroxymethyl polystyrene resin (1%

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cross-linked, 1.0 meq/g, loading) and placed in the holder block. The apparatus is assembled, swelled with 3 mL of CH_2Cl_2 , and drained as set forth in Example 1.

5 To three sets of 12 reaction wells within the array is added 2 mL of a 0.2 M CH_2Cl_2 solution of either 3-(2,4,5-trifluorophenyl)-3-oxo-1-propanoic acid, 3-(2,3,4,5-tetrafluorophenyl)-3-oxo-1-propanoic acid, or 3-(2,4-dichloro-5-fluoropyridyl)-3-oxo-3-propanoic acid. An additional 1 mL of a CH_2Cl_2 stock solution
10 containing 0.4 M dicyclohexylcarbodiimide and 0.4 M 1-hydroxybenzotriazole is added to all 36 reaction wells of the array and the appropriate reaction tubes are submerged in the corresponding reaction wells. The apparatus is sealed as set forth in Example 1 and
15 agitated in a sonic bath for 12 hours, while maintaining a positive chilled nitrogen flow through the manifold as set forth in Example 2. At the end of the reaction, the reaction tube array is drained and sparged as set forth in Example 1.

20 The reaction tube array is then washed using CH_2Cl_2 (10 times) as set forth in Example 3.

Into every well of the array is added 3 mL of a solution of acetic anhydride/triethyl orthoformate (1:1 v/v) and the apparatus is sealed as set forth in
25 Example 1. The apparatus is then heated at 150°C (oil bath temp.) for 3 hours, while maintaining a positive chilled nitrogen flow through the manifold as set forth in Example 2. At the end of the reaction, the reaction tube array is drained and sparged as set forth in
30 Example 1.

The reaction tubes are washed as set forth in Example 3, using CH_2Cl_2 (10 times, 3 mL.)

To three sets of 12 reaction wells within the array is added 3 mL of a t-butanol solution 0.13 M in both
35 ethyl amine and t-butoxide, or cyclopropyl amine and t-butoxide, or 2,4-difluoroaniline and t-butoxide. The

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apparatus is reassembled as set forth in Example 1, immersed in an oil bath (60°C) and heated for 12 hours as set forth in Example 3. At the end of the reaction, the reaction tube array is allowed to cool and then
5 drained and sparged as set forth in Example 1.

The reaction tubes are then washed as set forth in Example 3, using CH_2Cl_2 (12 times, 3 mL.)

To four sets of nine reaction wells within the array is added 3 mL of a 0.26 M acetonitrile solution
10 of either piperazine, piperidine, N-methyl piperazine, or 3-(ethylaminomethyl)pyrrolidine. The apparatus is assembled as set forth in Example 1, and heated in an oil bath (80°C) for 4 hours as set forth in Example 3. At the end of the reaction, the reaction tube array is
15 allowed to cool and then drained and sparged as set forth in Example 1.

The reaction tubes are then washed as set forth in Example 3, using CH_2Cl_2 (12 times, 3 mL.)

The reaction tubes are submerged in reaction wells
20 containing 3 mL of 2N NaOH in 1:1 dioxane:water. The apparatus is assembled as set forth in Example 1, and allowed to stand at room temperature for 6 hours to affect ester hydrolysis. At the end of the reaction, the reaction tube array is drained and sparged as set
25 forth in Example 1.

The reaction tubes are then washed as set forth in Example 3, using 3 x 3 mL of 0.1N NaOH in dioxane:water (1:1). The filtrates are then combined and the dioxane is removed by evaporation as set forth in Example 3.
30 The resulting aqueous solutions are acidified with 5 N HCl solution and evaporated to give the 36 individual expected crude quinolones.

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EXAMPLE 5

Synthesis of Keto-ureas

The synthesis of eight keto-ureas is summarized in Scheme 5 and Table 5.

5 100 g of Merrifield's chloromethyl polystyrene resin (1-3% cross-linked, loading = 0.78 meq/g, 200-400 mesh) is heated for 20 hours at 150-155°C in 500 mL of DMSO with 5 eq of NaHCO₃ to form the resin-bound benzaldehyde. The resin is filtered and washed
10 sequentially with excess DMSO, hot water, dioxane:water (2:1), water, dioxane, acetone, ethanol, CH₂Cl₂, and benzene. The dried aldehyde resin is then reacted with O-methylhydroxylamine hydrochloride (7.5 eq) and
15 pyridine (7.8 eq) in refluxing ethanol to form the resin-bound oxime, which is washed with excess MeOH and CH₂Cl₂, and then directly reduced with sodium cyanoborohydride (10 eq) in EtOH saturated with HCl at room temperature for 3 hours using methyl orange as an
20 indicator and adjusting the pH to 3.1 with additional EtOH/HCl. The resin is washed sequentially with excess MeOH, hot water, dioxane:water (1:1), dioxane, and CH₂Cl₂ and used directly below.

25 100 mg of O-Methyl hydroxylamine resin from above is measured into each of the eight gas dispersion tubes (reaction tubes). The apparatus is assembled, swelled with 3 mL of CH₂Cl₂, and drained as set forth in Example 1.

30 To facilitate coupling of the Boc-amino benzoic acids, the appropriate reaction tubes are submerged in reaction wells containing 3 mL of a solution of 0.08 M diisopropylethylamine, 0.16 M BOP (Benzotriazol-1-yloxy-tris-(dimethylamino)-phosphonium hexafluorophosphate), and 0.08 M Boc-amino benzoic acid (either
35 3-amino-4-chlorobenzoic acid or 4-amino-3-hydroxybenzoic acid) in CH₂Cl₂. The apparatus is sealed as set forth in Example 1 and agitated in a

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sonic bath for 8 hours while maintaining a positive nitrogen flow through the manifold. At the end of the reaction, the reaction tube array is drained and sparged as set forth in Example 1.

5 The reaction tube array is subjected to the standard wash cycle set forth in Example 1.

 The appropriate reaction tubes are submerged in reaction wells containing a solution of 4 mL of sat. HCl-Ether:dioxane (2:1). The apparatus is sealed as
10 set forth in Example 1 and agitated in a sonic bath for 2 hours, while maintaining a positive nitrogen flow through the manifold. At the end of the reaction, the reaction tube array is drained and sparged as set forth in Example 1.

15 The reaction tube array is subjected to the standard wash cycle set forth in Example 1.

 The appropriate reaction tubes are submerged in reaction wells containing a solution of 3 mL of 0.16 M phenyl isocyanate or ethyl isocyanate in dioxane with
20 an internal standard (e.g., anthracene at 2-4 mg/mL). The apparatus is sealed as set forth in Example 1 and agitated in a sonic bath for 6 hours, while maintaining a positive nitrogen flow through the manifold. The reaction progress is monitored by GC/ISTD as set forth
25 in Example 1. At the end of the reaction, the reaction tube array is drained and sparged as set forth in Example 1.

 The reaction tube array is subjected to the standard wash cycle set forth in Example 1.

30 In order to cleave the product from the solid support, the reaction tubes are submerged in reaction wells containing a solution of 3 mL of dry CH_2Cl_2 at -78°C . The apparatus is sealed as set forth in Example 1 and placed in an isopropanol/dry ice bath
35 while maintaining a positive chilled nitrogen flow through the manifold. A solution of either

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methylmagnesium bromide 1.4 M in toluene/tetrahydrofuran or phenylmagnesium bromide 1.0 M in tetrahydrofuran (0.2 mL) is added to the top of the appropriate reaction tubes through the gasket at the top of the manifold and the apparatus is sonicated at -78°C (maintained at -10°C by circulating chilled fluid through the sonic bath). After 2 hours, 10% HCl (0.5 mL) is added to each reaction tube through the gasket at the top of the manifold and the apparatus is allowed to warm to room temperature. At the end of the reaction, the reaction tube array is drained and sparged as set forth in Example 1.

To isolate and purify the products, the reaction tubes are washed as set forth in Example 1, using 3 mL portions of methanol. The methanol extraction protocol is repeated until the filtrates are free of any organic components as determined by TLC (1-2 times). The appropriate HCl and methanol filtrates are then combined and concentrated on a Speed-Vac® in tared vials to afford eight discrete keto-ureas.

EXAMPLE 6

Synthesis of N²-Substituted Hydantoins

The synthesis of eight N²-substituted hydantoins is summarized in Scheme 6 and Table 6.

100 mg each of the commercially available Fmoc-protected amino acids on Wang resins (Fmoc-Alanine and Fmoc-Phenylalanine, loading = 0.37-0.60 meq/g, 200-400 mesh) are measured into two sets of four gas dispersion tubes (reaction tubes). The apparatus is assembled, swelled with 3 mL of DMF, and drained as set forth in Example 1.

To deprotect the Fmoc amino acids, the appropriate reaction tubes are submerged in reaction wells containing a solution of 4 mL of 25% piperidine in DMF (v/v) containing an internal standard (anthracene at

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2-4 mg/mL). The apparatus is sealed as set forth in Example 1 and agitated in a sonic bath while maintaining a positive nitrogen flow through the manifold. The reaction progress is monitored by removing a sample of the filtrate (e.g., 10-100 μ L) and analyzing for the Fmoc-piperidine adduct and dibenzofulvene by GC/ISTD calibration methods. At the end of the reaction, the reaction tube array is drained and sparged as set forth in Example 1.

The reaction tube array is subjected to the standard wash cycle set forth in Example 1.

The appropriate reaction tubes are submerged in reaction wells containing a solution of 4 mL of 0.10 to 0.30 M acetaldehyde or benzaldehyde in dioxane containing an internal standard (e.g., anthracene at 2-4 mg/mL). The apparatus is sealed and agitated in a sonic bath for 12 hours, while maintaining a positive nitrogen flow through the manifold. The reaction progress is monitored by GC/ISTD calibration methods as set forth in Example 1. At the end of the reaction, the reaction tube array is drained and sparged as set forth in Example 1.

The reaction tube array is subjected to the standard wash cycle set forth in Example 1.

The appropriate reaction tubes are submerged in reaction wells containing a solution of 4 mL of 0.10-0.30 M sodium cyanoborohydride in methanol with an indicator (e.g., methyl orange at 2-4 mg/mL). The apparatus is sealed as set forth in Example 1 and agitated in a sonic bath while maintaining a positive nitrogen flow through the manifold. The reaction progress is monitored by visually observing the color of the filtrates. When neutral or basic conditions are observed, the pH is adjusted by injection of dilute aqueous HCl into the appropriate reaction tubes through the gasket at the top of the manifold. Stabilization

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of pH changes indicates completion of the reduction reaction. At the end of the reaction, the reaction tube array is drained and sparged as set forth in Example 1.

5 The reaction tube array is subjected to the standard wash cycle set forth in Example 1.

 The appropriate reaction tubes are submerged in reaction wells containing a solution of 4 mL of 0.10-0.30 M trimethylsilyl isocyanate or allyl
10 isocyanate in DMF containing an internal standard (e.g., anthracene at 2-4 mg/mL). The apparatus is sealed as set forth in Example 1 and agitated in a sonic bath for 6 hours, while maintaining a positive nitrogen flow through the manifold. The reaction
15 progress is monitored by removing a sample of the filtrate (10-100 μ L), derivatization with an appropriate amine or alcohol, and analysis by GC/ISTD calibration methods as set forth in Example 1. At the
20 end of the reaction, the reaction tube array is drained and sparged as set forth in Example 1.

 The reaction tube array is subjected to the standard wash cycle set forth in Example 1.

 In order to cleave the product from the solid support, the reaction tubes are submerged in reaction
25 wells containing a solution of 4 mL of aqueous 6N HCl. The apparatus is sealed as set forth in Example 1, placed in an oil bath and heated at 95 to 100°C for 2 hours while maintaining a positive chilled nitrogen flow through the manifold as set forth in Example 2.
30 At the end of the reaction, the reaction tube array is drained and sparged as set forth in Example 1.

 To isolate and purify the products, the reaction tubes are washed as set forth in Example 1 using 4 mL portions of methanol. The methanol extraction protocol
35 is repeated until the filtrates are free of any organic components as determined by TLC (1-2 times). The

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appropriate HCl and methanol filtrates are then combined and concentrated on a Speed-Vac® in tared vials to afford eight discrete hydantoins.

5

EXAMPLE 7

Synthesis of (R)-4-Benzamido-5-oxopentanoic Acids

The synthesis of 12 oxopentanoic acids is summarized in Scheme 7 and Table 7.

5 g of α -2-Trimethylsilylethyl-N-BOC-glutamic acid
10 is linked through the γ -carboxyl group to commercially available p-benzyloxybenzyl alcohol resin (loading = 1.2 meq/g, 200-400 mesh) using N,N'-diisopropylcarbodiimide (4 eq) and HOBT (2 eq) in 3 mL of DMF for 24 hours. The resulting resin is then washed with
15 excess MeOH and CH₂Cl₂ and used directly below.

95 to 105 mg of the above formed resin is measured into each of 12 gas dispersion tubes (reaction tubes). The apparatus is assembled, swelled with 3 mL of dioxane, and drained as set forth in Example 1.

20 The reaction tubes are submerged in reaction wells containing a solution of tetrabutylammonium fluoride (5 eq) in dioxane (3 mL) at room temperature. The apparatus is sealed as set forth in Example 1 and agitated at room temperature for 8 hours in a sonic
25 bath as set forth in Example 5, while maintaining a positive nitrogen flow through the manifold. At the end of the reaction, the reaction tube array is drained and sparged as set forth in Example 1.

The reaction tubes are washed as set forth in
30 Example 3, using 2 x 3 mL each of the following solutions, dioxane:H₂O (1:1), dioxane and CH₂Cl₂.

The reaction tubes are submerged in reaction wells containing a solution of ethylchloroformate (3 eq) and triethylamine (3 eq) in dioxane (3 mL) at -10°C. The
35 apparatus is sealed as set forth in Example 1 and agitated in a sonic bath at room temperature for

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4 hours as set forth in Example 5, while maintaining a positive nitrogen flow through the manifold. At the end of the reaction, the reaction tube array is allowed to warm to room temperature and then drained and sparged as set forth in Example 1.

The reaction tubes are washed as set forth in Example 3, using 4 x 2 mL of anhydrous DMF. Because of the labile nature of the above formed acyl-carbonates, the reaction tubes are rapidly washed in order to minimize unwanted hydrolysis.

The appropriate reaction tubes are submerged in reaction wells containing a solution of 4 eq of either n-butylamine, cycloheptylamine, 8-azaspiro[4.5]decane, or dipentylamine in dioxane (4 mL) at -10°C. The apparatus is sealed as set forth in Example 1 and agitated in a sonic bath at -10°C as set forth in Example 5 for 1 hour and then 3 hours at room temperature, while maintaining a positive nitrogen flow through the manifold. At the end of the reaction, the reaction tube array is drained and sparged as set forth in Example 1.

The reaction tubes are washed as set forth in Example 3, using 2 x 3 mL each of the following solutions, dioxane:H₂O (1:1), dioxane:1N HCl (1:1), dioxane:H₂O (1:1), dioxane:0.1N NaOH (1:1), dioxane:H₂O (1:1), and dioxane.

The reaction tubes are submerged in reaction wells containing a solution of 20% piperidine in DMF at room temperature. The apparatus is sealed as set forth in Example 1 and agitated at room temperature in a sonic bath for 6 hours as set forth in Example 5, while maintaining a positive nitrogen flow through the manifold. At the end of the reaction, the reaction tube array is drained and sparged as set forth in Example 1.

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The reaction tubes are washed as set forth in Example 3, using 2 x 3 mL each of the following solutions, DMF, dioxane:H₂O (1:1), dioxane and CH₂Cl₂.

5 The appropriate reaction tubes are submerged in reaction wells containing a solution of 2 eq of either benzoyl chloride, 3-methoxybenzoyl chloride, or 3-nitrobenzoyl chloride in pyridine (3 mL) at 0°C. The apparatus is sealed as set forth in Example 1 and agitated in a sonic bath at 0°C for 3 hours as set
10 forth in Example 5, while maintaining a positive nitrogen flow through the manifold. At the end of the reaction, the apparatus is allowed to warm to room temperature, and the reaction tubes are drained and sparged as set forth in Example 1.

15 The reaction tubes are washed as set forth in Example 3, using 3 x 2 mL each of the following solutions, dioxane, dioxane:H₂O (1:1), and dioxane.

In order to cleave the product from the solid support, the reaction tubes are submerged in reaction
20 wells containing 3 mL each of trifluoroacetic acid. The apparatus is sealed as set forth in Example 1 and agitated in a sonic bath at 0°C for 2 hours and then at room temperature for 4 hours as set forth in Example 5, while maintaining a positive nitrogen flow through the
25 manifold. After 6 hours in a sonic bath, the apparatus is allowed to warm to room temperature, and the reaction tubes are drained and sparged as set forth in Example 1.

The reaction tubes are washed as set forth in
30 Example 3, using 2 x 3 mL of methylene chloride. The filtrates are then combined and concentrated as set forth in Example 3. The crude products are then redissolved, transferred to tared vials, and reconcentrated to yield the final crude products.

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EXAMPLE 8

Synthesis of Diketopiperazines

The synthesis of 40 diketopiperazines is summarized in Scheme 8 and Table 8.

5 95 to 105 mg of Merrifield's resin (loading = 0.66 meq/g, 200-400 mesh) is measured into each of 40 gas dispersion tubes (reaction tubes) containing a magnetic stirring bar. The apparatus is assembled, swelled with 3 mL of DMF, and drained as set forth in
10 Example 1.

The reaction tubes are submerged into wells containing a solution of 75.0 mg BOC 4-hydroxyproline cesium salt in 5 mL of DMF. The apparatus is assembled and sealed with clamps as set forth in Example 1, and
15 placed into an oil bath over a magnetic stirring plate while maintaining a positive chilled nitrogen flow through the manifold as set forth in Example 2 and reacted at 50 °C. After 24 hours, the reaction tube array is drained and sparged as set forth in Example 1.

20 The reaction tube array is subjected to the standard wash cycle set forth in Example 1, ending however in CH_2Cl_2 , to remove residual reagents and byproducts.

The appropriate reaction tubes are submerged in
25 reaction wells containing a solution of 5 mL of 0.07 M acyl halide (either, benzoyl bromide, acetyl chloride, 4-biphenylcarbonyl chloride, p-anisoyl chloride, 4-chlorobenzoyl chloride, 4-nitrobenzoyl chloride, pivaloyl chloride, or trifluoroacetyl chloride) and
30 0.07 M triethylamine in CH_2Cl_2 . The apparatus is sealed as set forth in Example 1 and agitated as above while maintaining a positive nitrogen flow through the manifold. After 72 hours, the reaction tube array is drained and sparged as set forth in Example 1, using
35 DMF, methanol, and CH_2Cl_2 .

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5 The reaction tubes are submerged in 5 mL of 0.08 M TFA in CH_2Cl_2 to remove the BOC protecting group from N_1 . The apparatus is assembled and the reactions stirred as above. After 72 hours, the reaction tubes are drained and washed with DMF, methanol, 1.0 M TFA in CH_2Cl_2 , and CH_2Cl_2 as set forth in Example 1.

10 A solution (5 mL) of 0.07 M BOC amino acid, 0.07 M diisopropylcarbodiimide and 0.07 M TFA in CH_2Cl_2 are stirred for 3 hours at room temperature. The appropriate reaction tubes are then submerged in the above reaction wells, and the apparatus is assembled as set forth in Example 1. Five BOC amino acids are employed, BOC-glycine, BOC-alanine, BOC-valine, BOC-phenylalanine, and BOC-diphenylglycine. After 15 72 hours, the reaction tube array is drained and sparged as set forth in Example 1.

The reaction tube array is subjected to the standard wash cycle set forth in Example 1, ending however in CH_2Cl_2 .

20 Cyclization to the final product and thus cleavage from the resin is accomplished by removal of the BOC (t-butyloxycarbonyl) group of the amino acid. The reaction tubes are submerged in reaction wells containing a solution of 5 mL of saturated HCl in CH_2Cl_2 . The apparatus is assembled as set forth in Example 1 and the reactions stirred as above at room temperature. After 24 hours, the reaction tube array is drained and sparged as set forth in Example 1. The contents in the reaction wells are concentrated to 25 dryness using a stream of nitrogen as set forth in Example 3.

30 A solution of 5 mL of 1.0 M triethylamine in CH_2Cl_2 is added to each reaction well and the reaction tubes are then submerged in these wells. The apparatus is assembled as set forth in Example 1, and the reactions 35

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stirred as above. After 24 hours, the reaction tube array is drained and sparged as set forth in Example 1.

5 The contents in the reaction wells are concentrated to dryness using a stream of nitrogen as set forth in Example 3. These crude products are partitioned between CH_2Cl_2 and a saturated solution of NaHCO_3 using a Tecan® robot to deliver the necessary liquids and to remove the aqueous layer. The basic aqueous wash is repeated and the residual CH_2Cl_2 layers are dried by
10 passing the samples through a bed of MgSO_4 contained in standard SPE cartridges employing the vacuum SPE apparatus described above, giving the crude diketopiperazines.

15

EXAMPLE 9

Synthesis of Tetrahydro-4-hydroxy-6-[2-(1H-pyrrol-1yl)ethyl]-2H-pyran-2-ones

The synthesis of three 2H-pyranones is summarized in Scheme 9 and Table 9.

20 Three dispersion tubes are loaded each with 100 ± 5 mg of hydroxymethyl polystyrene resin (1% cross-linked, 1.0 meq/g, loading) and placed in the holder block. The apparatus is assembled, swelled with 3 mL of acetonitrile, and drained as set forth in Example 1.

25 The appropriate reaction tubes are submerged in reaction wells containing 2 mL of a solution of either 0.2 M acetic anhydride, propanoic anhydride, or 2-phenylacetic anhydride in acetonitrile, and 1.0 mL of a 0.4 M solution of triethylamine in CH_2Cl_2 . The
30 apparatus is sealed as set forth in Example 1, and agitated in a sonic bath while maintaining a positive nitrogen flow through the manifold for 12 hours. At the end of the reaction, the reaction tube array is drained and sparged as set forth in Example 1.

35 The reaction tubes are washed as set forth in Example 3, using 10 x 4 mL of CH_2Cl_2 .

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The reaction tubes are submerged in reaction wells containing 3 mL of a solution of 0.2 M lithium diisopropylamide in THF and the apparatus is sealed as set forth in Example 1 and agitated in a sonic bath at room temperature as set forth in Example 5, while maintaining a positive nitrogen flow through the manifold for 4 hours.

1 mL of a solution (in THF) of either 1.0 M 5-[2-(4-fluorophenyl)-3-phenyl-4-carboxamidophenyl-5-isopropyl-1-pyrrololyl]-3-oxo-1-pentanoic acid methyl ester (to wells #1 and #2), or 1.0 M 5-[2,3,4-triphenyl-5-methyl-1-pyrroloyl]-3-oxo-1-pentanoic acid methyl ester (to well #3) is added to the top of the appropriate reaction tubes through the gasket at the top of the manifold and sonicated at room temperature for 12 hours as set forth in Example 5. At the end of the reaction, the reaction tube array is drained and sparged as set forth in Example 1.

The reaction tubes are washed as set forth in Example 3, using 10 x 4 mL of CH_2Cl_2 .

The reaction tubes are submerged in reaction wells containing 2.0 mL of a 0.26 M THF solution of diethyl methoxy borane and the apparatus is sealed as set forth in Example 1. 1.0 mL of a 1.0 M THF solution of sodium borohydride is added to the top of the appropriate reaction tubes through the gasket at the top of the manifold and the apparatus is sonicated at room temperature for 4 hours as set forth in Example 5. At the end of the reaction, the reaction tube array is drained and sparged as set forth in Example 1.

The reaction tubes are washed as set forth in Example 3, using 10 x 4 mL of CH_2Cl_2 .

The reaction tubes are submerged in reaction wells containing 3.0 mL of 3% hydrogen peroxide in 1:1 dioxane:water and the apparatus is assembled as set forth in Example 1, and sonicated at room temperature.

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for 6 hours as set forth in Example 5 to affect boron oxidation. At the end of the reaction, the reaction tube array is drained and sparged as set forth in Example 1.

5 The reaction tubes are washed as set forth in Example 3, using 10 x 3 mL of dioxane:water (1:1).

10 The reaction tubes are submerged in reaction wells containing a solution of 0.1 M HCl in 1:1 dioxane:water (3 mL). The apparatus is assembled as set forth in Example 1, and sonicated at room temperature for 6 hours as set forth in Example 5 to affect hydrolysis/ring closure. At the end of the reaction, the reaction tube array is drained and sparged as set forth in Example 1.

15 The reaction tubes are washed as set forth in Example 3, using 3 x 3 mL of dioxane:water (1:1). The appropriate filtrates are then combined and concentrated as set forth in Example 3 to give the three individual expected lactone products.

20

EXAMPLE 10

Synthesis of N-Arylpiperazines

The synthesis of 12 N-arylpiperazines is summarized in Scheme 10 and Table 10.

25 100 mg of hydroxymethyl polystyrene (2% cross-linked, 1.2 meq/g, loading) resin is measured into each of 12 gas dispersion tubes (reaction tubes). The apparatus is assembled, swelled with 3 mL of DMF, and drained as set forth in Example 1.

30 To facilitate coupling of the acid, the appropriate reaction tubes are submerged in reaction wells containing a solution of 0.09 M 4-chlorobutyric acid or 3-chloropropionic acid; and 0.09 M N,N'-diisopropylcarbodiimide and 0.09 M 4,4-dimethylaminopyridine in
35 DMF (3 mL) at room temperature. The apparatus is sealed as set forth in Example 1 and agitated in a

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sonic bath for 2 hours, while maintaining a positive nitrogen flow through the manifold. At the end of the reaction, the reaction tube array is drained and sparged as set forth in Example 1.

5 The reaction tube array is subjected to the standard wash cycle set forth in Example 1.

10 The appropriate reaction tubes are submerged in reaction wells containing a solution of 3 mL of either 0.09 M piperazine, phenylpiperazine, or 1-(2-pyridyl) piperazine; 0.18 M potassium carbonate; and 0.18 M potassium iodide in DMF. The apparatus is sealed as set forth in Example 1, placed in an oil bath, and heated at 95 to 100°C while maintaining a positive chilled nitrogen flow through the manifold as set forth in Example 2. After 6 hours, the reaction is stopped and the reaction tube array is drained and sparged as set forth in Example 1.

15 The reaction tube array is subjected to the standard wash cycle set forth in Example 1.

20 The appropriate reaction tubes are submerged in reaction wells containing a solution of 3 mL of toluene-20% 1,2-dimethoxyethane at 0°C. The apparatus is sealed as set forth in Example 1 and agitated in a sonic bath at 0°C as set forth in Example 5, while maintaining a positive argon flow.

25 To form the enolate, 0.5 mL of a solution (in THF) of 0.15 M trityllithium is added to the top of the appropriate reaction tubes through the gasket at the top of the manifold and sonicated at 0°C for 15 minutes as set forth in Example 5.

30 After 15 minutes, 0.5 mL of 0.28 M acid chloride (either nicotinoyl chloride or naphthoyl chloride) in THF is added to the top of the appropriate reaction tubes through the gasket at the top of the manifold and sonicated at room temperature for 1 hour as set forth in Example 5. At the end of the reaction, the

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reaction tube array is drained and sparged as set forth in Example 1.

The reaction tube array is subjected to the standard wash cycle set forth in Example 1.

5 In order to cleave the product from the solid support, the reaction tubes are submerged in reaction wells containing a solution of 3 mL of dry HBr in TFA. The apparatus is sealed as set forth in Example 1 and agitated in a sonic bath while maintaining a positive
10 nitrogen flow through the manifold. After 1 hour, the reaction is stopped, and the reaction tube array is drained and sparged as set forth in Example 1.

To isolate and purify the products, the reaction tubes are washed as set forth in Example 1, using 3 mL
15 portions of methanol. The methanol extraction protocol is repeated until the filtrates are free of any organic components as determined by TLC (1-2 times). The appropriate HCl and methanol filtrates are then combined and concentrated on a Speed-Vac® in tared
20 vials to afford 12 discrete products.

In an example of a post-cleavage reaction, a modified Wolff-Kishner reduction is done on half of the amount of each of the isolated compounds. The compounds are placed in the appropriate reaction wells
25 containing 3 mL of 0.09 M potassium hydroxide, and 0.09 M hydrazine hydrate in diethylene glycol. The reaction tubes are filled with 0.5 g sodium sulfate (to absorb water) and submerged in the reaction wells. The apparatus is sealed as set forth in Example 1, placed
30 in an oil bath, and heated at 80°C while maintaining a positive chilled nitrogen flow through the manifold as set forth in Example 2. After 12 hours, the reaction is stopped and the reaction tube array is drained and sparged as set forth in Example 1. Each product is
35 isolated by adding 4 mL of a solution of 1:1 water:CH₂Cl₂. Each organic layer is separated, dried,

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and concentrated as set forth in Example 8 to yield another 12 desired products.

EXAMPLE 11

5 Synthesis of Benzisothiazolones

The synthesis of nine benzisothiazolones is summarized in Scheme 11 and Table 11.

95 to 105 mg of Merrifield's resin (loading = 0.66 meq/g, 200-400 mesh) is measured into each of nine
10 gas dispersion tubes (reaction tubes). The apparatus is assembled, swelled with 3 mL of dioxane and drained as set forth in Example 1.

The appropriate reaction tubes are submerged in reaction wells containing a solution of 5 mL of either
15 0.2 M thiosalicylic acid, 5-chloro-thiosalicylic acid, or 2-mercaptonicotinic acid; and 0.4 M triethylamine in dioxane and magnetic stirring bars. The apparatus is sealed with clamps as set forth in Example 1 and placed over a magnetic stirring plate, while maintaining a
20 positive nitrogen flow through the manifold. After 96 hours, the reaction is stopped and the reaction tube array is drained and sparged as set forth in Example 1.

The reaction tubes are washed as set forth in Example 3, using 3 x 3 mL each of water, 1N HCl, 1N
25 HCL:dioxane (1:1), and finally dioxane to remove residual reagents and byproducts.

The reaction tubes are submerged in reaction wells containing a solution of 5 mL of 0.20 M carbonyldiimidazole in dioxane. The apparatus is assembled and
30 agitated as above. After 5 hours, the reaction is stopped and the reaction tube array is drained and sparged as set forth in Example 1.

The reaction tubes are washed as set forth in Example 3, using 4 x 3 mL each of DMF then dioxane.

35 The appropriate reaction tubes are submerged in reaction wells containing a solution of 5 mL of either

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0.20 M 4-methoxyaniline, cyclohexylamine, or benzylamine in dioxane. The apparatus is assembled as set forth in Example 1, while maintaining a flow of cold nitrogen. The apparatus is placed in an oil bath over a magnetic stirring plate and the reactions are warmed to reflux and stirred as above. After 24 hours, the reaction is stopped and the reaction tube array is drained and sparged as set forth in Example 1.

The reaction tubes are washed as set forth in Example 3, using 4 x 3 mL each of DMF, methanol, CH₂Cl₂, and then dioxane.

The reaction tubes are submerged in reaction wells containing a solution of 5 mL of 0.13 M NaBrO₂ in dioxane/water (8:1). The apparatus is assembled as set forth in Example 1, while maintaining a flow of nitrogen. The apparatus is placed over a magnetic stirring plate and stirred at room temperature as above. After 1 hour, the reaction is stopped and the reaction tube array is drained and sparged as set forth in Example 1.

The reaction tubes are washed as set forth in Example 3, using 4 x 3 mL each of dioxane, water, dioxane/water (1:1), methanol, and then CH₂Cl₂.

Cyclization to the final product and thus cleavage from the resin is accomplished by addition of trichloroacetic anhydride to the sulfoxide and rearrangement as taught by Wright, et al, Tetrahedron Letters 1991;33:153. Thus, reaction tubes are submerged in reaction wells containing a solution of 5 mL of CH₂Cl₂. The apparatus is assembled as set forth in Example 1, while maintaining a flow of nitrogen. The apparatus is then placed in a cold bath at 0°C and the reactions stirred as above. Trichloroacetic anhydride (0.18 mL) is then added to the top of each reaction tube through the gasket at the top of the manifold and the temperature of the reaction is allowed

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to warm to 25°C over 7 hours. At the end of the reaction, the reaction tube array is drained and sparged as set forth in Example 1.

5 The reaction tubes are washed as set forth in Example 3, using 2 x 3 mL each of CH₂Cl₂, dioxane, MeOH, and CH₂Cl₂.

10 The appropriate filtrates are then combined and concentrated as set forth in Example 3. These crude products are partitioned between CH₂Cl₂ and 2N NaOH using a Tecan® robot to deliver the necessary liquids and to remove the aqueous layer. The basic aqueous wash is repeated and the residual CH₂Cl₂ layers are dried by passing the samples through a bed of MgSO₄ contained in standard SPE cartridges employing the vacuum SPE apparatus described above to yield the crude
15 benzisothiazolones.

EXAMPLE 12

Synthesis of Isoindolone-Based Spirosuccinimides

20 The synthesis of nine spirosuccinimides is summarized in Scheme 12 and Table 12.

A solution of 0.10 mol cyanoacetic acid and 0.10 mol carbonyldiimidazole are combined in 500 mL of dioxane. After 5 hours, 10 g of benzylhydroxy
25 polystyrene resin (loading = 1.08 meq/g, 200 400 mesh) is added. The reaction is refluxed for 8 hours, cooled to room temperature, filtered, and washed with DMF, methanol, dioxane, and finally CH₂Cl₂. The product resin, benzyl cyanoacetate polystyrene resin, is used
30 directly in the next step.

95 of 105 mg of the resin prepared above (loading = 1.08 meq/g, 200-400 mesh) is measured into each of nine gas dispersion tubes (reaction tubes). The apparatus is assembled, swelled with 3 mL of DMF and drained as
35 set forth in Example 1.

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The appropriate reaction tubes are submerged in reaction wells containing a solution of 5 mL of either 0.20 M 3-imino-1-oxoisindoline, 6-chloro-3-imino-1-oxoisindoline, or 6-phenyl-1-oxoisindoline in diglyme with stirring magnetic bars. The 3-imino-1-oxoisindoline employed are prepared by the method of Wrobel, et al, Journal of Medicinal Chemistry 1992;35:4613. The apparatus is sealed with clamps as set forth in Example 1, and placed in an oil bath over a magnetic stirring plate while maintaining a positive flow of cold nitrogen as set forth in Example 2. The reaction vessels are warmed to reflux, and after 2 hours the reaction is stopped, and the reaction tube array is drained and sparged as set forth in Example 1.

The reaction tubes are washed as set forth in Example 3, using 3 x 3 mL each of dioxane, CH_2Cl_2 , and DMF.

The appropriate reaction tubes are submerged in reaction wells containing a solution of 0.20 M alkyl halide (either methyl iodide, benzyl bromide and 3-bromobenzyl bromide) and 1 mmol K_2CO_3 in 5 mL of DMF. The apparatus is assembled as set forth in Example 1, while maintaining a positive flow of cold nitrogen as set forth in Example 2. The oil bath is then warmed to 100°C and agitated as above. After 24 hours, the reaction is stopped and the reaction tube array is drained and sparged as set forth in Example 1.

The reaction tubes are washed as set forth in Example 3, using 3 x 3 mL each of DMF, water, MeOH, dioxane, CH_2Cl_2 , and DMSO.

The reaction tubes are submerged in reaction wells containing a solution of 5 mL of 0.20 M potassium cyanide in DMSO. The apparatus is assembled as set forth in Example 1 and agitated as above. After 24 hours at 25°C, the reaction is stopped and the

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reaction tube array is drained and sparged as set forth in Example 1.

5 The reaction tubes are washed as set forth in Example 3, using 3 x 3 mL each of DMF, water, MeOH, water, and 10% aqueous HCl.

10 The reaction tubes are submerged in reaction wells containing a solution of saturated dry HCl in MeOH. The apparatus is assembled as set forth in Example 1 and agitated as above. After 3 days at 25°C, the reaction is stopped and the reaction tube array is drained and sparged as set forth in Example 1.

15 The reaction tubes are washed as set forth in Example 3, using 3 x 3 mL each of MeOH, dioxane, and CH₂Cl₂.

The appropriate filtrates are then combined and concentrated as set forth in Example 3.

20 The crude products are dissolved in glacial HOAc and a holder block with nine clean empty reactions tubes is attached. The apparatus is assembled as set forth in Example 1 and agitated as above. After 24 hours at 25°C, the reaction is stopped and the reaction tube array is drained and sparged as set forth in Example 1.

25 The reaction tubes are washed as set forth in Example 3, using 3 x 3 mL of MeOH.

30 The appropriate filtrates are then combined and concentrated as set forth in Example 3 to yield the crude spiro[1H-isoindole-1,3'-pyrrolidine]-2',3,5' (2H)-triones.

EXAMPLE 13

Synthesis of Pilocarpine Analogs

The synthesis of four pilocarpine analogs is summarized in Scheme 13 and Table 13.

35 95 to 105 mg of 2-Chloro-triphenylmethyl resin as taught by Barlos, et al, Tetrahedron Letters

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1989;30:3947 is measured into each of four gas dispersion tubes (reaction tubes). The apparatus is assembled, swelled with 3 mL of DMF, and drained as set forth in Example 1.

5 Fmoc-histidine methyl ester is coupled to the resin. The reaction tubes are submerged in reaction wells containing a solution of 0.2 M Fmoc-histidine methyl ester and 0.4 M pyridine in 3 mL dry THF. The apparatus is sealed as set forth in Example 1 and
10 agitated in a sonic bath for 6 hours, while maintaining a positive nitrogen flow through the manifold. At the end of the reaction, the reaction tube array is drained and sparged as set forth in Example 1.

15 The reaction tube array is subjected to the standard wash cycle set forth in Example 1.

To deprotect the Fmoc amino acids, reaction tubes are submerged in reaction wells containing 3 mL of a solution of 25% piperidine in DMF. The apparatus is sealed as set forth in Example 1 and placed in a sonic
20 bath, while maintaining a positive nitrogen flow through the manifold. The reaction is stopped after 4 hours. At the end of the reaction, the reaction tube array is drained and sparged as set forth in Example 1.

25 The reaction tube array is subjected to the standard wash cycle set forth in Example 1.

The appropriate reaction tubes are submerged in reaction wells containing a solution of 0.2 M anhydride (either acetic anhydride or isobutyric anhydride) and 0.4 M triethylamine in 3 mL of dry THF. The apparatus
30 is sealed as set forth in Example 1 and agitated in a sonic bath for 12 hours, while maintaining a positive nitrogen flow through the manifold. At the end of the reaction, the reaction tube array is drained and sparged as set forth in Example 1.

35 The reaction tube array is subjected to the standard wash cycle set forth in Example 1.

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The apparatus is sealed as set forth in Example 1 while maintaining a positive nitrogen flow through the manifold. A solution of lithium aluminum hydride (LAH) in dry THF (0.1 M, 3 mL) is then added to the top of each reaction tube through the gasket at the top of the manifold. The apparatus is placed in an oil bath heated at 67°C for 8 hours, while maintaining a positive chilled nitrogen flow through the manifold (as set forth in Example 2). The apparatus is cooled to room temperature and 1 mL of water is added to the top of each reaction tube through the gasket at the top of the manifold, and the apparatus sonicated at room temperature for 30 minutes to neutralize any unreacted LAH. At the end of the reaction, the reaction tube array is drained and sparged as set forth in Example 1.

The reaction tube array is subjected to the standard wash cycle set forth in Example 1, except that a base wash cycle is added to remove the neutralized LAH salts.

The cyclic carbamate is formed according to the method of Gonzalez, et al, Tetrahedron Letters 1989;30:2145. The reaction tubes are submerged in reaction wells containing a solution of 0.2 M diethyl carbonate and 0.4 M NaOMe in MeOH. The apparatus is sealed as set forth in Example 1 and agitated in a sonic bath for 12 hours, while maintaining a positive nitrogen flow through the manifold. At the end of the reaction, the reaction tube array is drained and sparged as set forth in Example 1.

The reaction tube array is subjected to the standard wash cycle set forth in Example 1.

The appropriate reaction tubes are submerged in reaction wells containing a solution of 0.2 M iodoalkane (either 1-iodohexane, or iodomethane) in 3 mL of CH_2Cl_2 . The apparatus is sealed as set forth in Example 1 and agitated in a sonic bath while

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maintaining a positive nitrogen flow through the manifold for 1 hour. The apparatus then placed in an oil bath heated at 37°C, while maintaining a positive chilled nitrogen flow through the manifold (as set forth in Example 2) for 6 hours. At the end of the reaction, the reaction tube array is drained and sparged as set forth in Example 1.

The reaction tube array is subjected to the standard wash cycle set forth in Example 1.

To isolate and purify the products, the reaction tubes are submerged in reaction wells containing a solution 3 mL each of MeOH. The apparatus is sealed as set forth in Example 1, placed in an oil bath and heated at 64°C while maintaining a positive chilled nitrogen flow through the manifold as set forth in Example 2. At the end of the reaction, the reaction tube array is allowed to cool and then drained and sparged as set forth in Example 1.

The reaction tubes are washed as set forth in Example 1, using 3 mL portions of methanol. The methanol extraction protocol is repeated until the filtrates are free of any organic components as determined by TLC (2 times). The appropriate methanol filtrates are then combined and concentrated on a Speed-Vac® in tared vials to afford four discrete products.

EXAMPLE 14

3-Substituted 1-(Aminomethyl)-3,4-dihydro-5,6-dihydroxy-1H-2-benzopyrans

The synthesis of six benzopyrans is summarized in Scheme 14 and Table 14.

10 g of Merrifield's chloromethyl polystyrene resin (2% cross-linked, loading = 1.2 meq/g, 200-400 mesh) is reacted with excess sodium ethyl acetoacetate (10 eq) in DMF at 80°C for 16 hours. After washing with DMF

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and dioxane the crude beta-keto ester is hydrolyzed and decarboxylated 4N HCl:dioxane (1:1) at 60°C to give the phenethyl ketone resin, after washing with dioxane.

This is further reacted with excess catechol and cat.

5 p-TsOH in toluene at 90°C for 30 hours. The resin is thoroughly washed with toluene, dioxane, and CH₂Cl₂ to remove all catechol. The crude resin is then used directly below.

10 95 to 105 mg of the catechol ketal resin prepared above is measured into each of six gas dispersion tubes (reaction tubes). The reaction tubes are fitted into the appropriate holder block and swelled and drained as set forth in Example 1.

15 The appropriate reaction tubes are submerged in reaction wells containing a solution of nBuLi (1 eq) in THF at 0°C. The apparatus is sealed as set forth in Example 1 and agitated in a sonic bath (maintained at 0°C as set forth in Example 5), while maintaining a positive nitrogen flow through the manifold. After
20 1 hour, the bath is warmed to room temperature and sonicated for 3 hours more. The bath is then recooled to 0°C and a solution of either cyclohexyl- or phenyl-ethyleneoxide (2 eq) in THF (0.5 mL) is then added to the top of the appropriate reaction tube through the
25 gasket at the top of the manifold. The bath is again warmed to room temperature and sonicated for 2 hours. The apparatus is then removed from the sonic bath, and 0.5 mL of THF:H₂O (1:1) are added to the top of each reaction tube through the gasket at the top of the
30 manifold as above, to quench any excess n-BuLi. At the end of the reaction, the reaction tube array is drained and sparged as set forth in Example 1.

The reaction tubes are washed as set forth in Example 3, using 3 x 3 mL each of dioxane:sat NH₄Cl
35 (1:1), dioxane:H₂O (1:1), and dioxane.

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The appropriate reaction tubes are submerged in reaction wells containing a solution of (2 eq) of either bromoacetaldehyde dimethyl acetal (4 reaction tubes) or [(formylamino)methyl]acetaldehyde dimethyl acetal (2 reaction tubes); and BF_3OEt_2 (3 eq) in Et_2O (3 mL) at 0°C . The apparatus is sealed as set forth in Example 1 and agitated in a sonic bath at room temperature for 24 hours, while maintaining a positive nitrogen flow through the manifold. At the end of the reaction, the reaction tube array is drained and sparged as set forth in Example 1.

The reaction tubes are washed as set forth in Example 3, using 3 x 3 mL each of dioxane: H_2O (1:1), dioxane: H_2O 0.1N NaOH (1:1), dioxane: H_2O (1:1), dioxane.

The appropriate four reaction tubes are submerged in reaction wells containing a solution of either allylamine or benzylamine (2 eq) in dioxane (4 mL) at room temperature. The other two reaction tubes (those which are reacted with [(formylamino)methyl]-acetaldehyde dimethyl acetal above) are submerged in wells containing a solution of dioxane:MeOH:15% NaOH (2:2:1) 5 mL at room temperature. The apparatus is sealed as set forth in Example 1 and agitated in a sonic bath at 60°C as set forth in Example 5, while maintaining a positive chilled nitrogen flow through the manifold as set forth in Example 2. After 8 hours the reaction is stopped, the reaction tube array is allowed to cool, and then drained and sparged as set forth in Example 1.

The reaction tubes are washed as set forth in Example 3, using 3 x 2 mL each of dioxane, dioxane: H_2O (1:1), dioxane.

In order to cleave the products from the solid support, the reaction tubes are submerged in reaction wells containing a solution of 3 mL of 5 M HCl in

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dioxane:ethanol (1:1). The apparatus is sealed as set forth in Example 1 and agitated in a sonic bath at 80°C for 4 hours as set forth in Example 5, while maintaining a positive chilled nitrogen flow through the manifold as set forth in Example 2. After 4 hours the reaction is stopped, the reaction tube array is allowed to cool, and then drained and sparged as set forth in Example 1.

The reaction tubes are washed as set forth in Example 3, using 3 x 3 mL of dioxane:ethanol (1:1). The filtrates are then combined and concentrated as set forth in Example 3. The crude products are then redissolved, transferred to tared vials, and reconcentrated. The final crude products including, *cis*-1-(aminomethyl)-3-cyclohexyl-3,4-dihydro-5,6-dihydroxy-1*H*-2-benzopyran hydrochloride; *cis*-1-(aminomethyl)-3-phenyl-3,4-dihydro-5,6-dihydroxy-1*H*-2-benzopyran hydrochloride; *cis*-1-[(allylamino)methyl]-3-cyclohexyl-3,4-dihydro-5,6-dihydroxy-1*H*-2-benzopyran hydrochloride; *cis*-1-[(allylamino)methyl]-3-phenyl-3,4-dihydro-5,6-dihydroxy-1*H*-2-benzopyran hydrochloride; *cis*-1-[(benzylamino)methyl]-3-cyclohexyl-3,4-dihydro-5,6-dihydroxy-1*H*-2-benzopyran hydrochloride; *cis*-1-[(benzylamino)methyl]-3-phenyl-3,4-dihydro-5,6-dihydroxy-1*H*-2-benzopyran hydrochloride; are set forth in Table 14.

EXAMPLE 15

6,7-Dihydro-4*H*-pyrazolo[1,5-*a*]pyrrolo[3,4-*d*]pyrimidine-5,8-diones

The synthesis of eight pyrimidinediones is summarized in Scheme 15 and Table 15.

15 g of Merrifield's chloromethyl polystyrene resin (2% cross-linked, loading = 0.7 meq/g, 200-400 mesh) is reacted with excess cesium acrylate (10 eq) and NaI (0.1 eq) in DMF at 80°C for 14 hours to yield the

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d sired ester. The resin is thoroughly washed with DMF, dioxane:H₂O (1:1), dioxane, and CH₂Cl₂ to remove excess reagents. The crude resin is then used directly below.

5 95 to 105 mg of the acrylate ester resin prepared above is measured into each of eight gas dispersion tubes (reaction tubes). The reaction tubes are fitted into the holder block and swelled and drained as set forth in Example 1.

10 The appropriate reaction tubes are submerged in reaction wells containing a solution of either cyclohexylamine or n-hexylamine (1 eq) in 3 mL of DMF at room temperature. The apparatus is sealed as set forth in Example 1 and agitated in a sonic bath
15 (maintained at room temperature as set forth in Example 5), while maintaining a positive nitrogen flow through the manifold. After 24 hours, a solution of diethyl oxalate (1 eq) and Na₂CO₃ (1 eq) in 1 mL of DMF
20 is added to the top of the each reaction tube through the gasket at the top of the manifold. The bath is warmed to 60°C and sonicated for 2 hours. At the end of the reaction, the reaction tube array is allowed to cool and then drained and sparged as set forth in Example 1.

25 The reaction tubes are washed as set forth in Example 3, using 3 x 2 mL each of dioxane:sat. NH₄Cl (1:1), dioxane:1N HCl (1:1), dioxane:H₂O (1:1), and dioxane.

30 In order to cleave the products from the solid support, the appropriate reaction tubes are submerged in reaction wells containing a solution of (1 eq) of either 5-phenylpyrazol-3-amine or 5-(4-chlorophenyl) pyrazol-3-amine in HOAc (4 mL) at room temperature. The apparatus is sealed as set forth in Example 1 and
35 agitated in a sonic bath at 100°C for 2 hours as set forth in Example 5, while maintaining a positive

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chilled nitrogen flow through the manifold as set forth in Example 2. At the end of the reaction, the reaction tube array is allowed to cool and then drained and sparged as set forth in Example 1.

5 The reaction tubes are washed as set forth in Example 3, using 3 x 2 mL of dioxane:ethanol. The filtrates are then combined and concentrated as set forth in Example 3. The crude products are then redissolved, transferred to tared vials, and
10 reconcentrated.

 In an example of a post-cleavage reaction, a solution of either 2-chloroacetophenone (2 eq) and K_2CO_3 (1.1 eq) in DMF (3 mL) or 1-bromo-5-hexene (2 eq) and K_2CO_3 (1.1 eq) in DMSO (3 mL) is added to the
15 appropriate reaction wells. A holder block with clean empty reaction tubes is added and the apparatus is sealed and agitated as set forth in Example 1 in a sonic bath at 40°C for 22 hours as set forth in
20 Example 5, while maintaining a positive chilled nitrogen flow through the manifold as set forth in Example 2. At the end of the reaction, the reaction tube array is allowed to cool and then drained and sparged as set forth in Example 1

 The crude reaction mixtures are transferred to the
25 corresponding tubes in the SPE apparatus equipped with 20 mL columns and 5 μ m filters. Each reaction mixture is diluted and mixed with 15 mL of H_2O to precipitate the desired products. The suspensions are then filtered and washed with H_2O and Et_2O (3 x 2 mL, each).
30 The resulting solids are then dissolved with CH_2Cl_2 (3 x 2 mL washes) run through the filter and collected in clean tubes. These solutions are then concentrated as set forth in Example 3. The crude products are then redissolved, transferred to tared vials, and
35 reconcentrated. The final crude products including, 6-Cyclohexyl-6,7-dihydro-4-(phenylmethyl)-2-phenyl-

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4H-pyrazolo[1,5a]pyrrolo[3,4-d]-pyrimidine-5,8-dione;
6-Cyclohexyl-6,7-dihydro-4-(2-oxo-2-phenylethyl)-
2-phenyl-4H-pyrazolo[1,5-a]pyrrolo[3,4-d]-pyrimidine-
5,8-dione; 6-Cyclohexyl-6,7-dihydro-4-(phenylmethyl)-
2-(4-chlorophenyl)-4H-pyrazolo[1,5-a]pyrrolo[3,4-d]-
pyrimidine-5,8-dione; 6-Cyclohexyl-6,7-dihydro-
4-(2-oxo-2-phenylethyl)-2-(4-chlorophenyl)-
4H-pyrazolo[1,5-a]pyrrolo[3,4-d]-pyrimidine-5,8-dione;
6-Hexyl-6,7-dihydro-4-(phenylmethyl)-2-phenyl-
4H-pyrazolo[1,5-a]pyrrolo[3,4-d]-pyrimidine-5,8-dione;
6-Hexyl-6,7-dihydro-4-(2-oxo-2-phenylethyl)-2-phenyl-
4H-pyrazolo[1,5-a]-pyrrolo[3,4-d]-pyrimidine-5,8-dione;
6-Hexyl-6,7-dihydro-4-(phenylmethyl)-2-(4-chloro-
phenyl)-4H-pyrazolo[1,5-a]pyrrolo[3,4-d]-pyrimidine-
5,8-dione; 6-Hexyl-6,7-dihydro-4-(2-oxo-2-phenylethyl)-
2-(4-chlorophenyl)-4H-pyrazolo[1,5-a]pyrrolo[3,4-d]-
pyrimidine-5,8-dione are set forth in Table 15.

EXAMPLE 16

20 Synthesis of Tepoxalin Derivatives

The synthesis of eight tepoxalin derivatives is summarized in Scheme 16 and Table 16.

95 to 105 mg of benzylhydroxy polystyrene resin (loading = 1.08 meq/g, 200-400 mesh) is measured into each of eight gas dispersion tubes (reaction tubes). The apparatus is assembled, swelled with 3 mL of DMF, and drained as set forth in Example 1.

A solution (5 mL) of either 0.20 M 6-phenyl-4,6-dioxohexanoic acid, or 6-(4-chlorophenyl)-4,6-dioxohexanoic acid and 0.20 M carbonyldiimidazole in CH_2Cl_2 are stirred for 6 hours at room temperature. The two 6-aryl-4,6-dioxohexanoic acids employed are prepared by the method of Murray, et al, Journal of Organic Chemistry 1990;55:3424.

The appropriate reaction tubes are submerged in the reaction wells above. The apparatus is sealed as set

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forth in Example 1 and agitated in a sonic bath while maintaining a positive nitrogen flow through the manifold. After 12 hours, the reaction is stopped and the reaction tube array is drained and sparged as set forth in Example 1.

The reaction tubes are washed as set forth in Example 3, using 3 x 3 mL each of DMF, MeOH, dioxane, CH₂Cl₂, and MeOH.

The appropriate reaction tubes are submerged in reaction wells containing a solution of 5 mL of either 0.2 M 4-methoxyphenylhydrazine hydrochloride or 4-methylphenylhydrazine hydrochloride and 0.2 M triethylamine in MeOH. The apparatus is sealed as set forth in Example 1 and agitated in a sonic bath, while maintaining a positive nitrogen flow through the manifold. After 12 hours, the reaction is stopped and the reaction tube array is drained and sparged as set forth in Example 1.

The reaction tubes are washed as set forth in Example 3, using 3 x 3 mL each of DMF, MeOH, dioxane, and CH₂Cl₂.

Cleavage is accomplished by aminolysis of the resin ester linkage. Thus, the appropriate reaction tubes are submerged in reaction wells containing a solution of 5 mL of either 0.20 M methylhydroxylamine hydrochloride or benzylhydroxylamine hydrochloride and 0.2 M triethylamine in CH₂Cl₂ along with stirring bars. The apparatus is sealed with clamps as set forth in Example 1 and placed in an oil bath over a magnetic stirring plate while maintaining a positive flow of cold nitrogen as set forth in Example 2. The reaction vessels are warmed to reflux and stirred as above. After 24 hours, the reaction is stopped and the reaction tube array is drained and sparged as set forth in Example 1.

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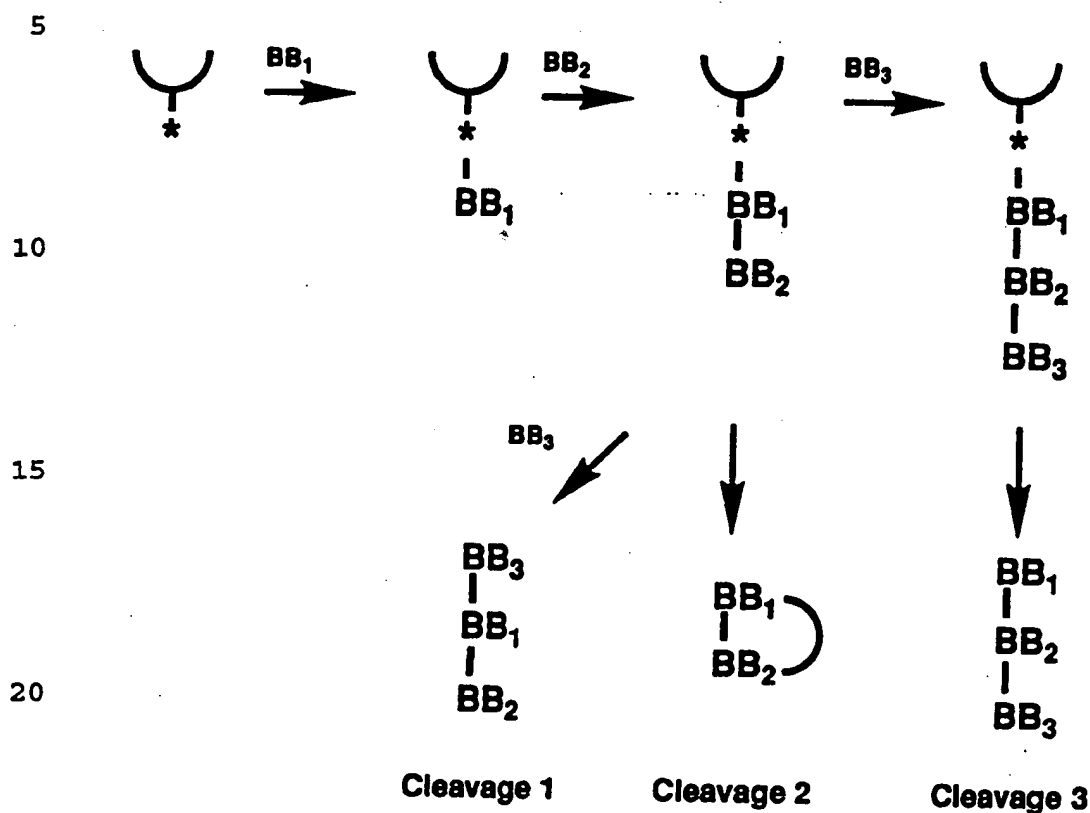
The reaction tubes are washed as set forth in Example 3, using 3 x 3 mL each of MeOH, dioxane, and CH₂Cl₂.

5 The appropriate filtrates are then combined and concentrated as set forth in Example 3. These crude products are partitioned between CH₂Cl₂ and 1 M HCl using a Tecan® robot to deliver the necessary liquids and to remove the aqueous layer. The acidic aqueous wash is repeated and the residual CH₂Cl₂ layers are
10 dried by passing the samples through a bed of MgSO₄ contained in standard SPE cartridges employing the vacuum SPE apparatus described above to yield tepoxalin and the expected tepoxalin derivatives.

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SCHEME 1

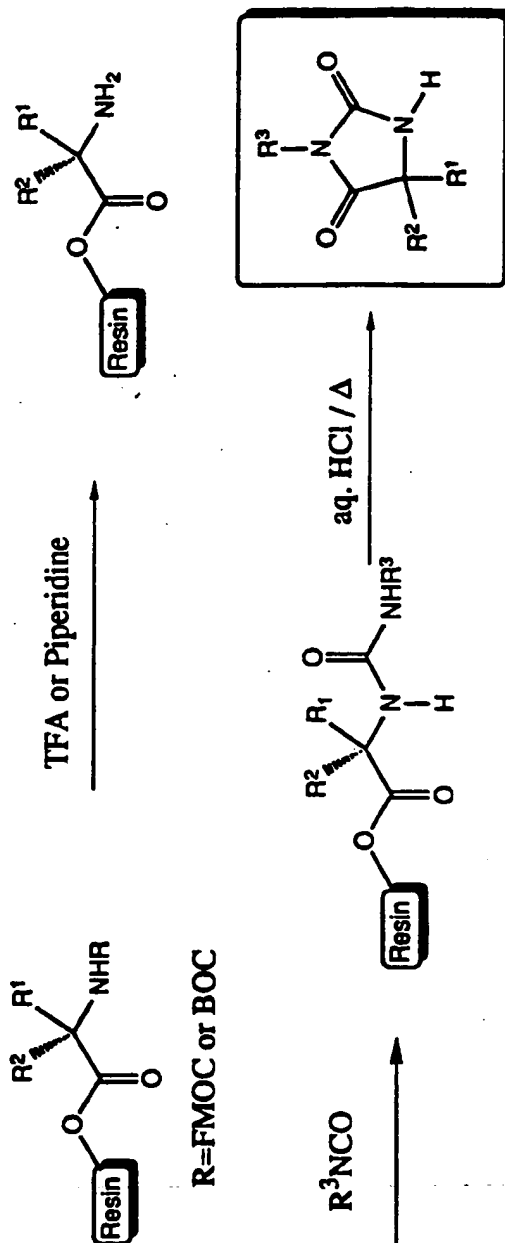


BB_N represents the building blocks which are bifunctional to allow sequential attachment. The curved structure represents the solid support where the asterisk is the functionality capable of covalently attaching the growing molecule to the solid support.

The three possible cleavage modes are illustrated. Cleavage 1 represents the third building block attacking the solid support linkage to cleave the final molecule. This provides structural variation at the former site of attachment to the support. Cleavage 2 represents a distal functionality attacking the solid support linkage to cleave the final molecule as a cyclized product. Cleavage 3 represents a cleavage by an invariant agent. This provides a constant functional group at the former site of attachment to the support.

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SCHEME 2



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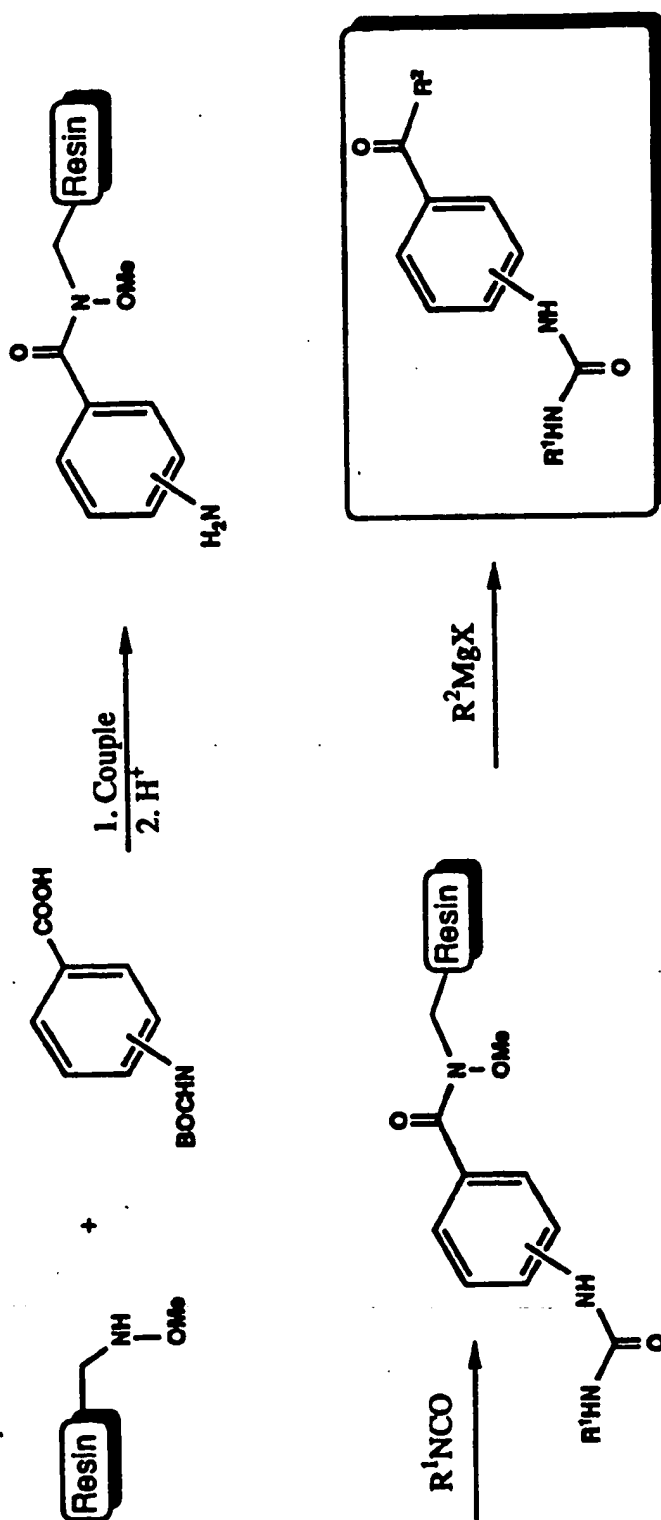
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SCHEME 5



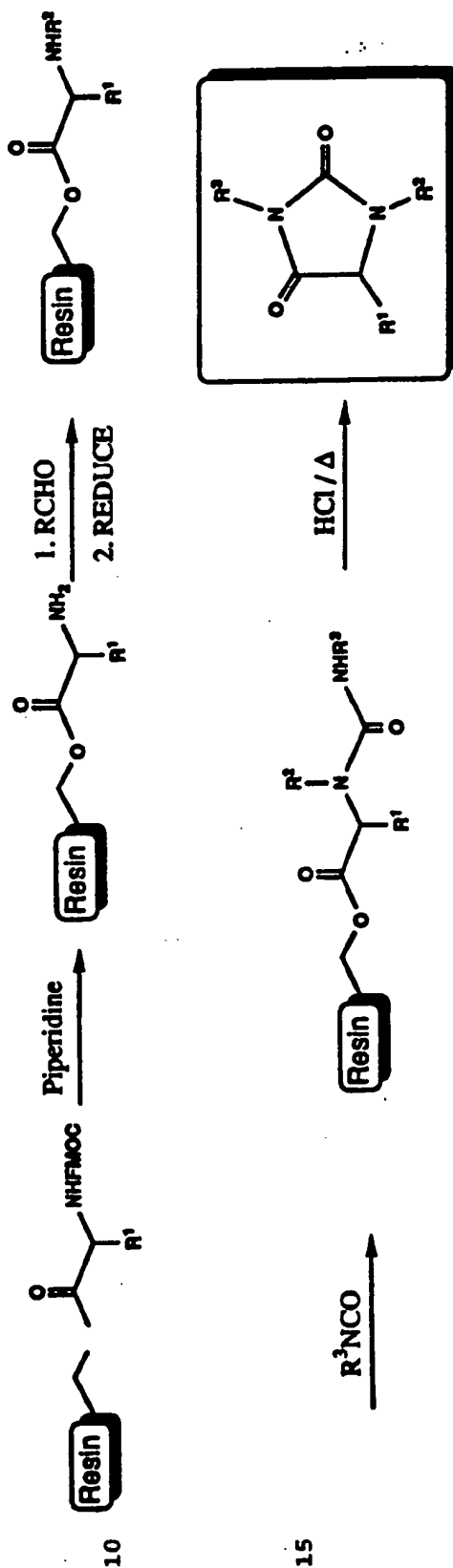
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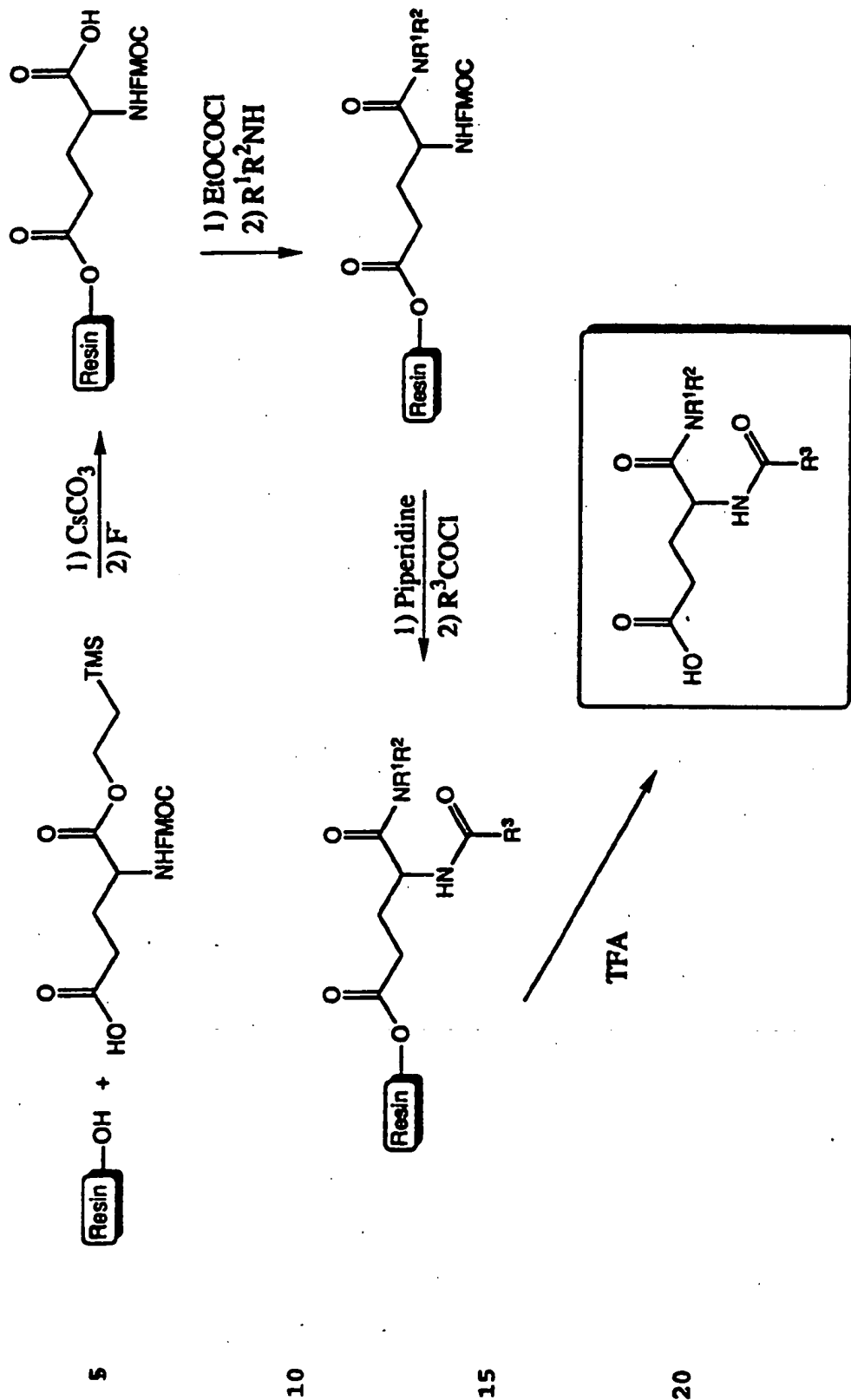
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SCHEME 6

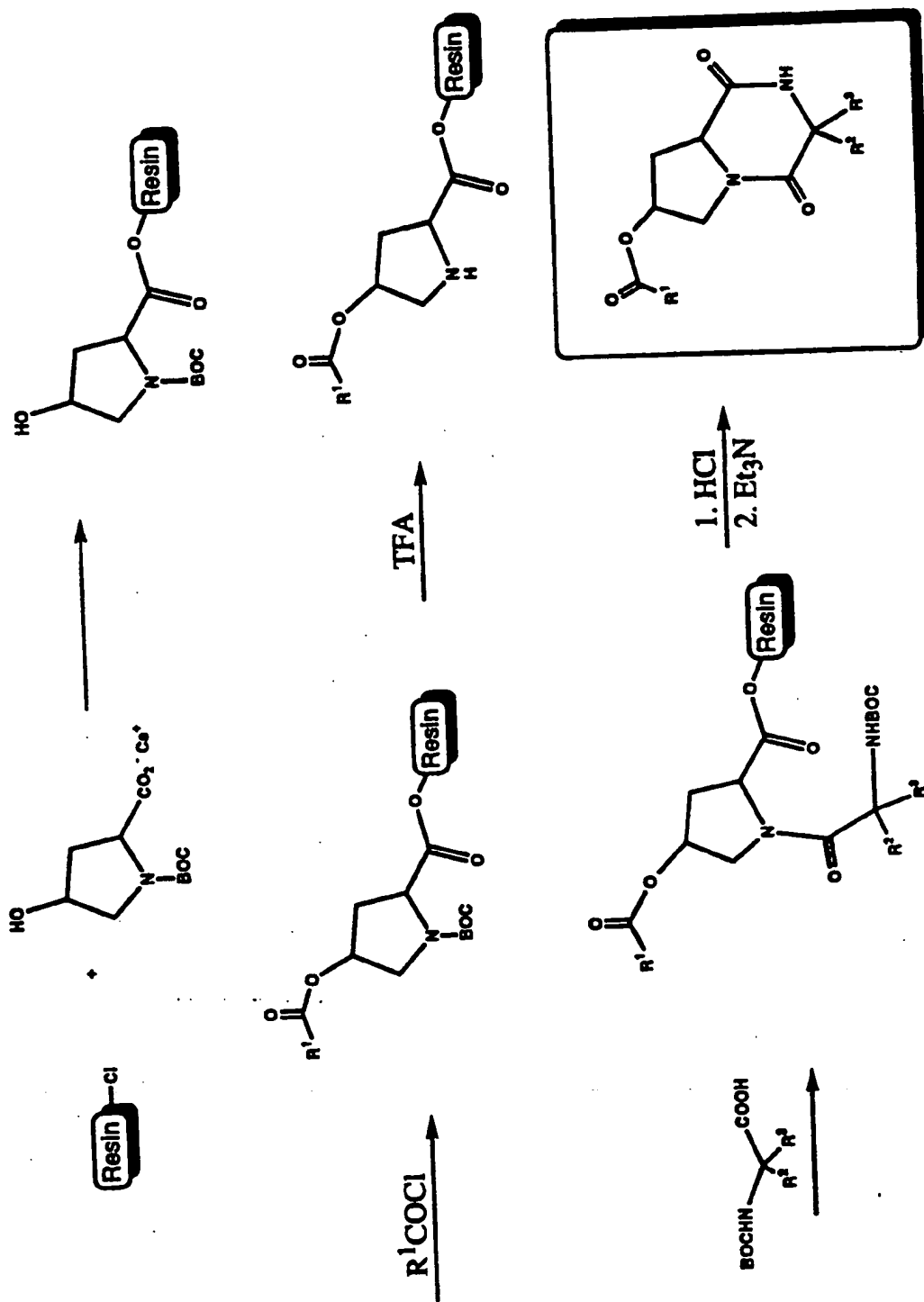


SCHEME 7

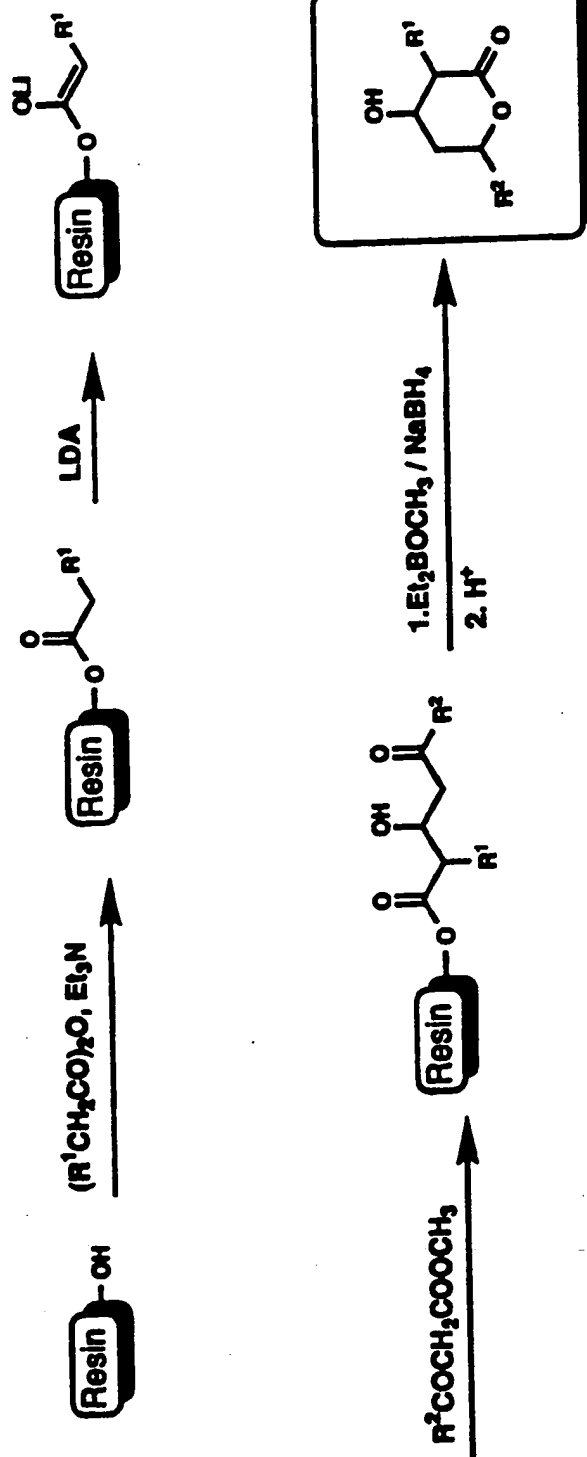


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SCHEME 8

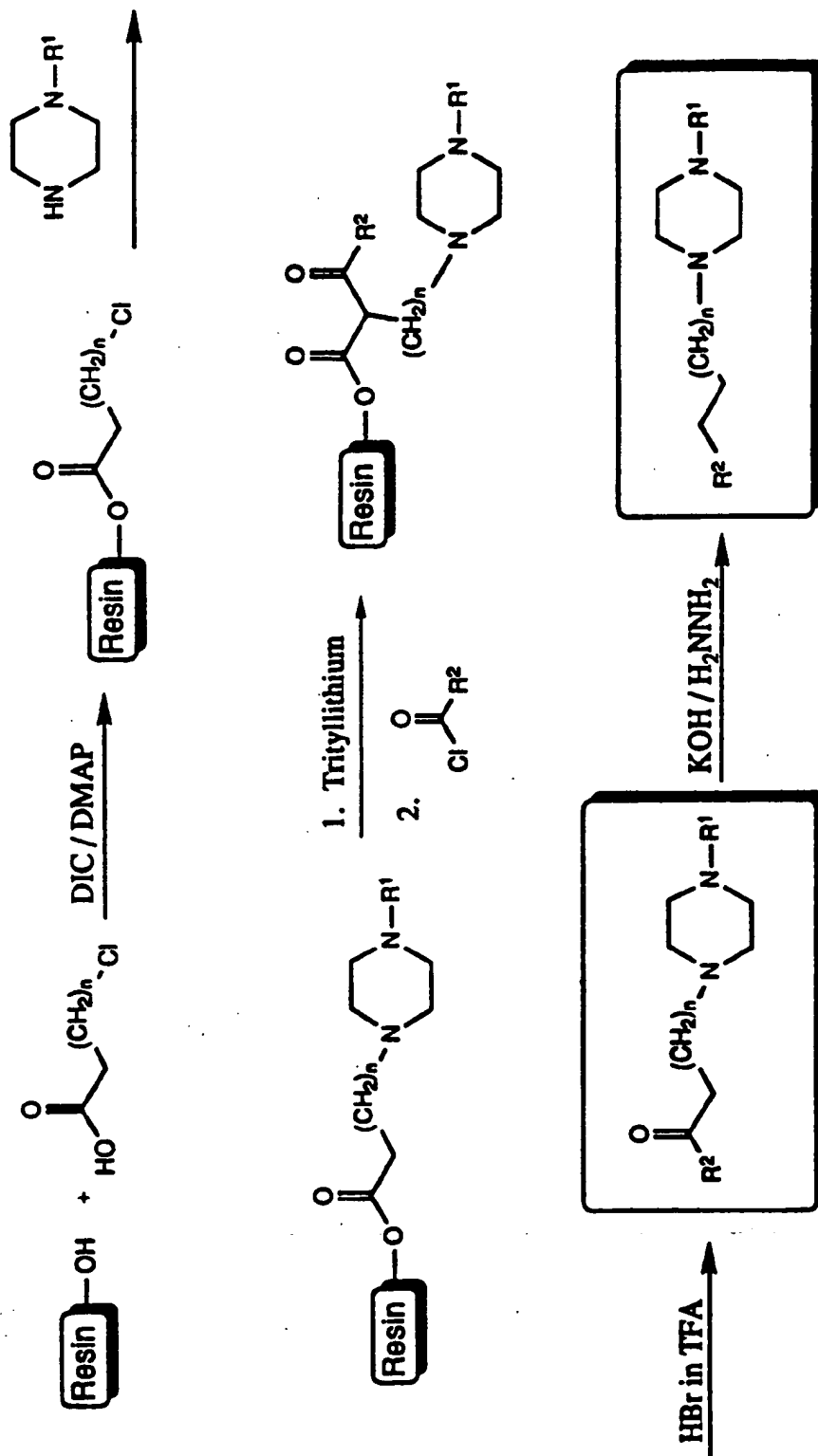


SCHEME 9



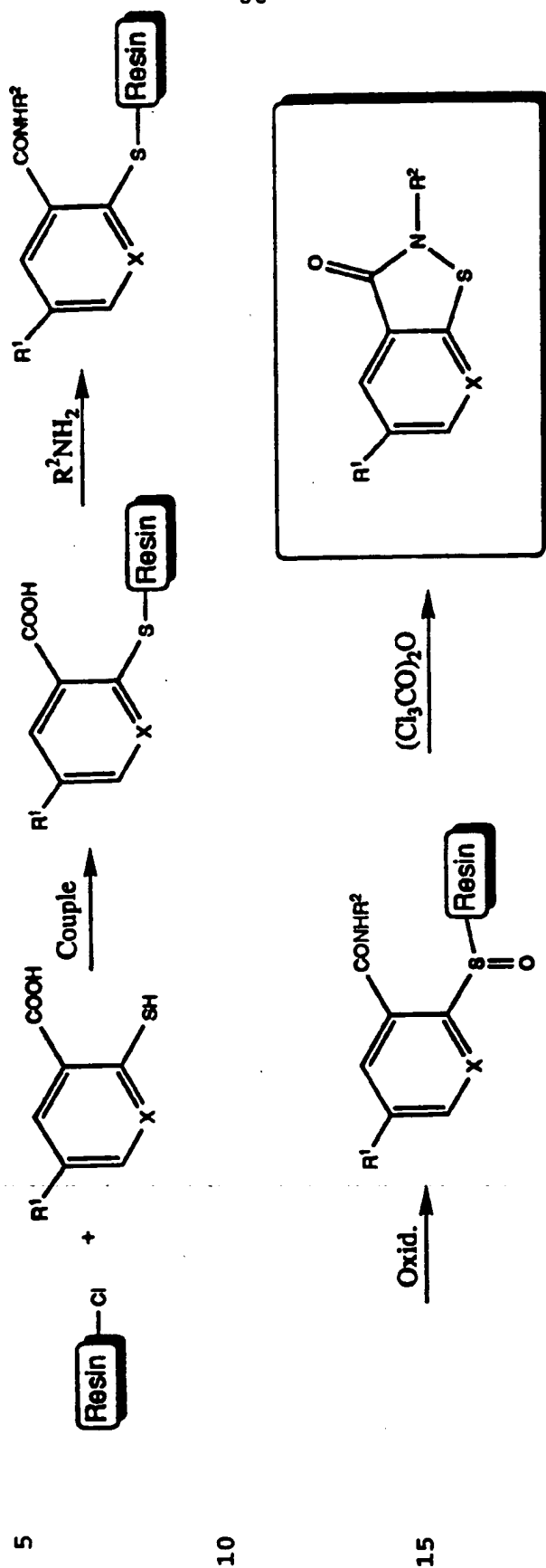
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SCHEME 10

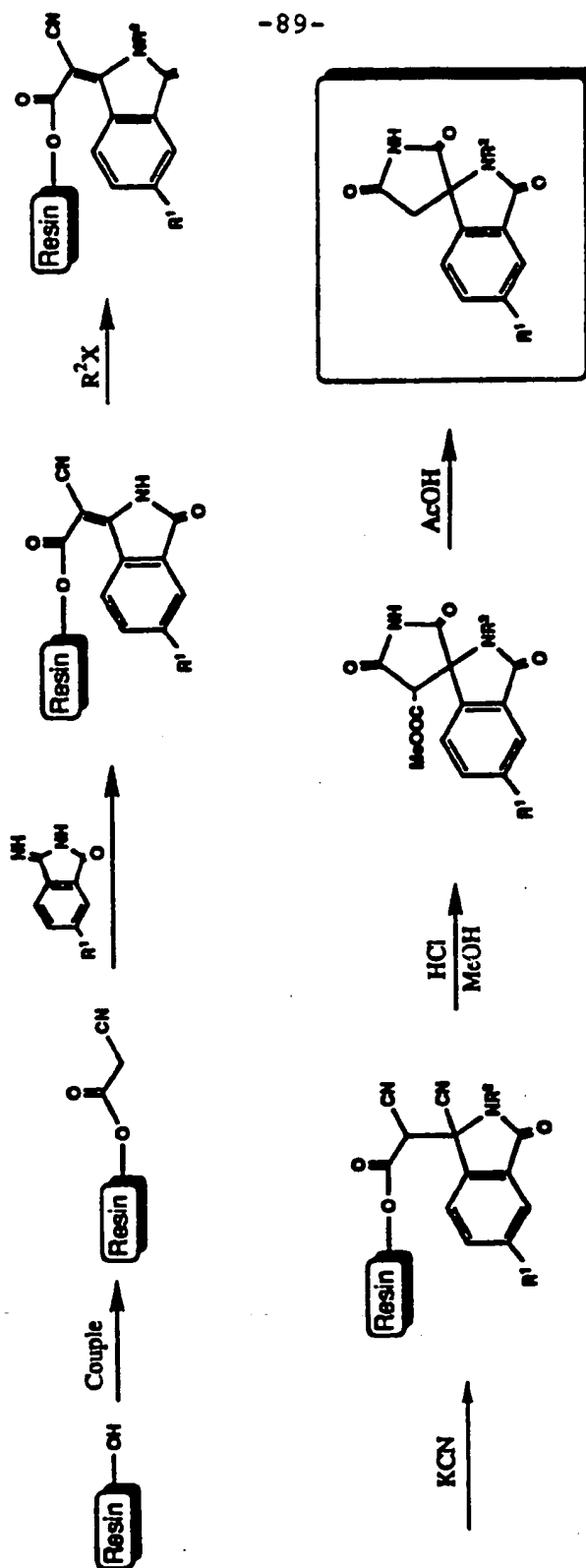


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SCHEME 11

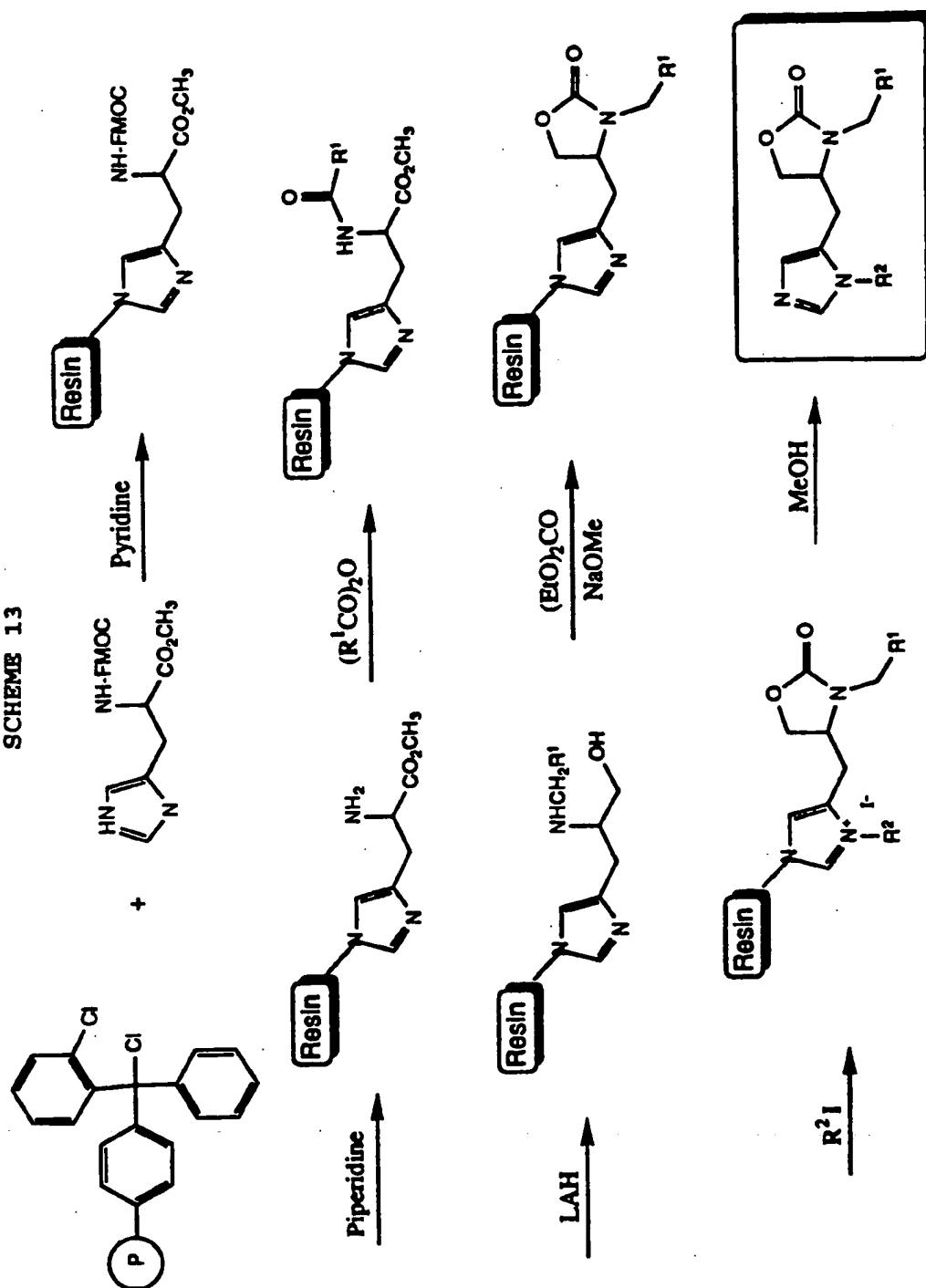


SCHEME 12



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SCHEME 13



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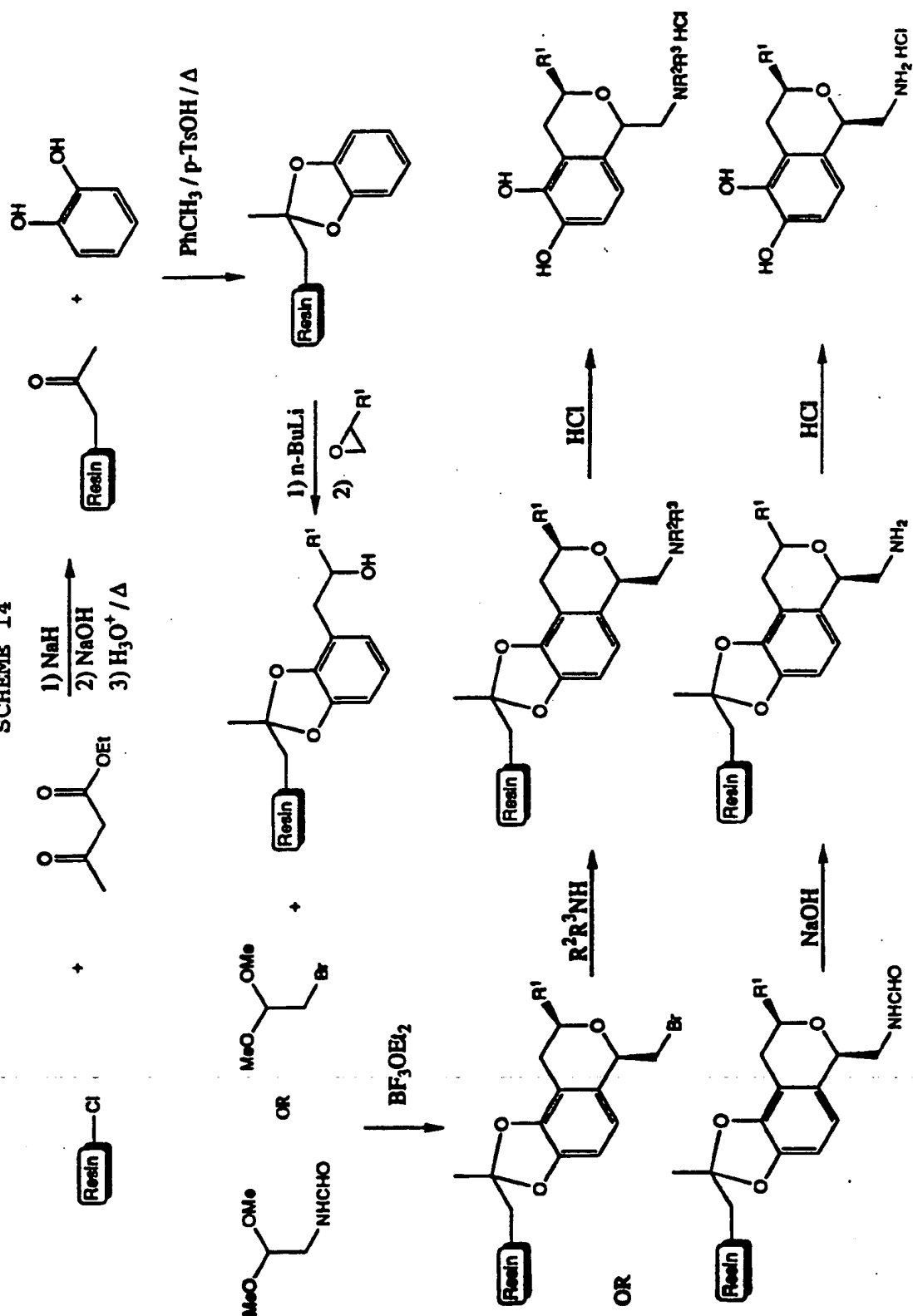
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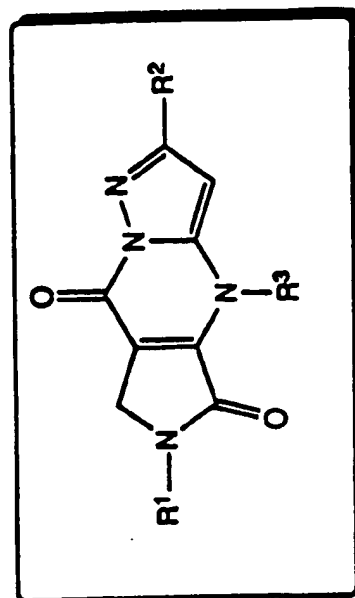
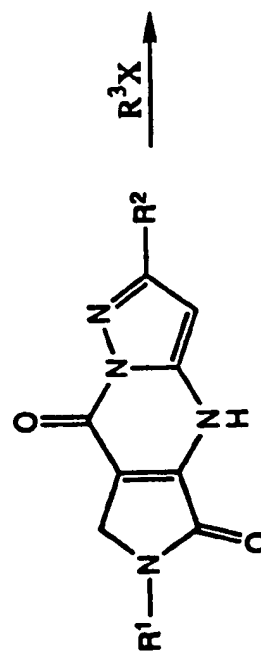
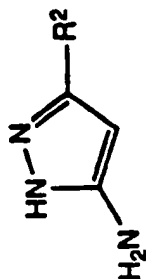
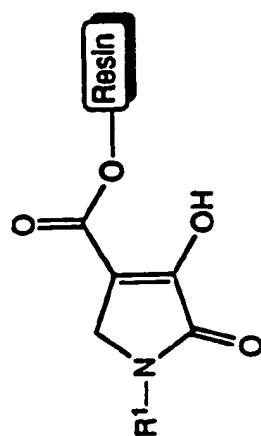
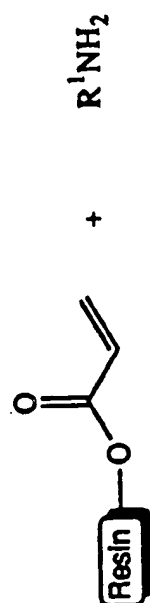
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SCHEME 14

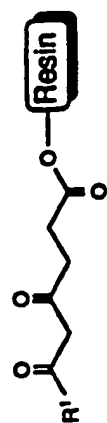


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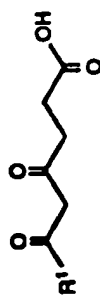
SCHEME 15



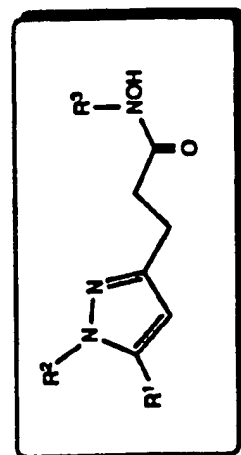
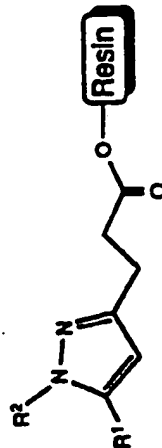
SCHEME 16



Couple



+


$$\text{R}^3\text{NHOH HCl}$$
 $\text{R}^2\text{NHNH}_2, \text{HCl}$

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TABLE 1. Dipeptides

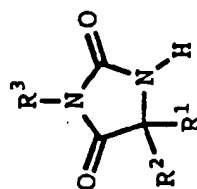
	<u>Dipeptide</u>	<u>Molecular Formula</u>	<u>M.W. g/mol</u>	<u>Yield (%)*</u>
5	AlaPhe	$C_{12}H_{16}N_2O_3 \cdot TFA$	350.29	40
	AlaGly	$C_5H_{10}N_2O_3 \cdot TFA$	260.17	41
	AlaIle	$C_9H_{18}N_2O_3 \cdot TFA$	316.28	85
	AlaLeu	$C_9H_{18}N_2O_3 \cdot TFA$	316.28	57
10	AlaAla	$C_6H_{12}N_2O_3 \cdot TFA$	274.19	45
	AlaPro	$C_8H_{14}N_2O_3 \cdot TFA$	300.23	28
	AlaVal	$C_8H_{16}N_2O_3 \cdot TFA$	302.25	77
	AlaTrp	$C_{14}H_{17}N_3O_3 \cdot TFA$	389.33	37
	IlePhe	$C_{15}H_{22}N_2O_3 \cdot TFA$	392.37	35
15	IleGly	$C_8H_{16}N_2O_3 \cdot TFA$	302.25	26
	IleIle	$C_{12}H_{24}N_2O_3 \cdot TFA$	358.36	85
	IleLeu	$C_{12}H_{24}N_2O_3 \cdot TFA$	358.36	30
	IleAla	$C_9H_{18}N_2O_3 \cdot TFA$	316.28	38
	IlePro	$C_{11}H_{20}N_2O_3 \cdot TFA$	342.31	29
20	IleVal	$C_{11}H_{23}N_3O_3 \cdot TFA$	344.33	62
	IleTrp	$C_{11}H_{23}N_3O_3 \cdot TFA$	431.41	32

* Yields based upon indicated loading of commercially available functionalized resins (0.37-0/60 mmeq./gm)

25

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TABLE 2. Hydantoins



Number	R1	R2	R3	Yield mg	Yield %
1	H	methyl	H	4.1	67%
2	H	benzyl	H	2.5	38%
3	H	H	H	3.3	65%
4	H	s-butyl	H	3.1	42%
5	H	i-butyl	H	4.9	61%
6	H	i-propyl	H	4.9	58%
7	H	2-methylindole	H	5.0	35%
8	phenyl	phenyl	H	1.4	5%
9	H	methyl	butyl	1.6	17%
10	H	benzyl	butyl	3.9	47%
11	H	H	butyl	1.0	13%
12	H	s-butyl	butyl	5.3	48%
13	H	i-butyl	butyl	0.7	7%
14	H	i-propyl	butyl	0.9	8%
15	H	2-methylindole	butyl	0.9	5%
16	phenyl	phenyl	butyl	1.6	5%

* Yields based upon reported loading of commercially available functionalized resins (0.34-1.04 mEq/g)

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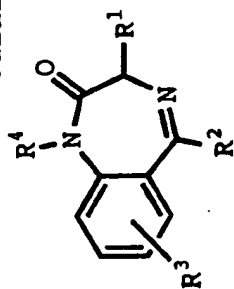
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TABLE 2. Hydantoins (continued)

Number	R1	R2	R3	Yield mg	Yield %
17	H	methyl	allyl	0.3	4%
18	H	benzyl	allyl	2.4	29%
19	H	H	allyl	3.7	48%
20	H	s-butyl	allyl	3.6	36%
21	H	i-butyl	allyl	5.0	54%
22	H	i-propyl	allyl	1.6	14%
23	H	2-methylindole	allyl	1.9	11%
24	phenyl	phenyl	allyl	2.1	7%
25	H	methyl	2-trifluorotolyl	2.6	23%
26	H	benzyl	2-trifluorotolyl	2.2	23%
27	H	H	2-trifluorotolyl	2.9	28%
28	H	s-butyl	2-trifluorotolyl	5.7	46%
29	H	i-butyl	2-trifluorotolyl	4.7	37%
30	H	i-propyl	2-trifluorotolyl	4.9	33%
31	H	2-methylindole	2-trifluorotolyl	3.0	15%
32	phenyl	phenyl	2-trifluorotolyl	0.0	0%
33	H	methyl	4-methoxyphenyl	3.1	22%
34	H	benzyl	4-methoxyphenyl	3.5	32%
35	H	H	4-methoxyphenyl	5.6	46%
36	H	s-butyl	4-methoxyphenyl	11.5	81%
37	H	i-butyl	4-methoxyphenyl	3.2	21%
38	H	i-propyl	4-methoxyphenyl	4.1	24%
39	H	2-methylindole	4-methoxyphenyl	4.9	22%
40	phenyl	phenyl	4-methoxyphenyl	3.0	7%

* Yields based upon reported loading of commercially available functionalized resins (0.34-1.04 mEq/g)

TABLE 3. Benzodiazepines



Number	R1	R2	R3	R4	Yield mg	Yield ‡	IC ₅₀ nM
1	methyl	phenyl	H	H	10.0	66‡	5795
2	methyl	phenyl	5-chloro	H	13.0	72‡	89
3	methyl	4-methoxyphenyl	H	H	14.0	81‡	>1 mM
4	methyl	phenyl	5-nitro	H	16.0	87‡	482
5	methyl	phenyl	4-methyl	isopropyl	7.0	37‡	>1 mM
6	methyl	phenyl	5-nitro	methyl	6.0	31‡	1000
7	methyl	cyclohexyl	H	H	7.0	43‡	>1 mM
8	methyl	thienyl	H	H	15.0	96‡	5902
9	H	phenyl	H	H	24.0	109‡	515
10	H	phenyl	5-chloro	H	13.0	54‡	32
11	H	4-methoxyphenyl	H	H	12.0	50‡	28255
12	H	phenyl	5-nitro	H	10.0	37‡	16
13	H	phenyl	4-methyl	isopropyl	<1	<5‡	18581
14	H	phenyl	5-nitro	methyl	15.0	56‡	64
15	H	cyclohexyl	H	H	4.0	18‡	25723
16	H	thienyl	H	H	17.0	75‡	1117
Y1 lds based on indicated loading of commercially available functionalized resins (0.37-0.60 mmEq/g)							

† IC₅₀ values were also obtained for the commercially available Diazepam (0.67 nM), Nordiazepam (0.2 nM), and Nitrazepam (1.46 nM) the latter 2 corresponding to samples Number 10 and 12, respectively.

TABLE 3. Benzodiazepines (continued)

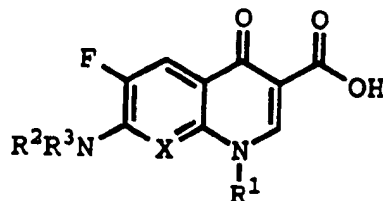
Number	R1	R2	R3	R4	Yield mg	Yield %	IC ₅₀ nM
17	2-methylindole	phenyl	H	H	12.0	54%	>1 mM
18	2-methylindole	phenyl	5-chloro	H	7.0	29%	>1 mM
19	2-methylindole	4-methoxyphenyl	H	H	6.0	25%	>1 mM
20	2-methylindole	phenyl	5-nitro	H	5.0	19%	12700
21	2-methylindole	phenyl	4-methyl	isopropyl	4.0	15%	>1 mM
22	2-methylindole	phenyl	5-nitro	methyl	3.0	12%	>1 mM
23	2-methylindole	cyclohexyl	H	H	7.0	31%	>1 mM
24	2-methylindole	thienyl	H	H	7.0	31%	>1 mM
25	4-hydroxytolyl	phenyl	H	H	12.0	60%	>1 mM
26	4-hydroxytolyl	phenyl	5-chloro	H	11.0	50%	>1 mM
27	4-hydroxytolyl	4-methoxyphenyl	H	H	9.0	42%	>1 mM
28	4-hydroxytolyl	phenyl	5-nitro	H	8.0	35%	>1 mM
29	4-hydroxytolyl	phenyl	4-methyl	isopropyl	9.0	39%	>1 mM
30	4-hydroxytolyl	phenyl	5-nitro	methyl	<1	<5%	>1 mM
31	4-hydroxytolyl	cyclohexyl	H	H	21.0	104%	>1 mM
32	4-hydroxytolyl	thienyl	H	H	30.0	145%	>1 mM
33	i-propyl	phenyl	H	H	17.0	77%	>1 mM
34	i-propyl	phenyl	5-chloro	H	10.0	39%	>1 mM
35	i-propyl	4-methoxyphenyl	H	H	7.0	29%	>1 mM
36	i-propyl	phenyl	5-nitro	H	7.0	27%	>1 mM
37	i-propyl	phenyl	4-methyl	isopropyl	11.0	42%	>1 mM
38	i-propyl	phenyl	5-nitro	methyl	6.0	22%	>1 mM
39	i-propyl	cyclohexyl	H	H	7.0	30%	>1 mM
40	i-propyl	thienyl	H	H	20.0	87%	>1 mM

* Yields based on indicated loading of commercially available functionalized resins (0.37-0.60 mmEq/g)

† IC₅₀ values were also obtained for the commercially available Diazepam (0.67 nM), Nordiazepam (0.2 nM), and Nitrazepam (1.46 nM) the latter 2 corresponding to samples Number 10 and 12, respectively.

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TABLE 4. Quinolones

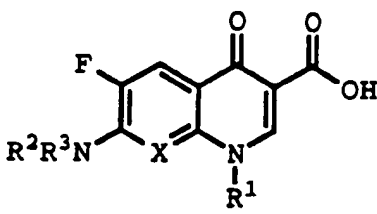





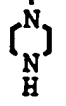



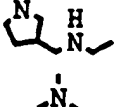











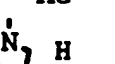
Cpd No.	R ¹	R ² R ³ N	X
1			CH
2			CH
3			CH
4			CH
5			CH
6			CH
7			CH
8			CH
9	Ethyl		CH
10	Ethyl		CH
11	Ethyl		CH
12	Ethyl		CH

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TABLE 4. Quinolones (continued)

5

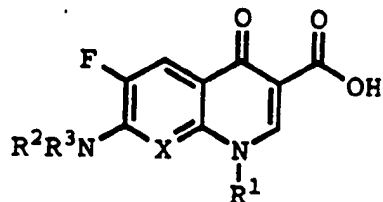


Cpd No.	R ¹	R ² R ³ N	X
10 13			CF
14			CF
15 15			CF
16			CF
20 17			CF
18			CF
25 19			CF
20 20			CF
21	Ethyl		CF
30 22	Ethyl		CF
23	Ethyl		CF
35 24	Ethyl		CF

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TABLE 4. Quinolones (continued)



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15

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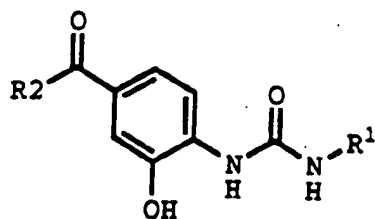
Cpd No.	R ¹	R ² R ³ N	X
25			N
26			N
27			N
28			N
29			N
30			N
31			N
32			N
33	Ethyl		N
34	Ethyl		N
35	Ethyl		N
36	Ethyl		N

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TABLE 5. Keto-ureas

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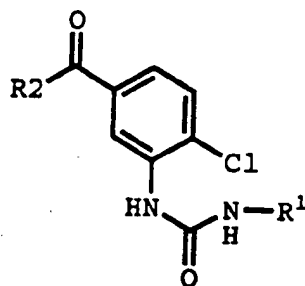


10

Cpd No.	R ¹	R ²
1	Ethyl	
2		
3	Ethyl	Methyl
4		Methyl

15

20



25

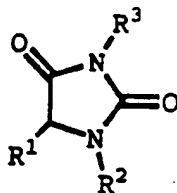
30

35

Cpd No.	R ¹	R ²
5	Ethyl	
6		
7	Ethyl	Methyl
8		Methyl

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TABLE 6. N²-Substituted Hydantoins

	Cpd No.	R ¹	R ²	R ³
5	1	CH ₃	CH ₃ CH ₂	H
10	2	CH ₃	CH ₃ CH ₂	CH ₂ CH=CH ₂
15	3	CH ₃	C ₆ H ₅ CH ₂	H
20	4	CH ₃	C ₆ H ₅ CH ₂	CH ₂ CH=CH ₂
	5	C ₆ H ₅ CH ₂	CH ₃ CH ₂	H
25	6	C ₆ H ₅ CH ₂	CH ₃ CH ₂	CH ₂ CH=CH ₂
	7	C ₆ H ₅ CH ₂	C ₆ H ₅ CH ₂	H
30	8	C ₆ H ₅ CH ₂	C ₆ H ₅ CH ₂	CH ₂ CH=CH ₂

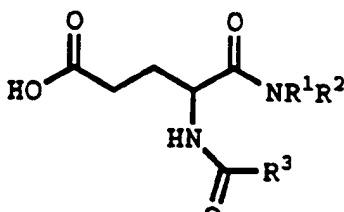
35





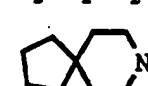

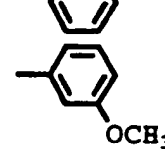

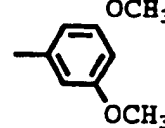
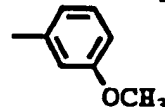

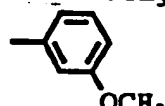
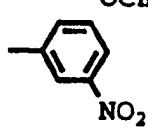

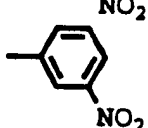
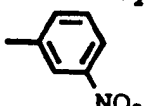

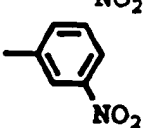
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TABLE 7. (R)-4-BENZAMIDO-5-OXOPENTANOIC ACIDS

5



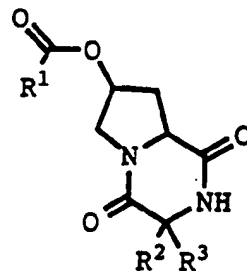
Cpd No.	R ¹ R ² N	R ³
1	nBuHN	
2		
3	(CH ₃ CH ₂ CH ₂ CH ₂ CH ₂) ₂ N	
4		
5	nBuHN	
6		
7	(CH ₃ CH ₂ CH ₂ CH ₂ CH ₂) ₂ N	
8		
9	nBuHN	
10		
11	(CH ₃ CH ₂ CH ₂ CH ₂ CH ₂) ₂ N	
12		

10
15
20
25
30
35

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TABLE 8. Diketopiperazines



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10

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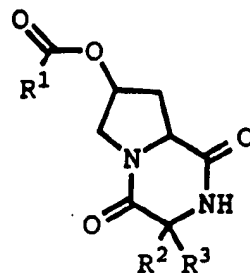
30

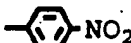



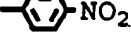
35

Cpd No.	R ¹	R ²	R ³
1		H	H
2		H	CH ₃
3		H	(CH ₃) ₂ CH
4		H	PhCH ₂
5		Ph	Ph
6		H	H
7		H	CH ₃
8		H	(CH ₃) ₂ CH
9		H	PhCH ₂
10		Ph	Ph
11		H	H
12		H	CH ₃
13		H	(CH ₃) ₂ CH
14		H	PhCH ₂
15		Ph	Ph
16		H	H
17		H	CH ₃
18		H	(CH ₃) ₂ CH
19		H	PhCH ₂
20		Ph	Ph

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TABLE 8. Diketopiperazines (continued)



Cpd No.	R ¹	R ²	R ³
21	-CH ₃	H	H
22	-CH ₃	H	CH ₃
23	-CH ₃	H	(CH ₃) ₂ CH
24	-CH ₃	H	PhCH ₂
25	-CH ₃	Ph	Ph
26	 NO ₂	H	H
27	 NO ₂	H	CH ₃
28	 NO ₂	H	(CH ₃) ₂ CH
29	 NO ₂	H	PhCH ₂
30	 NO ₂	Ph	Ph
31	-CF ₃	H	H
32	-CF ₃	H	CH ₃
33	-CF ₃	H	(CH ₃) ₂ CH
34	-CF ₃	H	PhCH ₂
35	-CF ₃	Ph	Ph
36	-tBu	H	H
37	-tBu	H	CH ₃
38	-tBu	H	(CH ₃) ₂ CH
39	-tBu	H	PhCH ₂
40	-tBu	Ph	Ph

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TABLE 9. 2H-Pyranones

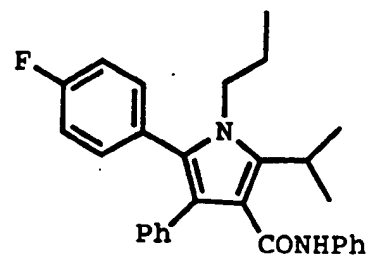


Cpd No.	R ¹	R ²
---------	----------------	----------------

10

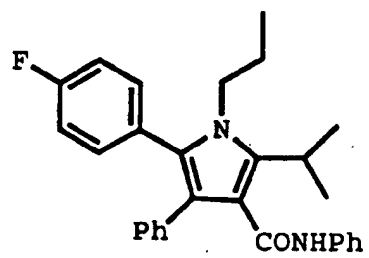
1

H



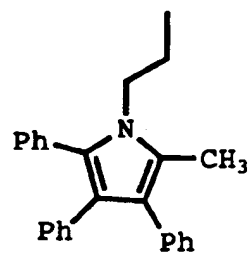
15

2

CH₃CH₂

20

3

C₆H₅CH₂

25

30

35

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TABLE 10. N-aryl Piperazines

5

Cpd No.

R¹

R²

Cpd No.

R¹

R²

10

1

H

7

H

15

2

H

8

H

3

9

20

4

10

25

5

11

6

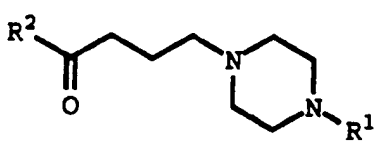
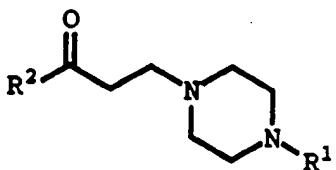
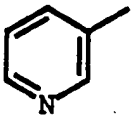
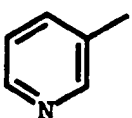
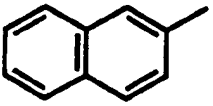
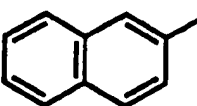

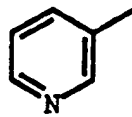

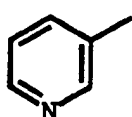

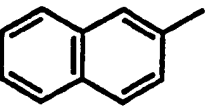

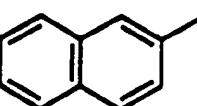
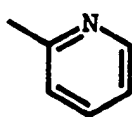
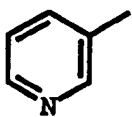
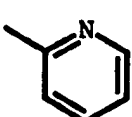
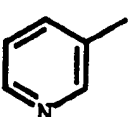
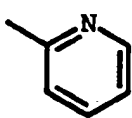
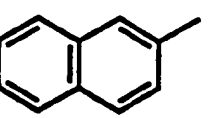
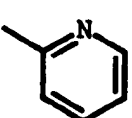
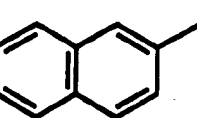
12

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TABLE 10. N-aryl Piperazines (continued)

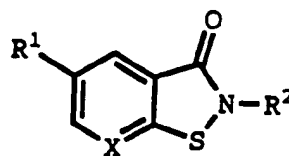
5						
	Cpd No.	R ¹	R ²	Cpd No.	R ¹	R ²
10	13	H		19	H	
15	14	H		20	H	
	15			21		
20	16			22		
25	17			23		
	18			24		

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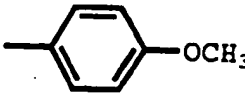

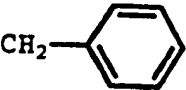
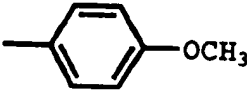

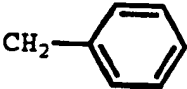
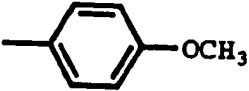

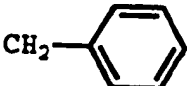
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TABLE 11. Benzoisothiazolones

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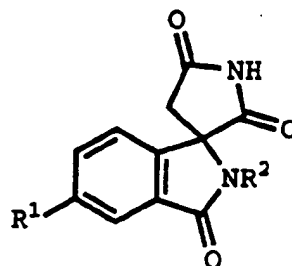
Cpd No.	X	R ¹	R ²
1	CH	H	
2	CH	H	
3	CH	H	
4	CH	Cl	
5	CH	Cl	
6	CH	Cl	
7	N	H	
8	N	H	
9	N	H	

35

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TABLE 12. Spirosuccimides



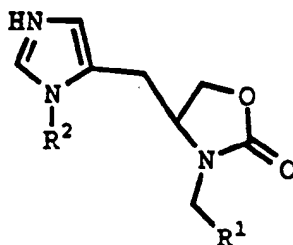
10

Cpd No.	R ¹	R ²
1	H	CH ₃
2	H	
15	3	
4	Cl	CH ₃
20	5	
6	Cl	
25	7	CH ₃
8		
30	9	
35		

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TABLE 13. Pilocarpine Analogs



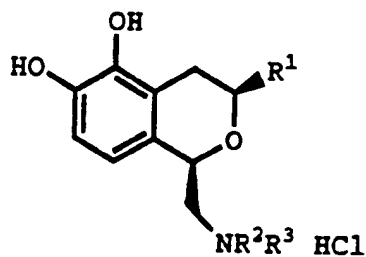
Cpd No.	R ¹	R ²
1	CH ₃	CH ₃
2	CH ₃	Hexyl
3	(CH ₃) ₂ CH	CH ₃
4	(CH ₃) ₂ CH	Hexyl

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TABLE 14. 3-Substituted 1-(aminomethyl)-3,4-dihydro-5,6-dihydroxy-1H-2-benzopyrans

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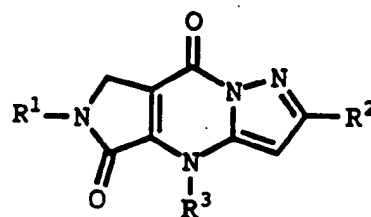
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Cpd No.	R^1	R^2R^3N
1		NH_2
2		NH_2
3		
4		
5		
6		

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TABLE 15. Pyrimidinediones



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Cpd No.	R ¹	R ²	R ³
1	Hexyl		
2	Hexyl		
3	Hexyl		
4	Hexyl		
5			
6			
7			
8			

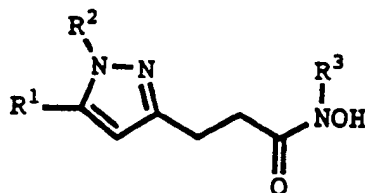
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TABLE 16. Tepoxalin Derivatives

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Cpd No.	R ¹	R ²	R ³
1			
2			Methyl
3			
4			Methyl
5			
6			Methyl
7			
8			Methyl

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CLAIMS

1. An apparatus for multiple, simultaneous synthesis of compounds comprising:
 - 5 a plurality of reaction tubes, each reaction tube having an upper end and a lower end, said lower end containing a filter device;
 - a reservoir block having a means for receiving the filter devices on the lower ends of said plurality of reaction tubes;
 - 10 a holder block with a plurality of apertures at least as large as the outside diameter of at least one of said plurality of reaction tubes;
 - a manifold located adjacent to said holder block, said manifold enclosing the upper ends of said plurality of reaction tubes;
 - 15 a means for providing a seal between said holder block and said reservoir block;
 - a means for providing a seal between said holder block and the manifold; and
 - 20 a means for fastening together said reservoir block, said holder block, and said manifold.
2. An apparatus according to Claim 1 in which at least one of the plurality of reaction tubes comprises gas dispersion tubes.
3. An apparatus according to Claim 2 in which the length of at least one of the plurality of gas dispersion tubes is from 50 to 300 mm.
4. An apparatus according to Claim 3 in which the outside diameter of the upper end of at least one of the plurality of gas dispersion tubes is from 5 to 25 mm.

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5. An apparatus according to Claim 4 in which the inside diameter of the upper end of at least one of the plurality of gas dispersion tubes is from 1 to 24 mm.
6. An apparatus according to Claim 5 in which the length of the filter on the lower end of at least one gas dispersion tube is from 1 to 300 mm.
7. An apparatus according to Claim 6 in which the outside diameter of the filter on the lower end of at least one gas dispersion tube is from 5 to 30 mm.
8. An apparatus according to Claim 7 in which the inside diameter of the filter on the lower end of at least one gas dispersion tube is from 1 to 24 mm.
9. An apparatus according to Claim 8 in which the filter on the lower end of at least one gas dispersion tube has a porosity of 10 to 400 μ .
10. An apparatus according to Claim 9 in which the filter on the lower end of at least one gas dispersion tube is constructed of fritted glass.
11. An apparatus according to Claim 10 which provides a means for equilibration of any pressure differentials within the sealed apparatus.
12. An apparatus according to Claim 11 in which at least one gas dispersion tube has at least one pressure equalization hole located above the filter on the lower end of the gas dispersion tube.

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13. An apparatus according to Claim 12 in which at least one pressure equalization hole is located from 1 to 250 mm above the filter on the lower end of the gas dispersion tube.
14. An apparatus according to Claim 13 in which at least one pressure equalization hole has a diameter from 100 to 2000 μm .
15. An apparatus according to Claim 1 in which the reservoir block provides a means for receiving the filter on the lower end of at least one of the plurality of reaction tubes.
16. An apparatus according to Claim 15 in which the means for receiving the filter on the lower end of the plurality of reaction tubes is a plurality of reaction wells.
- 5 17. An apparatus according to Claim 16 in which the filter located on the lower end of at least one of the plurality of reaction tubes is fit into at least one of the plurality of reaction wells in the reservoir block to form a reaction vessel.
18. An apparatus according to Claim 17 in which the reservoir block comprises a reservoir rack which is adapted to hold at least as many reaction wells as the number of reaction tubes.
- 5 19. An apparatus according to Claim 18 in which at least one of the plurality of reaction wells is sufficient in depth and diameter so that the reaction well may receive the filter on the lower end of one reaction tube and an amount of reactant required for organic synthesis.

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20. An apparatus according to Claim 19 in which the inside diameter of at least one of the plurality of reaction wells is from 7 to 100 mm.
21. An apparatus according to Claim 20 in which at least one of the reaction wells is removable.
22. An apparatus according to Claim 21 in which at least one of the reaction wells is constructed of glass.
23. An apparatus according to Claim 1 in which at least one of the apertures in the holder block is coaxial to the apertures in the reservoir block.
24. An apparatus according to Claim 23 in which the upper end of at least one of the plurality of reaction tubes passes through at least one of the plurality of apertures in the holder block.
25. An apparatus according to Claim 24 in which the holder block prevents torquing, wobbling, or vertical movement of the plurality of reaction tubes.
26. An apparatus according to Claim 1 in which the manifold has at least four side walls and one top wall, said top wall having an upper face and a lower face.
27. An apparatus according to Claim 26 in which the manifold is open on the end adjacent to the holder block.

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28. An apparatus according to Claim 27 in which the manifold has a plurality of ports on one or more walls.
29. An apparatus according to Claim 28 in which the manifold has two ports on one face.
30. An apparatus according to Claim 29 in which the manifold is constructed of a transparent material.
31. An apparatus according to Claim 30 in which the top wall of the manifold has a plurality of apertures which are coaxial to the plurality of apertures in the holder block.
32. An apparatus according to Claim 31 in which a plate is located adjacent to the upper face of the top wall of the manifold.
33. An apparatus according to Claim 32 in which the plate has a plurality of apertures which are coaxial to the plurality of apertures in the holder block.
34. An apparatus according to Claim 1 in which the means for providing a seal between the holder block and the manifold comprises at least one first gasket.
35. An apparatus according to Claim 1 in which the means for providing a seal between the holder block and the reservoir block comprises at least one second gasket.

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36. An apparatus according to Claim 33 in which the manifold and the plate are separated by at least one third gasket.
37. An apparatus according to Claim 36 in which the at least one third gasket is penetrable with a needle-like object.
38. An apparatus according to Claim 1 in which the reservoir block is releasably attached to the holder block and the manifold.
- 5 39. An apparatus according to Claim 38 in which the means for fastening together the reservoir block, the holder block, and the manifold are one or more spring clips affixed to one or more sides of the reservoir block, the holder block, or the manifold.
- 5 40. An apparatus according to Claim 39 in which the means for fastening together the reservoir block, the holder block, and the manifold are rods which extend longitudinally upward throughout each of the reservoir block, the holder block, and the manifold.
- 5 41. An apparatus according to Claim 35 in which a means for fastening the plate and the at least one third gasket of the upper face of the manifold is two or more spring clips attached to at least one side wall of the manifold which snap into grooves on the upper face of the plate.
42. An apparatus according to Claim 11 in which pressure equalization is achieved with a hollow tube placed around the upper end of at least ne

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- 5 of the gas dispersion tubes, said hollow tube having one or more protrusions located in two or more locations along its inner face, such that said one or more protrusions contact the gas dispersion tube and prevent the gas dispersion tube from lateral or vertical movement.
43. An apparatus according to Claim 11 in which pressure equalization is achieved with at least one capillary tube located adjacent and parallel to at least one gas dispersion tube, said
5 capillary tube extending from the reaction well, through an aperture in the holder block, into the manifold.
45. An apparatus according to Claim 1 in which the means for providing a seal between the holder block, at least one reaction tube, and the manifold consists of an o-shaped ring.
46. An apparatus according to Claim 45 in which a first spherical joint, opening downward and tooled with a groove to accommodate the o-shaped ring, is fused to the outer wall of the reaction tube.
47. An apparatus according to Claim 46 in which a second spherical joint having the same diameter and groove size of the first spherical joint and opening upward to accommodate the o-shaped ring is
5 fused to the top of at least one reaction well.
48. An apparatus according to Claim 47 in which the outside diameter of the first and second spherical joints is from 5 to 50 mm.

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- 5 49. An apparatus according to Claim 48 in which the position of the first spherical joint is above the filter and the pressure equalization holes and at a height necessary to adequately accommodate the filter within the sealed reaction well when the seal is achieved.
50. An apparatus according to Claim 49 in which the position of the first spherical joint may be from 1 to 299 mm above the top of the filter.
- 5 51. An apparatus according to Claim 50 in which the position of the corresponding second spherical joint fused to the top of the reaction well is also at a height necessary to adequately accommodate the filter within the sealed reaction well.
52. An apparatus according to Claim 51 in which the position of the second spherical joint may be from 1 to 299 mm above the bottom of the reaction well.
53. An apparatus according to Claim 52 in which the face of at least one aperture on the lower side of the holder block is chamfered to accommodate at least one first spherical joint.
54. An apparatus according to Claim 53 in which the face of at least one aperture on the upper side of the reservoir rack is chamfered to accommodate at least one second spherical joint.
55. An apparatus according to Claim 1 in which a vacuum box having an open upper face surrounds the reservoir block.

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56. An apparatus according to Claim 55 in which a second rack which may support one or more removable cartridges is located adjacent to the open upper face of the vacuum box.
- 5 57. An apparatus according to Claim 56 in which the means for providing a seal between the open upper face of the vacuum box and second rack is one or more gaskets adhered to the lower side of the base of the second rack.
- 5 58. An apparatus according to Claim 57 in which the means for fastening the vacuum box and second rack is two or more spring clips attached to the side walls of the vacuum box and which snap into the grooves on the upper side of the base of the second rack.
- 5 59. An apparatus according to Claim 58 in which one or more needle-like objects, corresponding in number and location to the reaction wells, are mounted in the base of the second rack, protrude through the gasket, and open into the test tubes.
60. An apparatus according to Claim 59 in which the removable cartridges are used in combination with the needle-like objects.
61. A method for the multiple, simultaneous synthesis of compounds using an apparatus according to Claim 1 comprising the steps of:
- (a) charging the apparatus with the reactants required for synthesis;
 - (b) synthesizing the compounds; and
 - (c) cleaving the compounds from a solid support within the apparatus.

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62. A method according to Claim 61 wherein in step (a) the reactants comprise:
a functionalized solid support;
a building block;
5 a coupling reagent; and
a solvent.
63. A method according to Claim 61 wherein in step (a) the reactants comprise:
a functionalized solid support;
a reactive building block; and
5 a solvent.
64. A method according to Claim 61 wherein in step (a) the reactants comprise:
a solid support with attached building block;
a second building block;
5 a coupling reagent; and
a solvent.
65. A method according to Claim 61 wherein in step (a) the reactants comprise:
a solid support with attached building block;
a second reactive building block; and
5 a solvent.
66. A method for the multiple, simultaneous synthesis of compounds using an apparatus according to Claim 1 comprising the steps of:
5 (a) charging the apparatus with a solid support with attached building block wherein the building block has a reactive moiety protected by a protecting group and a solvent;

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- 10 (b) removing the protecting group from the
reactive moiety with a deprotection
reagent;
- (c) removing the deprotection reagent;
- 15 (d) sequentially adding additional reactive
building blocks in solvents to
synthesize the compounds; and
- (e) cleaving the compounds from the solid
support within the apparatus.

67. A method for the multiple, simultaneous synthesis
of compounds using an apparatus according to
Claim 1 comprising the steps of:

- 5 (a) charging the apparatus with a solid
support with attached building block
wherein the building block has a
reactive moiety protected by a
protecting group and a solvent;
- 10 (b) removing the protecting group from the
reactive moiety with a deprotection
reagent;
- (c) removing the deprotection reagent;
- (d) adding a coupling reagent in a solvent;
- 15 (e) sequentially adding additional reactive
building blocks and optionally coupling
reagents in solvents to synthesize the
compounds; and
- (f) cleaving the compounds from the solid
support within the apparatus.

68. A method for the multiple, simultaneous synthesis
of compounds using an apparatus according to
Claim 1 comprising the steps of:

- 5 (a) charging the apparatus with a solid
support with attached building block and
a solvent;

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- 10 (b) adding a reagent for changing the
oxidation state of the reactive moiety;
(c) sequentially adding additional reactive
building blocks in solvents to
synthesize the compounds; and
(d) cleaving the compounds from the solid
support within the apparatus.

69. A method for the multiple, simultaneous synthesis
of compounds using an apparatus according to
Claim 1 comprising the steps of:

- 5 (a) charging the apparatus with a solid
support with attached building block and
a solvent;
(b) adding a reagent for changing the
oxidation state of the reactive moiety;
(c) adding a coupling reagent in a solvent;
10 (d) sequentially adding additional reactive
building blocks and optionally coupling
reagents in solvents to synthesize the
compounds; and
(e) cleaving the compounds from the solid
15 support within the apparatus.

70. A method according to Claims 62, 63, 64, 65, 66,
67, 68, or 69 wherein the solid support is
selected from the group consisting of cross linked
divinylbenzene-styrene copolymer, controlled pore
5 glass, polyacrylamide, poly(ethylene glycol),
poly(ethylene glycol) monomethyl ether, silica
gel, cellulose, and acrylic acid grafted
polypropylene.

71. A method for the multiple, simultaneous synthesis
of an array of compounds according to Claim 61.

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72. A method for the multiple, simultaneous synthesis of organic compounds according to Claim 71.
73. A method according to Claim 72 for the synthesis of organic compounds selected from the group consisting of peptides, hydantoins, benzodiazepines, quinolones, keto-ureas, N2-substituted hydantoins, (R)-4-benzamido-5-oxopentanoic acid derivatives, diketopiperazines, pilocarpine compounds, tepoxalin derivatives, 2H-pyranones, arylpiperazines, benzisothiazolones, spirosuccinamides, benzopyrans, and pyrimidinediones.
74. A method according to Claim 73 for the synthesis of an array of peptides.
75. A method according to Claim 73 for the synthesis of an array of benzodiazepines.
76. A method according to Claim 73 for the synthesis of an array of hydantoins.
77. A method for the multiple, simultaneous synthesis of inorganic compounds according to Claim 71.
78. A method for the multiple, simultaneous synthesis of organometallic compounds according to Claim 71.
79. A method for preparing an array of compounds according to Claim 71 whereby the cleavage of each compound from the solid support is accomplished by utilizing a functionality contained within the compound to attack and rupture the bond attaching the compound to the solid support thereby

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simultaneously giving an unattached cyclic product.

5 80. A method for preparing an array of compounds according to Claim 71 whereby the cleavage of each compound from the solid support is accomplished by utilizing a functionalized compound to attack and rupture the bond attaching the penultimate compound to the solid support thereby simultaneously giving an unattached product incorporating the attacking compound as a portion of its structure.

5 81. A method for preparing an array of compounds according to Claim 71 whereby the cleavage from the solid support of each compound of the array is accomplished by utilizing a functionalized compound to attack and rupture the bond attaching the penultimate compound to the solid support thereby simultaneously giving an unattached product incorporating an invariant functionality at one position within the final compound.

5 82. A method according to Claim 71 of simultaneously synthesizing multiple organic compounds, selected from the group consisting of:
nitrogen containing heterocyclic compounds,
selected from the groups consisting of:
imidazopyridines, dihydropyridines, nucleoside and nucleoside analogs, indazoles, piperidines, benzazepines, indoles and condensed indoles, quinolines and isoquinolines, pyrrolidines,
10 aminopyrimidines, pyrrolizidines, guanidines, and tetrazoles;

oxygen containing heterocycles, selected from the groups consisting of: benzopyrans, coumarins,

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- 15 prostaglandins and prostacyclins, psoralens,
 tetrahydrofurans and pyrans;
 nitrogen and sulfur containing compounds,
 selected from the groups consisting of: beta-
 lactams and cephalosporins;
 carbocyclic compounds, selected from the
20 groups consisting of: tocopherol analogs,
 vitamin D analogs, steroids, naphthalenes, and
 anthracene analogs;
 alicyclic compounds, selected from the groups
 consisting of: polyunsaturated alkenes,
25 hydroxypropanolamines, benzofused bicyclic amines,
 and aryl amides;
 cyclic peptides; and cyclic nucleotides.

83. A method according to Claim 82 for synthesizing a
series of compounds simultaneously in an array
format based on a structure of known biological
activity for the purpose of developing a structure
5 activity relationship for a biological agent
 selected from the groups consisting of:
 muscarinic agonists, antiepileptics,
 antidepressants, HMG CoA reductase inhibitors,
 anti-infectives, cholecystokinin receptor
10 antagonist/agonists, antipsychotics,
 acylCoA:cholesterol O-acyltransferase inhibitors,
 and anti-inflammatory agents.
84. A method according to Claim 71 employing the
apparatus to perform reaction optimization.
85. A method according to Claim 71 employing the
apparatus for statistical experimental design
strategies.

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86. A method according to Claim 71 employing the apparatus for quantitative structure activity relationship strategies.
87. A method for the multiple, simultaneous synthesis of compounds using an apparatus according to Claim 1 comprising the steps of:
charging the apparatus with the reactants
5 required for synthesis; and
synthesizing the compounds in solution.
88. A method according to Claim 87 employing the apparatus to perform reaction optimization.
89. A method according to Claim 87 employing the apparatus for statistical experimental design strategies.
90. A method according to Claim 87 employing the apparatus for quantitative structure activity relationship strategies.

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FIG. 1

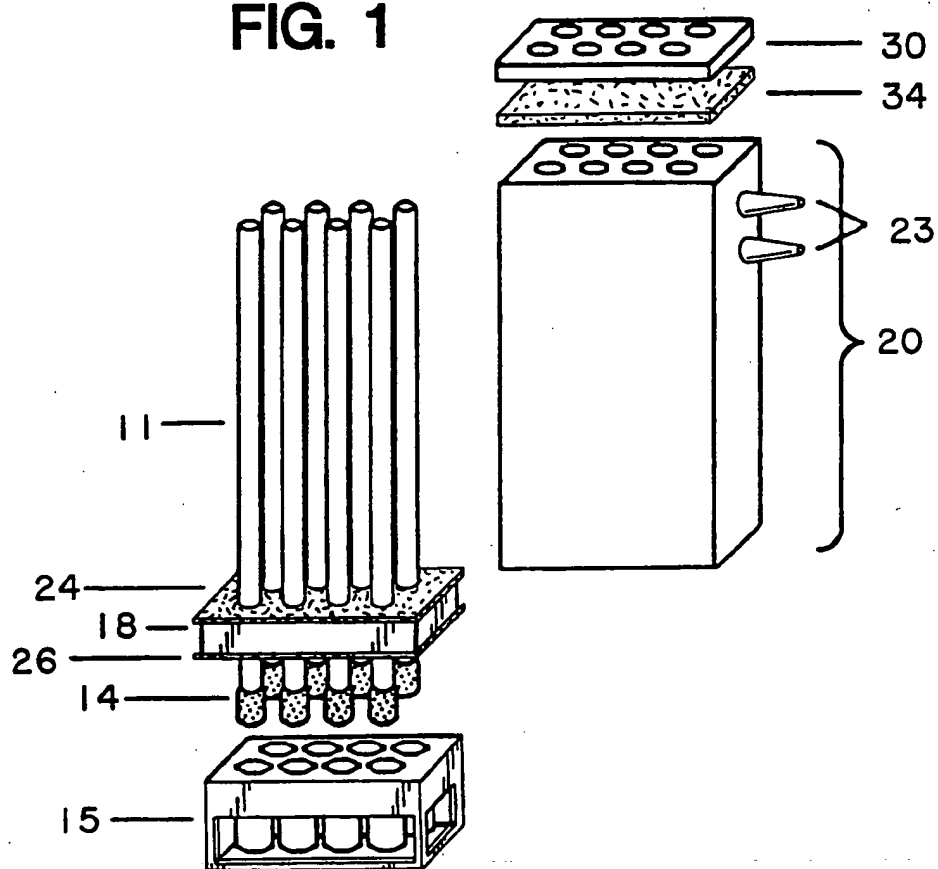


FIG. 3

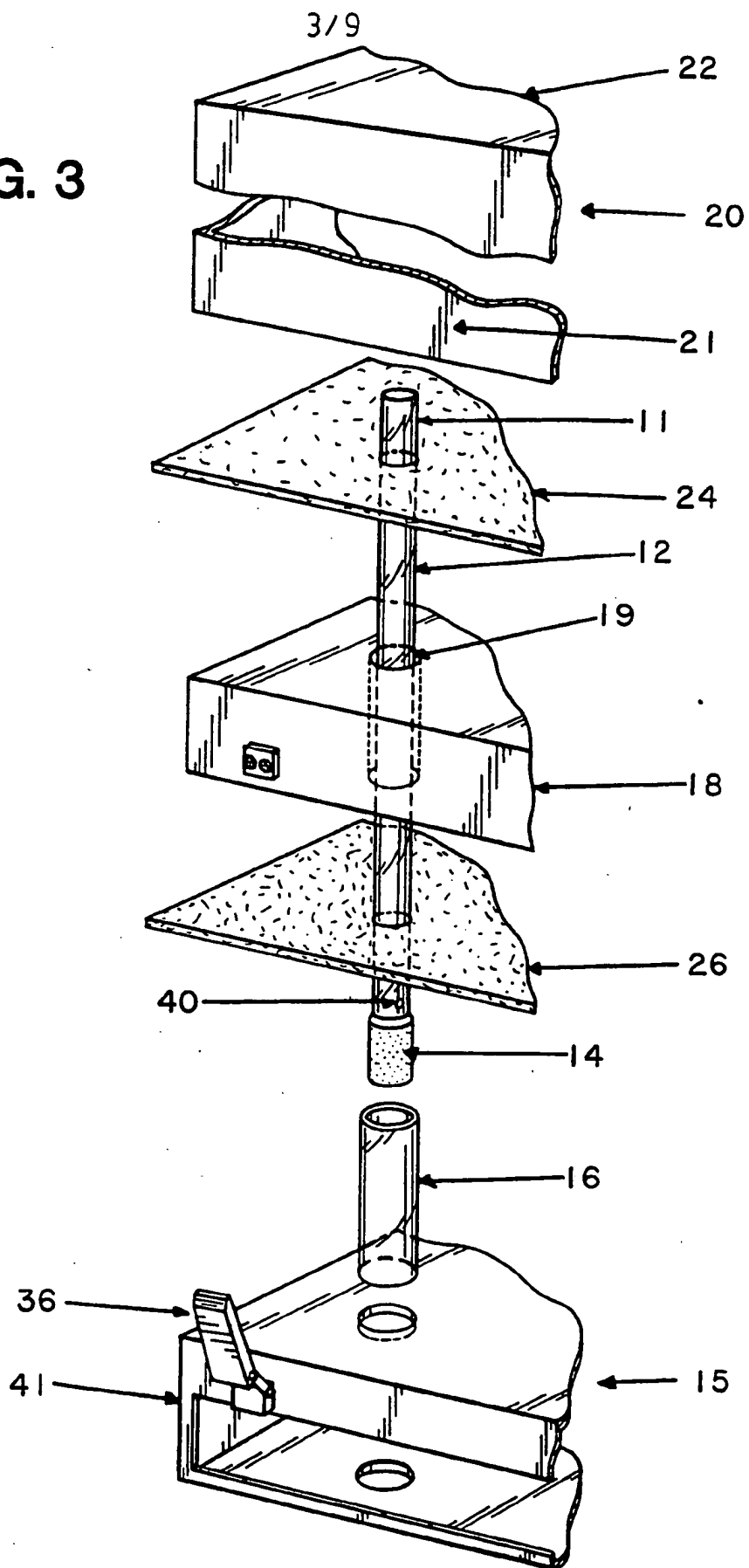
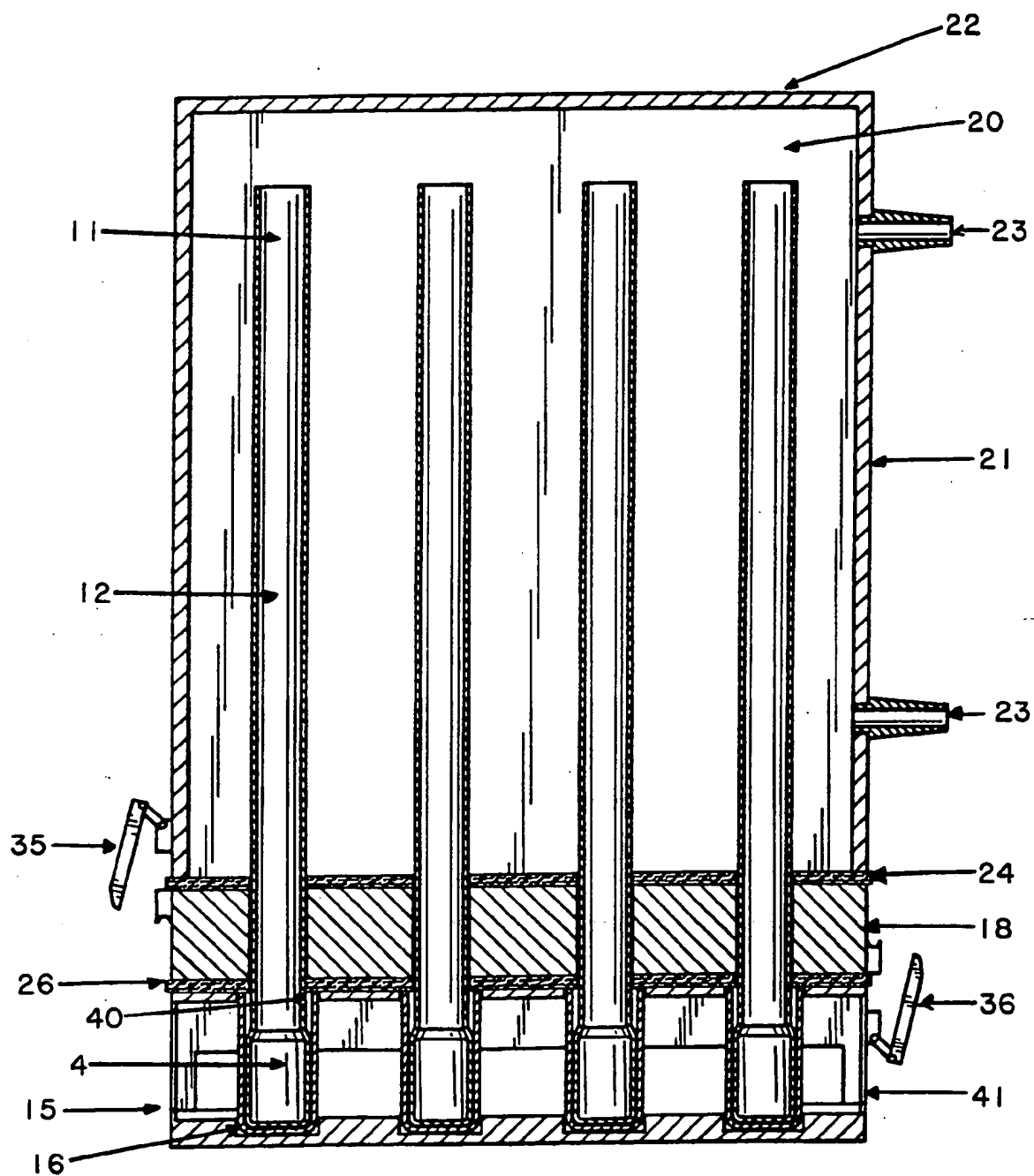


FIG. 4



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FIG. 5

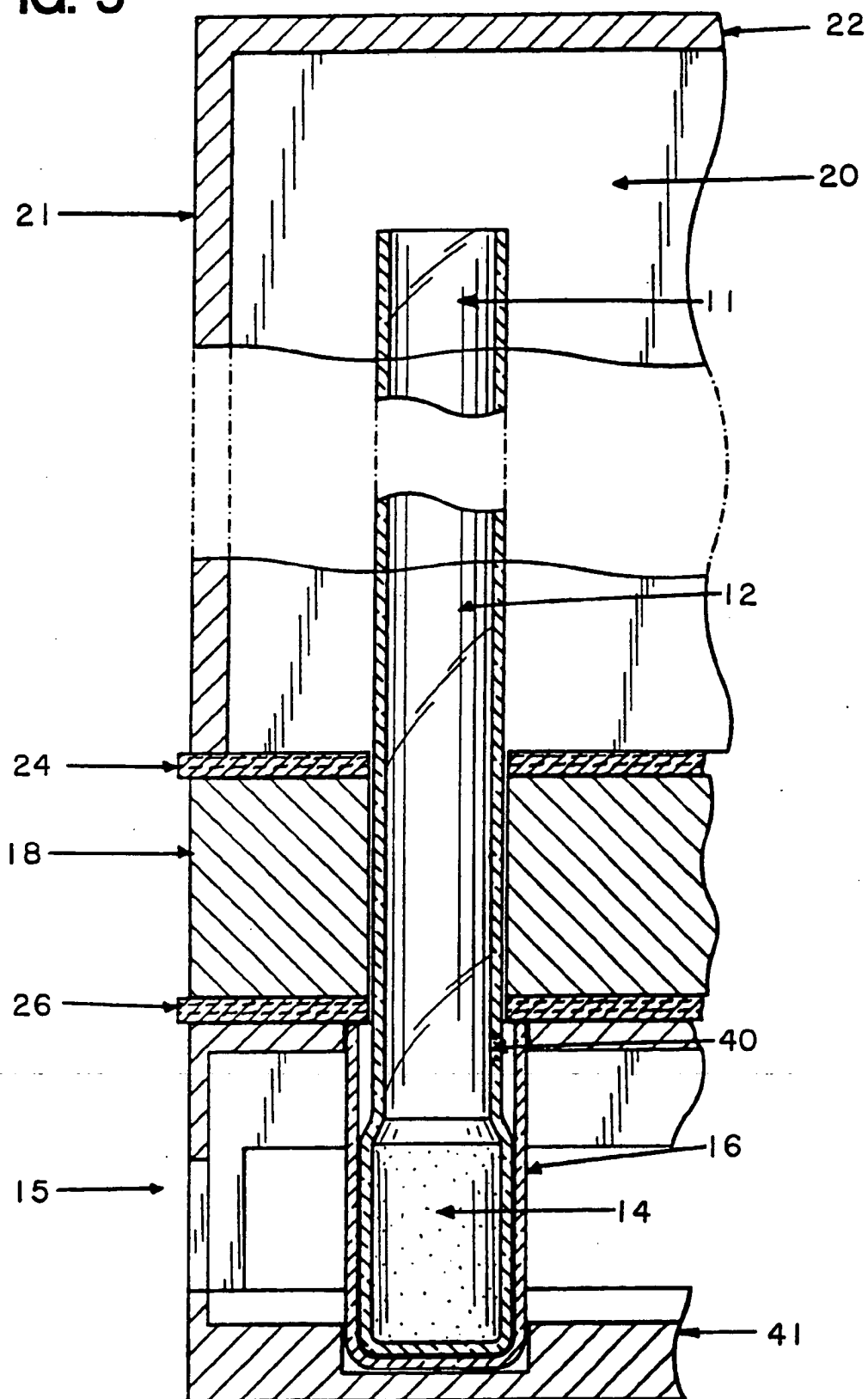
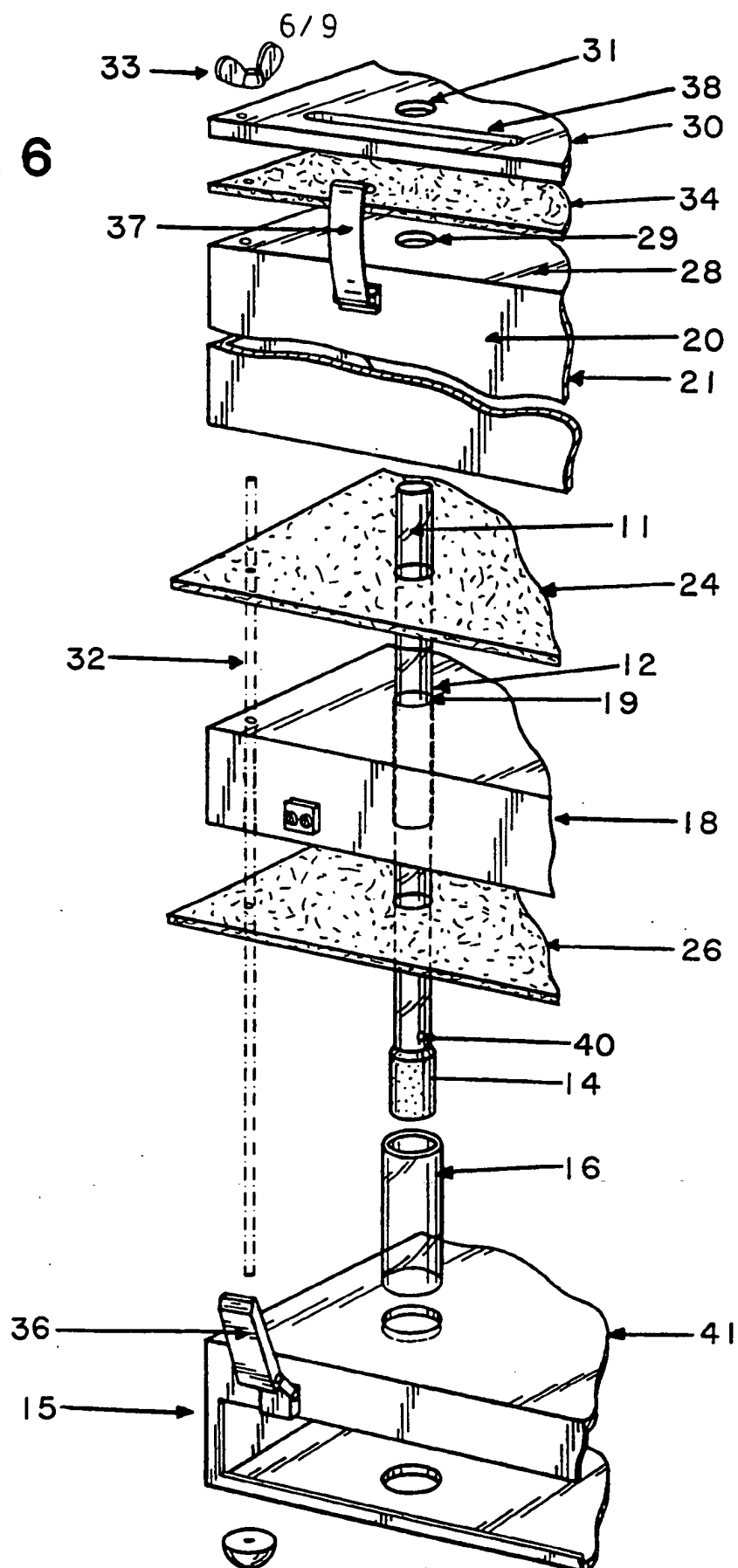


FIG. 6



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FIG. 7

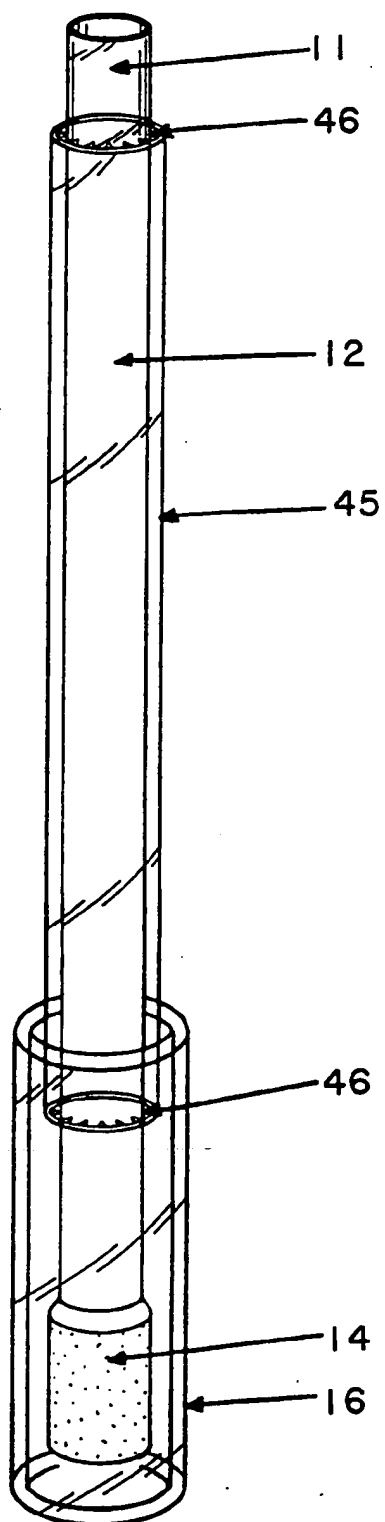


FIG. 8

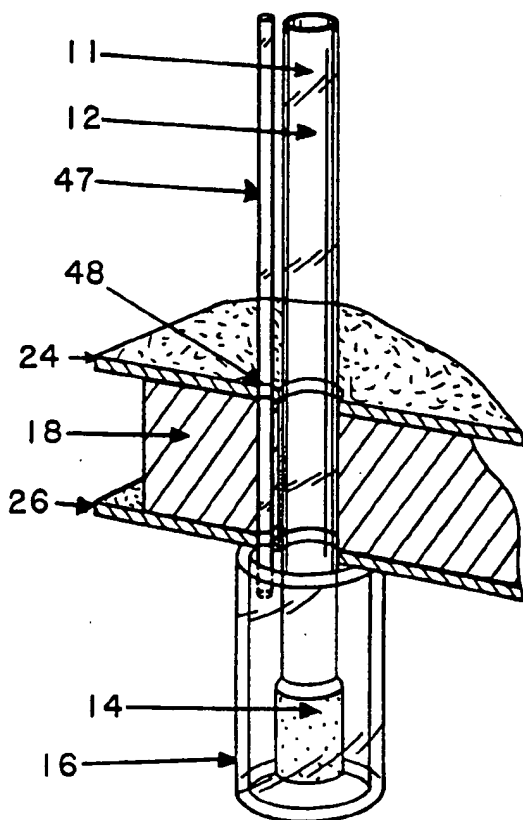
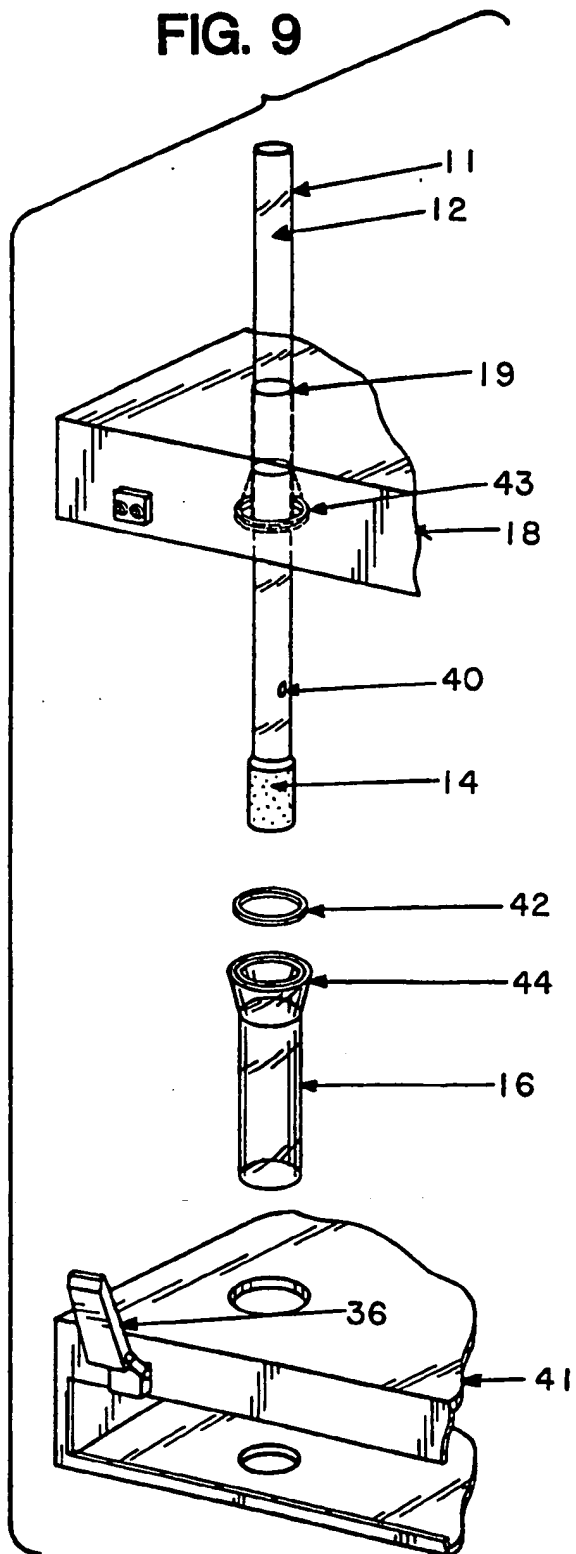
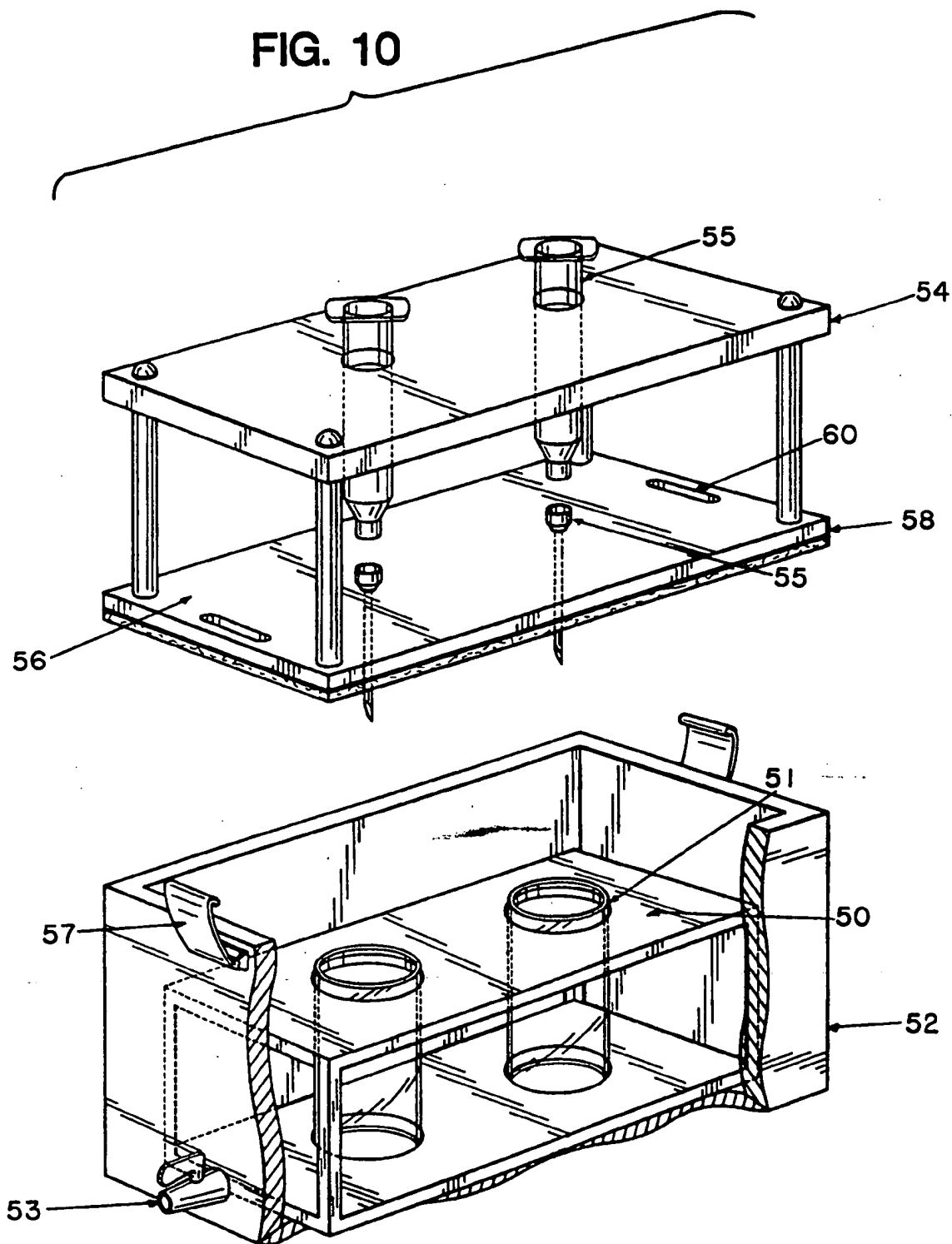


FIG. 9



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FIG. 10



INTERNATIONAL SEARCH REPORT

International Application No

PCT/US 93/09666

A. CLASSIFICATION OF SUBJECT MATTER
 IPC 5 B01J19/00 C07K1/04

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

IPC 5 B01J C07K

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	DE,A,37 23 004 (BOEHRINGER INGELHEIM KG) 26 January 1989	1,2, 9-11, 15-19, 21-30, 38,39, 61, 66-69, 71-74
Y	see the whole document	70,86 53,55, 62-65, 82,83
A	---	-/--

☒ Further documents are listed in the continuation of box C.

☒ Patent family members are listed in annex.

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Date of the actual completion of the international search

28 January 1994

Date of mailing of the international search report

22. 02. 94

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Stevnsborg, N

C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
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A	--- WO,A,92 10092 (AFFYMAX TECHNOLOGIES N.V.) 25 June 1992 see page 2, paragraph 1 - page 4, paragraph 1 see page 12, paragraph 3 - page 13, paragraph 1 see page 13, paragraph 3 - page 14, paragraph 1 see claims 1,3,4 -----	61, 71-74, 82,85

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Information on patent family members

International Application No

PCT/US 93/09666

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